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1	Evolutionary Importance of Intraspecific Variation in Sex Pheromones
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Highlights

Sex pheromones have traditionally been viewed as species recognition signals and
thus are expected to show little within-species variation in composition and amount.

Current research, however, reveals a high degree of intraspecific variation in sex
pheromone communication channels, suggesting they are molded by multiple
selection forces.

• Sex pheromone signaling can be costly and plastic, and signals as well as responses may be affected not only by between-species interactions (e.g. communication interference, host plants, geographic variation), but also by within-species interactions (mate choice) in addition to the genetic architecture, physiological state, and previous experience of individuals.

As variation in sex pheromone communication can be induced by intrinsic factors, its
evolution may not only follow ecological speciation, but also be the driver of
divergence.

Abstract

As sex pheromones in many insect species are important species-recognition signals that attract conspecifics and inhibit attraction between heterospecifics; sex pheromones have predominantly been considered to evolve due to interactions between species. Recent research, however, is uncovering roles for these signals in mate choice, and that variation within and between populations can be drivers of species evolution. Variation in pheromone communication channels arises from a combination of context-dependent, condition-dependent, or genetic mechanisms in both signalers and receivers. Variation can affect mate choice and thus gene flow between individuals and populations, affecting species' evolution. The complex interactions between intraspecific and interspecific selection forces calls for more integrative studies to understand the evolution of sex pheromone communication.

Sex Pheromones and Their Functions

Pheromones are semiochemicals involved in intraspecific communication, where species-specific signals are released by a sender that modify the behavior of a receiver [1]. Sex pheromones signal attraction and selection of potential mates. Sex pheromones are used by a variety of animal species, but the focus of this Review is on insect sex pheromones, as most pheromone research has been conducted on this class of animals. The traditional view is that sex pheromones are important species recognition signals to distinguish between species and thus under **stabilizing selection** (see Glossary) [2]. As closely related species can have similar sex pheromone signals that may only differ in the ratio of the different chemical constituents, the signaler and the receiver need to be finely tuned to recognize each other [3]. Small changes in pheromone release rate or ratio of the chemical constituents can affect attraction of the receiver [4].

Since sex pheromones are also mate recognition signals acting within species, their roles in reproductive isolation and speciation processes are important. Reproductive isolation can be shaped by **reproductive character displacement** [5]. In sex pheromone communication channels, reproductive character displacement has been found in closely related, sympatrically occurring taxa [6,7], which lessens communication interference between closely related and **sympatric species**, but can generate variation between populations that lead to divergence [5]. Intraspecific variation between geographically distant populations has been described in many species [8-10], suggesting that such communication interference and other environmental factors affect variation in pheromone communication.

Even though intraspecific variation between geographically isolated populations is generally accepted, variation within populations is still thought to be less common. Johansson & Jones [11], however, reviewed the role of sex pheromones in mate choice, showing the possibility of variation in these signals even within populations. We now know that sex pheromones can even be plastic, and this **plasticity** can be translated into quantitative [12,13] or qualitative [14] intrapopulational differences. This can take place over an individual's lifespan [12,15], as a consequence of seasonal development [13], or due to the vicinity of heterospecifics during development [14]. Since sex pheromones can provide information on the general condition of the signaler, and the receiver can select based on this variation, intraspecific variation in sex pheromones must be more common than previously assumed.

Why Has Intraspecific Variation Been Overlooked?

Although chemical communication is considered to be the oldest form of communication [16], humans are mostly visually oriented and thus much of the research focused on sexual signaling has been on variation in visual signaling. Also, it has proven challenging to analyze

individual-level variation in chemical extracts. For example, identification of the first moth sex pheromone required thousands of individuals pooled together [17], naturally obscuring any intrapopulation variation. In addition, research on sexual selection and sexual conflict theory is affected by biases of sex roles and assignment to different taxa in sexual selection research [18]. In moths, females are typically thought to be the signalers and release the sex pheromone, while males are the responding sex, which could explain the higher volume of literature on female sex pheromone variation in moths [19-22]. Males, however, also produce and release sex pheromone [23] to which females may respond, but this aspect has been little explored (Box I). Finally, female-signaling insect systems are well-represented amongst pest species, making them economically important. Sex pheromones are commonly used to monitor the presence and abundance of these pests and to disrupt mating. These applied aspects of insect chemical ecology likely increased the focus on sex pheromones at the species level. Thus, it is possible that, to date, our understanding of sex pheromone evolution, and in particular how intraspecific variation can affect a single species, has been held back by our research focus.

Mechanisms Underlying Intraspecific Sex Pheromone Variation

Both sender and receiver are involved in the process of mate attraction, and intraspecific variation in pheromone communication can either arise as a consequence of physiological variation in the signaler (sender-specific driver of variation) or the receiver (receiver-specific driver of variation). To add another layer of complexity, both the sender and the receiver experience physiological changes within their lifetime, adding within-individual variation to the between-individual variation at the population level. In addition, senders may become receivers and vice versa.

Sender's Perspective

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Diet, age, mating status, and vicinity to conspecifics are among the mechanisms that can produce physiological changes and contribute to maintenance of intraspecific variation in pheromone production. These changes can lead to both within- and between-individual variation. When considering intraspecific sex pheromone variation, it is important to first consider how they are produced. In some species, pheromone components and precursors are sequestered from the diet, while in others they are produced *de novo* within the animal [24]. Although both forms of pheromone production rely on the resources available, pheromones derived from the diet will be more vulnerable to environmental and host-related effects, and thus to external conditions. In contrast, species that produce their pheromones de novo are probably less susceptible to externally-induced variation, but may exhibit variation due to internal processes. This is especially likely if there are costs to pheromone production, as this could result in trade-offs with other processes. Sex pheromones can indeed be costly: nutritional state influences the amount of pheromone produced in cockroaches [12], smaller female moths have lower fitness when they are stimulated into pheromone signaling [25], and pheromone composition predicts the fitness of female moths [26]. The signaling sex may communicate its nutritional state to the receiver [27]. There is also ample evidence that sex pheromones are affected by suboptimal diet at the larval stage [15,28], in adults [12], or at both life stages [15]. Pheromone production can be recouped if a higher nutritional diet is obtained [12], resulting in a dynamic environment-individual interaction. Regardless of how sex pheromones are produced, their variation can also be a consequence of the internal physiological state and its changes, such as mating status [29,30] or aging [31,32]. Generally, a greater investment in pheromone signaling is expected in older unmated females [33]. Older females may also call earlier in the night to avoid competition with more attractive young females [34].

