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Author(s): Kulomäki, Suvi; Lahtinen, Elmeri; Perämäki, Siiri; Väisänen, Ari

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Determination of mercury at picogram level in natural waters with inductively coupled plasma mass spectrometry by using 3D printed metal scavengers

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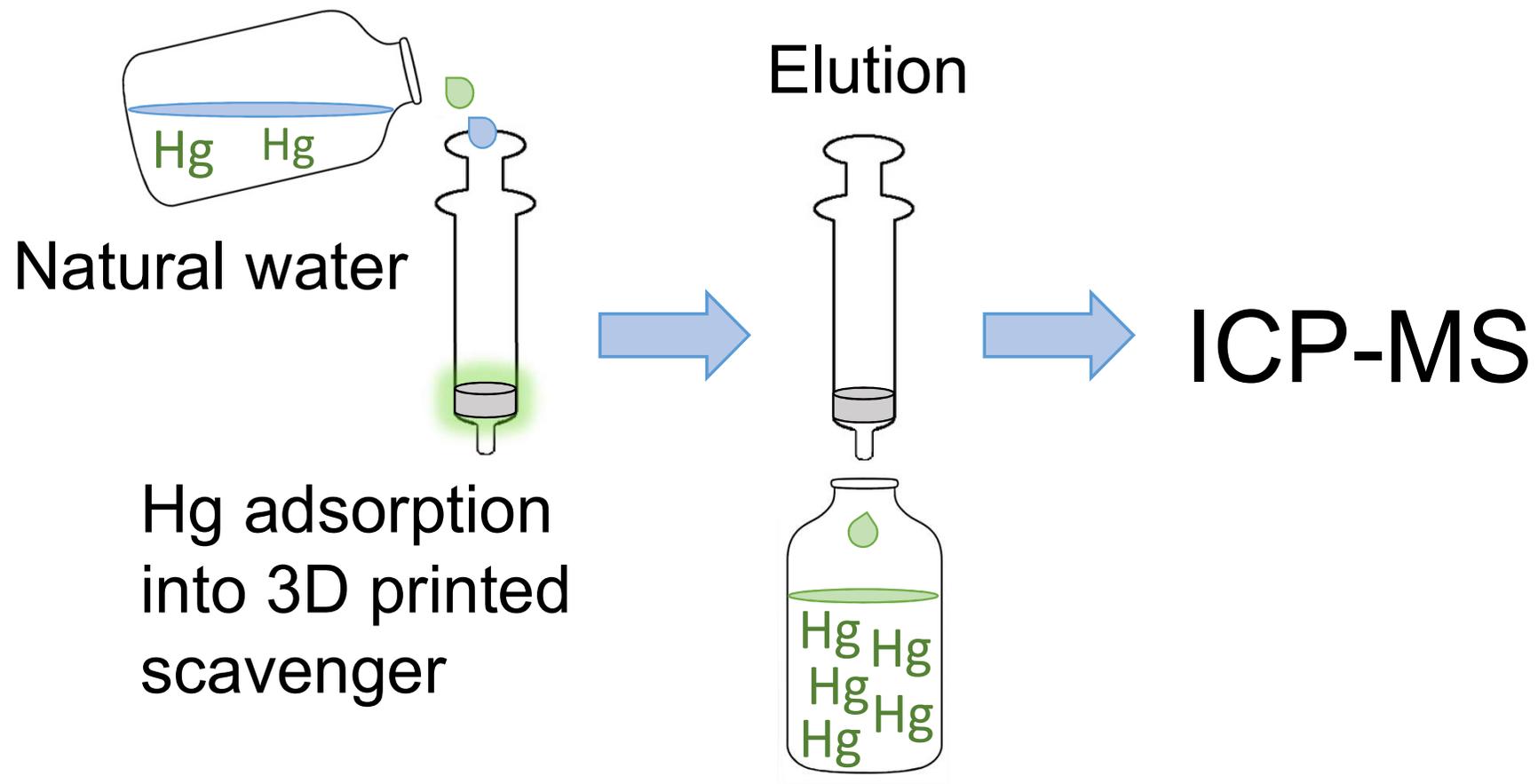
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1 **Determination of mercury at picogram level in natural waters with inductively coupled**
2 **plasma mass spectrometry by using 3D printed metal scavengers**