Other mechanisms that can affect pheromone production and release are pathogens [19], volatiles from host plants [31], conspecifics [35,36] or prey [37]. Verheggen et al. [37] recently found that pheromone production in the Asian lady beetles (*Harmonia axyridis*) (Figure 1A) is conditioned by the presence of prey, as exposure to volatiles of the prey initiated pheromone production. Also, the female calling behavior of the gregarious beet webworm moth (*Loxostege sticticalis*) is affected by the vicinity to male conspecifics [38]. Pheromone release can also be increased by crowding and sexual competition, for example in the American grasshopper (*Schistocerca americana*) [35]. It remains unclear if any of these changes represent adaptive strategies, i.e. changes in resource allocation depending on individual status, or are simply non-adaptive responses to environmental triggers. Examples that do suggest a level of adaptive response to the environment are developmental plasticity due to the chemical environment, when individuals have been exposed to the chemical signals of heterospecifics [14], as a consequence of environmental temperature [39], seasonal variation [13], or nocturnal light pollution [40].

Finally, genetic and behavioral mechanisms may contribute to within-population pheromone variation. For example, a point mutation identified in the tobacco budworm (*Heliothis virescens*) (Figure 1B) translates to a sex pheromone signal with a higher or lower ratio of two pheromone components [41]. The different ratios of the two components affect the signal's attractiveness to males, and females releasing a signal with a higher ratio of the two components are less attractive than females releasing pheromone signal with a lower ratio [36]. Unattractive females can obtain matings while in close proximity to attractive females and, therefore, unattractive females are maintained in the population [36]. Similarly, in the European corn borer (*Ostrinia nubilalis*), two genetically identified sex pheromone strains

[42] are maintained because males assortatively mate with females of their own pheromone strain [43].

Receiver's Perspective

The traditional view holds that signalers and receivers are finely tuned, where the predicted shape of the receiver's **preference function** corresponds to the distribution of the female pheromone signal in the population. Experimental evidence from the 1970-80's suggested that female pheromone signals experience stabilizing selection [3] and there is a lack of variation in male responses, because of optimal tuning to specific component ratios [2,4]. Variation in either the signaler or receiver, however, can introduce novelty in the communication channel even in established sender-receiver systems. For example, the rare receivers in the population that track changes in the pheromone signal [44], which might be possible through a **saltational event** [45]. New compounds in the signal may remain unperceived initially and only later do receivers evolve the preference for the new signal [46]. This scenario is known as the asymmetric tracking hypothesis [47]. An alternative and new conceptual model proposes that the preference trait evolves first in the receiver as a veiled preference, before the preferred trait is evolved in the signaler, and the receiver starts to select individuals with the newly preferred trait as soon as the trait arises [48].

Variation in receivers can take place at different levels, such as between- and within-individuals, and at different life stages, as a consequence of plasticity in the olfactory system [49]. In some species, population density can contribute to variation in the morphological structures of the receiver as a consequence of developmental plasticity. For example, gumleaf skeletonizer male moths (*Uraba lugens*) develop longer and more elaborate antennae to locate females in sparse populations [50]. This plasticity depends on the population density

during the developmental stage and suggests that individuals can be more or less sensitive to pheromone signals.

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Within the lifespan of an individual, plasticity in behavioral responses is well demonstrated in relation to circadian rhythms, as males of many moth species actively respond to pheromone sources at restricted times in the photoperiod [51]. A difference in gene expression in odor receptors seems to be at the base of this behavioral plasticity, linked to both the physiological state [52] and circadian rhythms [53] with, for example, effects on antennal sensitivity [54]. Responses to sex pheromones can also be modulated by experience, where pre-exposure to sex pheromones increases long-term sensibility and responsiveness [55,56]. The olfactory system dynamically adjusts to optimally perceive the surroundings [57] and differential receptor expression characterizes, for example, mated and unmated individuals [58]. After matings, some insects become less sensitive to sexual signals and cues for mating sites [59] either through a reduction in antenna sensitivity [54] or differences in pheromone processing in the central nervous system in the brain [60]. In other instances, environmental stress can have an effect on the receiver olfactory system. Sublethal pesticide doses was shown to increase the peripheral sensitivity of cotton leafworm (Spodoptera littoralis) males and increase their mating success [61]. A recent review [49] reports that the plasticity of the olfactory system occurs at all levels of the olfactory pathway. When studying the insect sensory system, it is challenging to integrate the information from the periphery to the brain, therefore, we have just begun to understand these interactions. As the olfactory system can plasticly respond to, for example, changes in the environment, physiological state, social interactions and experience, variation in the olfactory system among individuals is likely to lead to differences in sensitivity and responses.

Ecological Consequences of Intraspecific Variation: Effect on Attractiveness and **Mating Success** Intraspecific variation in sex pheromones can have important consequences in attractiveness, mating success and mating behavior, even when the source of the variation is entirely environmental. Furthermore, these consequences can be sex-specific. Volatiles produced by host plants can affect the production and response to sex pheromones. This can in turn affect mate location by increasing the calling behavior of signalers, for example by augmenting the frequency and calling duration [62], or increasing pheromone release/production [63]. Alternatively, host plant volatiles can enhance receivers' attraction to pheromone signals [62,64], by attracting a higher number of individuals [62,64] or by reducing the time to locate the pheromone source [64]. Finally, host plant volatiles can alter the response to ratios of pheromone components [65]. Thus, in general, host plants can influence species reproductive success during different stages of the insect lifespan, through both resource acquisition at the larval stage, and the interaction of plant volatiles with adult

the response to ratios of pheromone components [65]. Thus, in general, host plant volatiles can alter the response to ratios of pheromone components [65]. Thus, in general, host plants can influence species reproductive success during different stages of the insect lifespan, through both resource acquisition at the larval stage, and the interaction of plant volatiles with adult signals and behavior. Oxidative stress and pathogen infections can also affect sexpheromone based mate location [66,67]. Curiously, challenged immune systems and pathogen infections have been found to increase rather than decrease attraction in some insects, suggesting either terminal investment [67] or a strategy of pathogens to spread through populations via host manipulation [19]. Immune system challenges can also lead to sex-specific differences, for example only female mating success might be affected [68]. As populations vary in parasite load [69], variation in immune challenge could thus impact the dynamics of both pheromone signaling and sexual selection. In addition to host plants and pathogens, sex pheromone production and mating success can be affected by seasonal fluctuations. A good example is the squinting bush brown butterflies (*Bicyclus anynana*), in

which males that emerge in the wet season produce more pheromone and have higher mating success than those that emerge in the dry season [13].

Mate assessment is based on intraspecific variation in sexually selected traits. Sex pheromones can be under sexual selection when used as indicators of mate quality. Variation in sex pheromone composition can be used to avoid mating with siblings [70], which increases genetic variability and decreases the impact of deleterious alleles. For example, in *B. anynana*, females avoid mating with inbred males, which is assessed solely on the male-produced sex pheromone [71]. Sex pheromones are also used as signals to avoid matings that would lead to a lowered fitness and thus reduce the cost of matings. For example, sexually immature females of the cotton bollworm (*Helicoverpa armigera*) inhibit male attraction by temporarily releasing a repellent component in their pheromone signal [72]. Another example is the haplodiploid parasitic wasp, *Nasonia vitripennis*, in which females are more attracted to males with more pheromone, which correlates to a full sperm load [73]. As unfertilized eggs develop into males, this selection thus affects the population sex-ratio.

Sexual conflicts often arise over mating frequency. For example, in the parasitoid wasp families Pteromalidae and Braconidae, females tend to mate only once [30,74], while males mate multiple times. After mating, *Spalangia endius* (Pteromalidae) females actively resist males by increasing the release of a specific pheromone component, which prevents or terminates male courtship behavior [74]. Mated *Cotesia glomerata* (Braconidae) females stop releasing their attractive pheromone component in favor of the repellent only. These changes likely help mated females to avoid male harassment by reducing the attraction of males, and are also thought to help males locate virgin females, as their pheromone is distinct from mated females [30].

To conclude, intraspecific sex pheromone variation can have important consequences for the attractiveness and mating success of individuals, regardless of the source of this variation. If variation in signals and responses is heritable, there is the potential for different investment or responsive strategies to evolve.

Evolutionary Consequences of Intraspecific Sex Pheromone Variation

Evolution in sexual communication systems can occur when heritable differences in signals and/or responses increase the survival and reproductive output of the individual (Figure 2 & Table 1). Below we explore whether and how such evolution can occur in response to **ecological speciation** or as a driver of population divergence.

Ecological Speciation

Many herbivore species use chemicals from their host plants as precursors for compounds that make up the pheromone signal. Thus, host plant shifts can directly affect pheromone composition. If this is followed by changes in responses, leading to **assortative mating** and reproductive isolation, ecological speciation can occur [75]. Changes in mating signals following a host plant shift has been experimentally shown in the mustard beetle (*Phaedon cochleariae*) [76], where **cuticular hydrocarbons** used as mate recognition cues differ depending on the host plant on which they feed. Similar changes are hypothesized to have contributed to the speciation process between two parental species of flea beetles (*Altica fragaria* and *A. viridicyanea*) [77]. Pheromone divergence between populations linked to differences in host plants has also been described in the pine and larch strain of larch budmoth (*Zeiraphera diniana*) [78] and the chestnut tortrix (*Cydia splendana*) [79], and may represent the initial step towards reproductive isolation in these species. Changes in host

plants, however, do not automatically lead to differences in sex pheromones [80] or to host shift [81], suggesting that other processes are also involved in speciation events.

Plants damaged by conspecific herbivores can negatively affect the premating and mating behavior in the moth species *S. littoralis* [82]. Host plant volatiles released as a result of herbivory can decrease the calling behavior of the signaler and negatively impact the mating success. This interaction may result in an avoidance of egg laying on a suboptimal plant that has high competition for resources or is attractive to the enemies of the herbivore [82]. Thus, interactions between host plants and pheromone signals and/or responses are complex and not straightforward, and how these interactions are involved in ecological speciation likely depends on the plants and insects involved.

Pheromone Differentiation as Driver of Speciation

Reproductive isolation can also take place without ecological differentiation, through divergence in mate preference [83]. For example, in the primitive moth *Eriocrania semipurpurella-sangii* species complex, the ratios of components in the female pheromone do not only reflect geographic differences between populations, but also identify different species [84]. Since all the species share the same host plant, this is a case in which reproductive isolation likely has evolved as a result of divergence in sexual communication [84]. Such divergence could happen if some individuals of the choosing sex have a preference for a specific pheromone signal and choose those signalers. A good example comes from *B. anynana*, in which females developed a mating bias towards the pheromone signal they were exposed to, even when this was a new signal. As this mating bias can be transmitted to the next generation without the offspring being exposed to the new pheromone signal [85], such heritable learning can drive the evolution of assortative mating and

speciation. The potential of sex pheromones as drivers of speciation has been hypothesized for some *B. anynana* populations in which sex pheromone differentiation was found to be higher than genetic differentiation, suggesting that pheromone divergence precedes genetic divergence [8] (Box II).

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Potential mechanisms that can introduce variation and novelty in sex pheromones lie in the sequence variability of the genes involved in determining component ratios in pheromone signals [86], such as gene duplication and amino acid substitution in genes coding for specific enzymes in the pheromone pathways [87]. Variation in response and preference for pheromone signals may be due to divergence and evolution in olfactory receptors [88]. For example, chemosensory divergence in odorant receptors in the peripheral nervous system associated with reproductive isolation, have been found in two rarely hybridizing Heliconius species [58]. In O. nubilalis, however, genetic differences in the central nervous system (i.e. neurogenesis), instead of the odorant receptors, were recently found to be primarily responsible for the differential male response [89]. The discovery of the genetic architecture underlying variation in sexual signals and responses has just recently begun, and so far mostly focused on Lepidoptera [90]. Recent studies suggest that phenotypic variation in sex pheromones can be the result of single gene [42] or locus [91] variation or due to a number of loci [92]. In the moths species studied so far, the genomic regions involved in signaling and response are unlinked. This makes runaway selection unlikely and the lack of genetic **correlation** favors the evolution of complex traits [90]. Thus, how phenotypic variation in pheromone communication may lead to reproductive isolation and, ultimately, to speciation is still an evolutionary mystery.

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How Easily Can Pheromone Communication Channels Evolve?

As insect pheromones are used for pest management tactics, we have some knowledge on the evolution of sex pheromones in response to anthropogenically induced selection, which shows that pheromone communication channels can evolve relatively rapidly. Techniques such as pheromone trapping and **mating disruption** are attractive alternatives to pesticides, but the continuous use of artificial sex pheromones as disruptants may exert selective pressure on pest species to change their sexual signals and evolve 'resistance' to it [93], meaning that males are no longer disrupted by the artificial pheromone composition. Cases of such emerging resistance can be viewed as natural field experiments showing that sexual communication systems can sometimes evolve quickly. The first field case of male resistance to pheromone-based traps was reported in Japan, where males of the smaller tea tortrix (Adoxophyes honmai) stopped responding to synthetic lures after about 10 years of treatment, causing the efficacy of mating disruption to drop from > 96% to < 50% [94]. The strong selective pressure exerted on this species resulted in the evolution of a resistant population, in which females changed their pheromone composition and males broadened their pheromone response [95,96]. This is unlikely to be an isolated case, as intraspecific variation observed in the cosmopolitan pest, codling moth, Cydia pomonella, shows the same potential for a shift in female sexual signal in response to mating disruption [20].