3 Suvi Kulomäki*, Elmeri Lahtinen, Siiri Perämäki, Ari Väisänen

4 Department of Chemistry, Renewable Natural Resources and Chemistry of Living Environment,
5 University of Jyväskylä, P.O. Box 35, FI-40014 Jyväskylä, Finland

6 **Abstract**

7 The determination of ultra-trace concentrations of Hg in natural water samples via
8 preconcentration using 3D printed metal scavenger technique followed by inductively
9 coupled plasma mass spectrometry (ICP-MS) was developed. The determination of Hg in
10 certified reference material ERM-CA615 (groundwater) was performed with high accuracy
11 and precision resulting in recovery of $100 \pm 3\%$ and $RSD < 2.5\%$, respectively. Selective laser
12 sintering (SLS) 3D printing was used to fabricate the scavengers using a mixture of
13 polyamide-12 powder with thiol-functionalized silica. The preconcentration procedure is
14 based on the adsorption of the Hg on the scavenger and followed by elution of the
15 preconcentrated Hg from the filter with 0.3% thiourea in 8% HCl prior to its determination by
16 ICP-MS. A preconcentration factor of 92.8 can be achieved by filtering 495 mL of water
17 followed with the elution step. Very low instrumental detection limit and method detection
18 limits were obtained resulting in 0.013 and 0.037 ng L⁻¹, respectively. The method was
19 applied successfully for the determination of Hg in different lake and river water samples.
20 The developed method is the first preconcentration method enabling simple and accurate
21 determination of Hg in pg L⁻¹ concentrations in natural waters with ICP-MS.

22 **Keywords:** Mercury; Inductively coupled plasma mass spectrometry; Ultra-trace
23 concentration; Preconcentration; 3D printing; Natural water.

24 *Corresponding Author. E-mail address: suvi.t.kulomaki@jyu.fi (S. Kulomäki)

1 1. Introduction

2 Mercury is one of the most toxic elements due to its accumulative and persistent nature in the
3 environment and living organisms. It occurs naturally in the environment in a variety of
4 chemical and physical forms at very low concentrations [1]. In pristine natural waters, Hg
5 concentrations are typically between pg and ng L⁻¹ range. Recent reports estimate a total
6 mercury concentration in natural waters ranging from 0.03 to 90 ng L⁻¹, while contaminated
7 waters may contain up to several µg L⁻¹ of Hg [1,2]. Although mercury is not an abundant
8 element in nature, its presence in polluted waters and the uptake of Hg by aquatic media has
9 become an area of great environmental concern and a potential risk to human health [3]. All
10 forms of mercury are poisonous and its toxicity is generally increased by transformation into
11 organic forms via biomethylation [1]. Monitoring of mercury traces in the hydrosphere has
12 been regulated in many countries, e.g., in Europe by the European Water Framework
13 Directive [2,4]. Hg is included in Annex I of the Directive 2013/39/EU as priority hazardous
14 substance and a maximum allowable concentration of Hg and its compounds of 0.07 µg L⁻¹ in
15 inland and other surface waters has been established [5,6]. In Finland, heavy metal
16 concentrations in natural waters are usually at a very low level. For this reason, the method
17 detection limits for Ni, Cd, Pb, and Hg are stricter than those presented in the Water
18 Framework Directive. For Hg, the recommendation for method detection limit is 5 ng L⁻¹ [7].
19 Accurate quantification of Hg traces in natural waters is challenging due to very low Hg
20 concentrations. Highly sensitive detection techniques are available for mercury e.g. cold
21 vapor atomic fluorescence spectrometry (CV-AFS) and inductively coupled plasma mass
22 spectrometry (ICP-MS) but most modern analytical techniques do not succeed in direct
23 determination of low Hg concentrations [1,2,8]. Thus, a preconcentration step is needed to
24 reach the detection limits required for the environmental monitoring of Hg [9]. The analytical
25 performance of non-chromatographic methods using different types of preconcentration

1 methods has been reviewed previously [1,10,11]. Most of these methods are using a tedious
2 cold vapor generation coupled to atomic spectroscopic techniques with detection limits
3 between 0.05 and 500 ng L⁻¹ for Hg²⁺. Still, many of these approaches have inadequate
4 detection limit for ultra-trace Hg analysis in environmental samples and the development of
5 simple and sensitive methods are highly desirable.