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The continuous use of artificial pheromones can have an effect on both signalers and receivers. The signaling sex may evolve a different pheromone signal [95,97] and/or modify their behavior, by releasing pheromone for a longer time [98]. Pre-exposure to sex pheromones can affect gene expression of chemosensory genes and olfactory sensitivity [53] and result either in enhanced [55,56] or reduced [99] receiver sensitivity. All these factors can contribute to the evolution of resistance to artificial pheromones, and suggest that signaler and receiver can evolve and change in short time periods. Such quick evolutionary responses

to selection indicate that there is a high level of standing genetic variation in populations on which selection can act.

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Concluding Remarks and Future Perspective

There is an increasing awareness that intraspecific variation in sex pheromone communication channels, both in the signaler and the receiver, is more common than previously assumed. There is also an increasing number of studies that point to the existence of within-population variation, as these signals are used for mate assessment and choice. Thus, sex pheromone signals and responses can be under multiple selection forces (see Outstanding Questions) and seem able to evolve relatively quickly. This is particularly well represented by the fact that wild populations have been found to evolve resistance to synthetic sex pheromone composition, which suggests a high level of standing genetic variation within species on which selection can act. Finally, it is important to realize that individuals in nature can be affected simultaneously by several external factors (e.g. pathogens, vicinity to conspecifics, predators), which can trade-off with the physiology of the individual, so that populations always show some degree of variation. We therefore need to deepen our understanding on how variation in pheromone communication channels affects and is affected by the interaction between the internal state of signalers and receivers, their mutual mate choice and their environment. We also need a better understanding of the mechanisms behind this variation, whether these are genetic or plastic, and which molecular pathways are involved. With all this information it will be possible to move the field forward and better understand how the evolution of chemical communication between the sexes affects speciation.

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References

- 1. Karlson, P. and Lüscher, M. (1959) 'Pheromones': a new term for a class of biologically active substances. *Nature*, 183, 55-56
 - 2. Linn, C.E. and Roelofs, W.L. (1985) Response specificity of male pink bollworm moths to different blends and dosages of sex pheromone. *J. Chem. Ecol.* 11, 1583-1590
 - 3. Linn Jr, C.E. and Roelofs, W.L. (1983) Effect of varying proportions of the alcohol component on sex pheromone blend discrimination in male oriental fruit moths. *Physiol. Entomol.* 8, 291-306
 - 4. Baker, T.C. et al. (1981) Sex pheromone dosage and blend specificity of response by oriental fruit moth males. *Entomol. Exp. Appl.* 30, 269-279
 - 5. Pfennig, K. and Pfennig, D. (2009) Character displacement: ecological and reproductive responses to a common evolutionary problem. *Q. Rev. Biol.* 84, 253-276
 - 6. Yang, C.Y. et al. (2015) Sex pheromones and reproductive isolation in five mirid species. *PLoS One* 10, e0127051
 - 7. Meier, L.R. et al. (2016) Synergism between enantiomers creates species-specific pheromone blends and minimizes cross-attraction for two species of cerambycid beetles. *J. Chem. Ecol.* 42, 1181-1192
 - 8. Bacquet, P.M. et al. (2016) Differentiation in putative male sex pheromone components across and within populations of the African butterfly *Bicyclus anynana* as a potential driver of reproductive isolation. *Ecol. Evol.* 6, 6064-6084
 - 9. Darragh, K. et al. (2020) Species specificity and intraspecific variation in the chemical profiles of *Heliconius* butterflies across a large geographic range. *Ecol. Evol.* 10, 3895-3918
 - 10. Gao, K. et al. (2020) Geographic variation in sexual communication in the cotton bollworm, *Helicoverpa armigera*. *Pest Manag. Sci.* 76, 3596-3605
 - 11. Johansson, B.G. and Jones, T.M. (2007) The role of chemical communication in mate choice. *Biol. Rev.* 82, 265-289
- 12. Jensen, K. et al. (2017) Change in sex pheromone expression by nutritional shift in male cockroaches. *Behav. Ecol.* 28, 1393-1401
- 13. Balmer, A.J. et al. (2018) Developmental plasticity for male secondary sexual traits in a group of polyphenic tropical butterflies. *Oikos* 127, 1812-1821
- 438 14. Groot, A.T. et al. (2010) Phenotypic plasticity in sexual communication signal of a noctuid moth. *J. Evol. Biol.* 23, 2731-2738

440 15. Schultzhaus, J.N. et al. (2018) High fat diet alters *Drosophila melanogaster* sexual
 441 behavior and traits: decreased attractiveness and changes in pheromone profiles. *Sci.* 442 *Rep.* 8, 1-13

- 443 16. Wyatt (2014) Animals in a chemical world. In *Pheromones and Animal Behavior:* 444 *Chemical Signals and Signatures* (2nd edition), pp. 1-48, Cambridge University Press
 - 17. Butenandt, V.A. (1959) Uber den sexsual-lockstoff des seidenspinners *Bombyx mori. Reindarstellung und konstitution. Z. Naturforschg, b*, 14, 283
 - 18. Green, K.K. and Madjidian, J.A. (2011) Active males, reactive females: stereotypic sex roles in sexual conflict research?. *Anim. Behav.* 81, 901-907
 - 19. Burand, J.P. et al. (2005) Infection with the insect virus Hz-2v alters mating behavior and pheromone production in female *Helicoverpa zea* moths. *J. Insect Sci.* 5, 6
 - 20. Duménil, C. et al. (2014) Intraspecific variation in female sex pheromone of the codling moth *Cydia pomonella*. *Insects* 5, 705-721
 - 21. Groot, A.T. et al. (2014) Within-population variability in a moth sex pheromone blend: genetic basis and behavioural consequences. *Proc. R. Soc. Lond. B* 281, 20133054
 - 22. Cruz-Esteban, S. et al. (2018) Geographic variation in pheromone component ratio and antennal responses, but not in attraction, to sex pheromones among fall armyworm populations infesting corn in Mexico. *J. Pest Sci.* 91, 973-983
 - 23. Birch, M.C. et al. (1990) Scents and eversible scent structures of male moths. *Annu. Rev. Entomol.* 35, 25-54
 - 24. Foster, S.P. and Anderson, K.G. (2015) Sex pheromones in mate assessment: analysis of nutrient cost of sex pheromone production by females of the moth *Heliothis virescens*. *J. Exp. Biol.* 218, 1252-1258
 - 25. Harari, A.R. et al. (2011) Fitness cost of pheromone production in signaling female moths. *Evolution* 65, 1572-1582
 - 26. Blankers, T. et al. (2021) Sex pheromone signal and stability covary with fitness. *R. Soc. Open Sci.* In press
 - 27. Foster, S.P. and Johnson, C.P. (2011) Signal honesty through differential quantity in the female-produced sex pheromone of the moth *Heliothis virescens*. *J. Chem. Ecol.* 37, 717-723
 - 28. Darragh, K et al. (2019) Male pheromone composition depends on larval but not adult diet in *Heliconius melpomene*. *Ecol. Entomol.* 44, 397-405
 - 29. Lu, Q. et al. (2017) Sex pheromone titre in the glands of *Spodoptera litura* females: circadian rhythm and the effects of age and mating. *Physiol. Entomol.* 42, 156-162
 - 30. Xu, H. et al. (2019) The combined use of an attractive and a repellent sex pheromonal component by a gregarious parasitoid. *J. Chem. Ecol.* 45, 559-569
 - 31. Domínguez, A. et al. (2019) Influence of age, host plant and mating status in pheromone production and new insights on perception plasticity in *Tuta Absoluta*. *Insects* 10, 256
 - 32. Kartika, T. et al. (2021) Influence of Age and Mating Status on Pheromone Production in a Powderpost Beetle *Lyctus africanus* (Coleoptera: Lyctinae). *Insects* 12, 8
 - 33. Umbers, K.D. et al. (2015) The mothematics of female pheromone signaling: strategies for aging virgins. *Am. Nat.* 185, 417-432
- 34. Pham, H.T. et al. (2021) Age-dependent chemical signalling and its consequences for mate attraction in the gumleaf skeletonizer moth, *Uraba lugens*. *Anim Behav*. 173, 207-213

488 35. Stahr, C. et al. (2013) Chemical identification, emission pattern and function of male-489 specific pheromones released by a rarely swarming locust, *Schistocerca americana*. *J. Chem. Ecol.* 39, 15-27

- 36. van Wijk, M. et al. (2017) Proximity of signallers can maintain sexual signal variation under stabilizing selection. *Sci. Rep.* 7, 1-10
 - 37. Verheggen, F. et al. (2020) The Production of Sex Pheromone in Lady Beetles Is Conditioned by Presence of Aphids and Not by Mating Status. *J. Chem. Ecol.* 46, 590-596
 - 38. Xiao, Y. et al. (2020) Male Ventroposterior Brush Display Increases the Sexual Receptivity of Females in the Gregarious Beet Webworm *Loxostege sticticalis* (Lepidoptera: Crambidae). *J. Insect Behav.* 33, 184-192
 - 39. Dion, E. et al. (2016) Phenotypic plasticity in sex pheromone production in *Bicyclus anynana* butterflies. *Sci. Rep.* 6, 1-13
 - 40. Van Geffen, K.G. et al. (2015) Artificial night lighting disrupts sex pheromone in a noctuid moth. *Ecol. Entomol.* 40, 401-408
 - 41. Groot, A.T. et al. (2019) Within-population variability in a moth sex pheromone blend, part 2: selection towards fixation. *R. Soc. Open Sci.* 6, 182050
 - 42. Lassance, J.M. et al. (2010) Allelic variation in a fatty-acyl reductase gene causes divergence in moth sex pheromones. *Nature* 466, 486-489
 - 43. Malausa, T. et al. (2005) Assortative mating in sympatric host races of the European corn borer. *Science* 308, 258-260
 - 44. Baker, T.C. (2002) Mechanism for saltational shifts in pheromone communication systems. *Proc. Natl. Acad. Sci. U. S. A.* 99, 13368-13370
 - 45. Shimomura, K. et al. (2016) Saltational evolution of contact sex pheromone compounds of *Callosobruchus rhodesianus* (Pic). *Chemoecology* 26, 15-23
 - 46. Niehuis, O. et al. (2013) Behavioural and genetic analyses of *Nasonia* shed light on the evolution of sex pheromones. *Nature* 494, 345-348
 - 47. Phelan, P.L. (1992) Evolution of sex pheromones and the role of asymmetric tracking. In *Insect chemical ecology: an evolutionary approach* (Roitberg, B.D. and Isman, M.B., eds), pp. 265-314, Springer Science & Business Media
 - 48. Moehring, A.J. and Boughman, J. W. (2019) Veiled preferences and cryptic female choice could underlie the origin of novel sexual traits. *Biol. Lett.* 15, 20180878
 - 49. Anton, S. and Rössler, W. (2020) Plasticity and modulation of olfactory circuits in insects. *Cell Tissue Res.* 1-16
 - 50. Johnson, T.L. et al. (2017) Anticipatory flexibility: larval population density in moths determines male investment in antennae, wings and testes. *Proc. R. Soc. Lond.* B 284, 20172087.
 - 51. Groot, A.T. (2014) Circadian rhythms of sexual activities in moths: a review. *Front. Ecol. Evol.* 2, 43
 - 52. Jin, S. et al. (2017) Olfactory plasticity: variation in the expression of chemosensory receptors in *Bactrocera dorsalis* in different physiological states. *Front. Physiol.* 8, 672
 - 53. Wan, X. et al. (2015) Synthetic pheromones and plant volatiles alter the expression of chemosensory genes in *Spodoptera exigua*. *Sci. Rep.* 5, 1-11
- 532 54. Sollai, G. et al. (2018) Olfactory sensitivity to major, intermediate and trace components of sex pheromone in *Ceratitis capitata* is related to mating and circadian rhythm. *J. Insect Physiol.* 110, 23-33
- 55. Anderson, P. et al. (2003) Pre-exposure modulates attraction to sex pheromone in a moth. *Chem. Senses* 28, 285-291

537 56. Anderson, P. et al. (2007) Increased behavioral and neuronal sensitivity to sex pheromone after brief odor experience in a moth. *Chem. Senses* 32, 483-491

- 57. Levakova, M. et al. (2018) Moth olfactory receptor neurons adjust their encoding efficiency to temporal statistics of pheromone fluctuations. *PLoS Comput. Biol.* 14, e1006586
 - 58. van Schooten, B. et al. (2020) Divergence of chemosensing during the early stages of speciation. *Proc. Natl. Acad. Sci. U. S. A.* 117, 16438-16447
 - 59. Kromann, S.H. et al. (2015) Concurrent modulation of neuronal and behavioural olfactory responses to sex and host plant cues in a male moth. *Proc. R. Soc. Lond. B* 282, 20141884
 - 60. Diesner, M. et al. (2018) Mating-induced differential peptidomics of neuropeptides and protein hormones in *Agrotis ipsilon* moths. *J. Proteome Res.* 17, 1397-1414
 - 61. Lalouette, L. et al. (2016) Unexpected effects of sublethal doses of insecticide on the peripheral olfactory response and sexual behavior in a pest insect. *Environ. Sci. Pollut. Res.* 23, 3073–3085
 - 62. Gharaei, A.M. et al. (2020) Modulation of Reproductive Behavior of *Diaphania indica* (Lepidoptera: Pyralidae) by Preferred and Non-preferred Host Plants. *J. Insect Behav.* 33, 105-115
 - 63. Xu, H. and Turlings, T.C. (2018) Plant volatiles as mate-finding cues for insects. *Trends Plant Sci.* 23, 100-111
 - 64. McCormick, A.C. et al (2017) Exploring the effects of plant odors, from tree species of differing host quality, on the response of *Lymantria dispar* males to female sex pheromones. *J. Chem. Ecol.* 43, 243-253
 - 65. Hoffmann, A. et al. (2020) A plant volatile alters the perception of sex pheromone blend ratios in a moth. *J. Comp. Physiol.* 206, 553-570
 - 66. Ruiz-Guzmán, G. et al. (2021) Interactions between oxidative stress and attractiveness to mates and individual mate choice in the beetle *Tenebrio molitor*. *Ethology* 127, 109-116
 - 67. Reyes-Ramírez, A. et al. (2019) Female choice for sick males over healthy males: Consequences for offspring. *Ethology* 125, 241-249
 - 68. Barthel, A. et al. (2015) Sex-specific consequences of an induced immune response on reproduction in a moth. *BMC Evol. Biol.* 15, 1-12
 - 69. Altizer, S.M. et al. (2000) Associations between host migration and the prevalence of a protozoan parasite in natural populations of adult monarch butterflies. *Ecol. Entomol.* 25, 125-139
 - 70. Derstine, N.T. et al. (2017) Evidence for sex pheromones and inbreeding avoidance in select North American yellowjacket species. *Entomol. Exp. Appl.* 164, 35-44
 - 71. van Bergen, E. et al. (2013) The scent of inbreeding: a male sex pheromone betrays inbred males. *Proc. R. Soc. Lond. B* 280, 20130102
 - 72. Chang, H. et al. (2017) A pheromone antagonist regulates optimal mating time in the moth *Helicoverpa armigera*. *Curr. Biol.* 27, 1610-1615
 - 73. Ruther, J. et al. (2009) Quantity matters: male sex pheromone signals mate quality in the parasitic wasp *Nasonia vitripennis*. *Proc. R. Soc. Lond. B* 276, 3303-3310
 - 74. Mowles, S.L. et al. (2013) A female-emitted pheromone component is associated with reduced male courtship in the parasitoid wasp *Spalangia endius*. *PLoS One* 8, e82010
 - 75. Fitzpatrick, B.M. (2012) Underappreciated consequences of phenotypic plasticity for ecological speciation. *Int. J. Ecol.* 2012, 256017
 - 76. Geiselhardt, S. et al. (2012) Looking for a similar partner: host plants shape mating preferences of herbivorous insects by altering their contact pheromones. *Ecol. Lett.* 15, 971-977

587 77. Xue, H.J. et al. (2016) Contact pheromones of 2 sympatric beetles are modified by the host plant and affect mate choice. *Behav. Ecol.* 27, 895-902

- 78. Emelianov, I. et al. (2001) Host-induced assortative mating in host races of the larch budmoth. *Evolution* 55, 2002-2010
 - 79. Bengtsson, M. et al. (2014) Pheromone races of *Cydia splendana* (Lepidoptera, Tortricidae) overlap in host plant association and geographic distribution. *Front. Ecol. Evol.* 2, 46
 - 80. Frago, E. et al. (2019) Common pheromone use among host-associated populations of the browntail moth, *Euproctis chrysorrhoea*, displaying different adult phenologies. *Entomol. Gen.* 39, 295-306
 - 81. Vuts, J. et al. (2018) Host shift induces changes in mate choice of the seed predator *Acanthoscelides obtectus* via altered chemical signalling. *PLoS One* 13, e0206144
 - 82. Zakir, A. et al. (2017) Herbivore-induced changes in cotton modulates reproductive behavior in the moth *Spodoptera littoralis*. *Front. Ecol. Evol.* 5, 49
 - 83. Svensson, E.I. (2012) Non-ecological speciation, niche conservatism and thermal adaptation: how are they connected? *Org. Divers. Evol.* 12, 229-240
 - 84. Lassance, J.M. et al. (2019) Pheromones and barcoding delimit boundaries between cryptic species in the primitive moth genus *Eriocrania* (Lepidoptera: Eriocraniidae). *J. Chem. Ecol.* 45, 429-439
 - 85. Dion, E. et al. (2020) Early-exposure to new sex pheromone blends alters mate preference in female butterflies and in their offspring. *Nat. Commun.* 11, 1-8
 - 86. Lassance, J.M. et al. (2013) Functional consequences of sequence variation in the pheromone biosynthetic gene pgFAR for *Ostrinia* moths. *Proc. Natl. Acad. Sci. U. S. A.* 110, 3967-3972
 - 87. Buček, A. et al. (2015) Evolution of moth sex pheromone composition by a single amino acid substitution in a fatty acid desaturase. *Proc. Natl. Acad. Sci. U. S. A.* 112, 12586-12591
 - 88. de Fouchier, A. et al. (2017) Functional evolution of Lepidoptera olfactory receptors revealed by deorphanization of a moth repertoire. *Nat Commun.* 8, 1-11
 - 89. Koutroumpa, F.A. et al. (2016) Genetic mapping of male pheromone response in the European corn borer identifies candidate genes regulating neurogenesis. *Proc. Natl. Acad. Sci. U. S. A.* 113, E6401-E6408
 - 90. Groot, A.T. et al. (2016) The genetic basis of pheromone evolution in moths. *Annu. Rev. Entomol.* 61, 99-117
 - 91. Byers, K.J. et al. (2020) A major locus controls a biologically active pheromone component in *Heliconius* melpomene. *Evolution* 74, 349-364
 - 92. Byers, K.J. et al. (2021) Clustering of loci controlling species differences in male chemical bouquets of sympatric *Heliconius* butterflies. *Ecol. Evol.* 11, 89-107
 - 93. Evenden, M.L. (2016) Mating Disruption of Moth Pests in Integrated Pest Management: A Mechanistic Approach. In *Pheromone Communication in Moths: Evolution, Behavior and Application*. (Allison, J. and Cardé, R.T., eds), pp. 365-393, University of California Press, Oakland, California, USA.
- 94. Mochizuki, F. et al. (2002) Resistance to a mating disruptant composed of (Z)-11-tetradecenyl acetate in the smaller tea tortrix, *Adoxophyes honmai* (Yasuda)(Lepidoptera: Tortricidae). *Appl. Entomol. Zool.* 37, 299-304
- 632 95. Mochizuki, F. et al. (2008) Sex pheromone communication from a population 633 resistant to mating disruptant of the smaller tea tortrix, *Adoxophyes honmai* Yasuda 634 (Lepidoptera: Tortricidae). *Appl. Entomol. Zool.* 43, 293-298

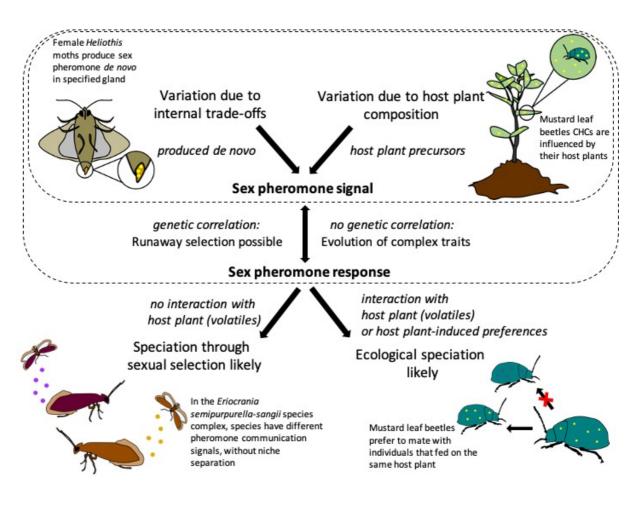
96. Tabata, J. et al. (2007) Sex pheromone production and perception in the mating disruption-resistant strain of the smaller tea leafroller moth, *Adoxophyes honmai. Entomol. Exp. Appl.* 122, 145-153
97. Evenden, M.L. and Haynes, K.F. (2001) Potential for the evolution of resistance to pheromone-based mating disruption tested using two pheromone strains of the cabbage looper, *Trichoplusia ni. Entomol. Exp. Appl.* 100, 131-134

- 98. Hodgdon, E.A. et al. (2020) Synthetic pheromone exposure increases calling and reduces subsequent mating in female *Contarinia nasturtii* (Diptera: Cecidomyiidae). *Pest Manag. Sci.* 77, 548-556
- 99. Judd, G.J. et al. (2005) Reduced antennal sensitivity, behavioural response, and attraction of male codling moths, *Cydia pomonella*, to their pheromone (E, E)-8, 10-dodecadien-1-ol following various pre-exposure regimes. *Entomol. Exp. Appl.* 114, 65-78
- 100. Groot, A.T. et al. (2013) One quantitative trait locus for intra-and interspecific variation in a sex pheromone. *Mol. Ecol.* 22, 1065-1080
- 101. Nieberding, C.M. et al. (2012) Cracking the olfactory code of a butterfly: the scent of ageing. *Ecol. Lett.* 15, 415-424
- 102. Chemnitz, J. et al. (2015) Beyond species recognition: somatic state affects long-distance sex pheromone communication. *Proc. R. Soc. Lond. B* 282, 20150832
- 103. Chemnitz, J. et al. (2017) Staying with the young enhances the fathers' attractiveness in burying beetles. *Evolution* 71, 985-994
- 104. Mérot, C. et al. (2015) Beyond magic traits: Multimodal mating cues in *Heliconius* butterflies. *Evolution* 69, 2891-2904
- Darragh, K. et al. (2017) Male sex pheromone components in *Heliconius* butterflies released by the androconia affect female choice. *PeerJ* 5, e3953.
- 106. González-Rojas, M.F. et al. (2020) Chemical signals act as the main reproductive barrier between sister and mimetic *Heliconius* butterflies. *Proc. R. Soc. Lond. B* 287, 20200587
- 107. Merot, C. et al. (2017) What shapes the continuum of reproductive isolation? Lessons from *Heliconius* butterflies. *Proc. R. Soc. Lond. B* 284, 20170335

Figure 1. Examples of Some Species for Which Intraspecific Variation in Sex Pheromone
Has Been Described in Literature. A) Asian lady beetle (*Harmonia axyridis*) (Photo: Chiara
De Pasqual); B) tobacco budworm (*Heliothis virescens*) (Photo: Jan van Arkel/IBED/UvA);
C) *Heliconius erato* (C) and *Heliconius melpomene* (D) (Photo: Melanie Brien)



Figure 2. Some Possible Routes to Speciation through Variation in Pheromone Production and Perception in Herbivorous Insects. Arrows indicate two of the possible alternative routes in the evolution of sex pheromone signals and responses. Pictures indicate examples discussed in this paper, clockwise from top left (*Heliothis* spp. [24]; mustard leaf beetle (*Phaedon cochleariae*) [75]; *Eriocrania semipurpurella-sangii* [83].



701 Table 1. Examples of Intraspecific Sex Pheromone Variation for Which Causes and/or

Consequences of the Variation Has Been Described and Genetic Basis or Heritability is

Known.

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	Level of variation ^a					
Species name	Quantitative b	Qualitative °	Cause of variation	Consequences of the variation	Genetic basis known or heritability calculated	References
Tobacco	_ d	Yes	Single point mutation	Female attractiveness & mating success affected		[21,35,40]
budworm (Heliothis virescens)	Yes	Yes	Pathogen infection boosted immune system	Sex specific: females mating success affected	Genetic basis known	[67]
Subflexus straw (Heliothis subflexa)	-	Yes	Phenotypic plasticity	Increase in assortative attraction	Genetic basis known	[14,99]
Squinting bush brown (Bicyclus anynana)	Yes	-	Seasonal polyphenis m or age	Mating success affected	Heritability known	[13,100]
Parasitic wasp (Nasonia vitripennis)	Yes	-	Sperm limitation (highly correlated with pheromone titre)	Attractions and mating success affected	Genetic basis known	[72]
Burying beetle (Nicrophorus vespilloides)	Yes	Yes	Diet or age, or parasite load	Male attractiveness affected	Unknown	[101,102]

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^a Variation in sex pheromones can take two forms: quantitative and qualitative.

⁷⁰⁷ b Quantitative when the variation is referred to the total amount of the components.

^{708 °} Qualitative when the variation is referred to the relative amounts and/or ratio of the components.

^{709 &}quot;-" refers to not-tested or not reported.

Outstanding Questions 1) How do interspecific and intraspecific selection forces interact and cause population divergence? Individuals often are under multiple selective pressures resulting from abiotic and biotic sources, which may act in opposite, similar or complex ways. 2) Is only one sex the signaler and the other sex the receiver, or is there mutual mate choice? As signalers may become receivers in different phases of mate assessment, it is important to determine which signals are being used by which sex at the different phases, and how these signals may interact or evolve separately. 3) How much of the intraspecific variation is determined by genetic factors and how much plasticity occurs in pheromone signals and responses? Since both sex pheromone signals and responses have been found to be plastic, and plasticity can accelerate as well as impede evolution, we need to gain more knowledge on the fitness consequences and heritability of this plasticity. 4) When using artificial pheromone blends for mating disruption, could resistance development be delayed by using variable pheromone lures? Knowledge on the level of intraspecific variation in the pheromone signal may be used to develop lures with different pheromone blends to reduce evolution of resistance to pheromone artificial traps.

Box I Visual vs Chemical signals

In butterflies, mate attraction is typically determined by visual cues, while sex pheromones are usually involved in short-range attraction. Current research is uncovering many aspects of sex pheromone roles in butterflies, suggesting that chemical signals are much more involved than previously expected in diurnal species that commonly rely on visual cues. *Heliconius* butterflies (Figure 1C,D in main text) are well known for exhibiting Müllerian mimicry, in which unrelated and chemically defended species mimic each other warning signal to more efficiently advertise their unpalatability to predators. When two species share identical wing pattern, such as *H. timareta* and *H. melpomene* in Peru, chemical signals are important to discriminate against heterospecifics [103]. Sex pheromones have a great importance in conspecific recognition, act as pre-mating barriers and affect mate choice [104]. As male pheromones and female-preference have been found to be heritable at least in some *Heliconius* butterfly species [105], pheromone communication may play a role in their speciation process. Thus, even in the visually-orientated butterflies, pheromones and other chemical signals, are used as pre-mating barriers in addition to wing patterns, making these species multi-modal signalers [106].

In *Bicyclus anynana*, the male sex pheromone can provide a wealth of information on the signaler, from the male's age to individual identity [100]. This pheromone is also a plastic trait as a consequence of a more or less climatic favorable season for their reproductive success [38], which has implications in their mating success [13]. Their sex pheromone can be more variable within populations than between populations and possibly acts as precursors of an ongoing speciation process [8].

In both *Heliconius* and *Bicyclus* study systems, short-range chemical signals are thus involved in species recognition and therefore under natural selection, but have also been shown to act in intraspecific mate assessment and thus under sexual selection. These examples show that in a male-signaling, female responding system, the progress of our understanding of the roles of pheromone signals seems to run counter to that in the more commonly studies females-signaling systems.

Box II Ecological and Evolutionary Consequences in the Burying Beetle (*Nicrophorus vespilloides*)

A comprehensive example of intraspecific sex pheromone variation that merges ecologicaland evolutionary consequences of its variation comes from burying beetles (*Nicrophorus*vespilloides). Intraspecific differences in sex pheromone have been shown to reflect males'
condition, with males being more attractive when they are in better nutritional condition, of
older age, have a larger body size and bear less parasites [101]. Burying beetles start to attract
females once they have found a carcass suitable for reproduction, by releasing a sex
pheromone. Both parents are known to feed on the carcass, so it is beneficial to both the
parents and the offspring to mate only after a carcass has been found. Interestingly, males that
have already performed brood care produce and release a higher amount of sex pheromone,
and attract more females. Because of this positive loop, in which individuals that perform
parental care produce more sex pheromone and become more attractive to females, the
interplay between food source and heightened expression of secondary sexual trait (sex
pheromone) has been suggested to have contributed to the evolution of parental care [102].

788	Glossary
789	Assortative attraction or assortative mating: individuals with similar phenotypes are
790	attracted and mate with one another more frequently than expected under random mating
791	
792	Cuticular hydrocarbons (CHCs): cuticular hydrocarbons are molecules on the surface of
793	adult insects, generally serving as anti-desiccation compounds. In a number of species,
794	CHCs have also be found to play important roles in insect communication. One of these
795	roles is attraction and selection of potential mates
796	
797	Ecological speciation: a form of speciation that arises as a consequence of reproductive
798	isolation due to a change in ecological factors (e.g. change in the host plant, presence of
799	predators or parasites and, in general, any environmental factor)
800	
801	Genetic correlation: two or more sets of genetic loci expected to co-vary as a consequence
802	of, for example, linkage disequilibrium or pleiotropy
803	
804	Mating disruption: a pest management technique which prevent individuals to successfully
805	locate potential mates through the release of a synthetic sex pheromone that mimics the
806	species sex pheromone
807	
808	Plasticity: the phenomenon for which organisms with the same genotype manifest alternative
809	phenotypes in response to environmental conditions, experienced usually during
810	developmental time
811	
812	Preference function: along a continuous range of a signaling trait values, most receivers

813	express a preference for intermediate values, while few show preference towards
814	extreme values
815	
816	Reproductive character displacement: selection to avoid hybridization between species.
817	When two species have overlapping habitats and (signaling) traits, divergence in one
818	or more traits evolves in response to unfavorable interactions, e.g. cross-species
819	attraction. When the divergence is due to selection to avoid hybridization between
820	races/strains within species, the term 'reinforcement' is used
821	
822	Runaway selection: due to genetic correlation between the sender and receiver's trait,
823	changes in the sender's signal are inherited together with changes for the preference
824	for that signal. This leads to a positive feedback loop, whereby both signal and
825	preference for the signal become exaggerated over time
826	
827	Saltational event: evolutionary event which generates greatly or completely different
828	phenotypes in a few generations, due to a mutation with major phenotypic effect. For
829	example, a mutation in an enzyme that changes the stereochemistry of a pheromone
830	component
831	
832	Signaling sex: the sex releasing a sexual communication signal to attract a potential mate
833	
834	Stabilizing selection: a form of selection in which the population mean trait converges to
835	intermediate values, e.g. when the mean pheromone signal is preferred over signals
836	deviating from the mean
837	
838	Sympatric species: species with overlapping geographic ranges that are close enough to

regularly interact