6 During the preconcentration step, the analyte is usually also separated from most of the
7 matrix elements, in order to eliminate possible interferences and lowering the detection limit
8 [12]. Various approaches have been applied for separation and preconcentration of Hg based
9 mainly on liquid-liquid extraction [13–15], co-precipitation [16,17] and solid-phase
10 extraction (SPE) [3,6,18–20]. SPE is the most commonly used preconcentration method for
11 trace metals due to its simplicity, flexibility, and ability to achieve high enrichment factors.
12 Extraction efficiency and selectivity are greatly dependent on the sorbent material used and
13 therefore the choice of the appropriate adsorbent with high affinity and selectivity for Hg is
14 the most critical step in SPE method development [12]. Numerous materials, such as
15 chelating resins [21], ion exchangers [22–24], modified carbon nanotubes [25,26] and
16 nanoparticles [2,3], modified silica gel [27] and polyurethane foam [28,29] have been
17 suggested as SPE sorbents. There are also some commercially available metal scavengers for
18 Hg but their disadvantage is that they are usually sold as powdery materials and hence
19 centrifugation or filtration is needed to recover the used adsorbent or to purify the sample
20 solutions from remaining particles. These difficulties make the use of the adsorbent laborious
21 and challenging. Moreover, the use of solid samplers instead of small particle material would
22 enable in situ sampling which would be almost impossible with a material consisting of small
23 particles. These challenges can be avoided by anchoring the chemically active component to
24 a sorbent matrix by using 3D printing.

1 Over the past few years, 3D printing has received increasing attention as a way to fabricate
2 macroscopic objects with actual functionality instead of just mechanical or aesthetic
3 properties [30–35]. 3D printing has found many potential applications in the fields of
4 separation sciences [36] and analytical chemistry [37]. However, many of these reports have
5 been focusing on either extrusion-based methods or stereolithography whereas methods such
6 as Selective Laser Sintering (SLS) have received less attention. This is even though the SLS
7 3D printing possesses some obvious advantages if targeting, for example, any kind of flow-
8 related processes. The challenge with the aforementioned extrusion-based and
9 stereolithography methods is that they produce objects, where the surfaces of the printed
10 objects are completely solid and the reactive surface area inside the object is not accessible.
11 The SLS 3D printing doesn't suffer from similar challenges as it uses a laser to partially
12 sinter small polymer beads together, leaving accessible voids between the polymer particles.
13 These voids are crucial for analytical chemistry purposes, as they allow the fluid to flow
14 through the pores of the objects and therefore allow for controlled chemical interaction
15 between the fluid and the object. This level of sintering, and therefore the porosity, can be
16 controlled by adjusting the printing parameters such as laser power, laser speed and
17 temperature [38–41]. Even different areas of the same object can be made to possess different
18 porosities. For example, the outer walls of the object can be made impermeable for fluids
19 while leaving the inside of the object highly macroporous [42]. The chemical functionality
20 can be embedded into the printed objects either by simply mixing the additive into the
21 starting material or by post-processing the printed objects. Almost anything can be used as
22 the additive since as long as the additive can withstand the temperature of the printing
23 process, it can be printed to form chemically functionalized macroscopic objects without
24 losing the activity of the additive during the process [42,43]. Turning powdery materials into

1 easily handleable objects with customizable shape, size and porosity often make their use in
2 actual applications much more feasible if compared to using them in their powdery form.

3 This paper presents a 3D printed Hg scavenger (3D-Thiol) method for the preconcentration
4 and determination of ultra-trace mercury concentrations in natural waters. A series of
5 sorption experiments were conducted using flow-through procedures to study the effect of
6 sorption, elution, and selectivity for mercury and some other metals. In addition, adsorption
7 isotherms for thiol functionalized silica (Thiol) and 3D-Thiol were determined. Finally, an
8 analytical method for Hg determination in ultra-trace concentrations was developed, based on
9 sorption of Hg on 3D-Thiol and elution using HCl and thiourea solution followed by ICP-MS
10 detection. To our knowledge, this is the very first study enabling Hg detection with ICP-MS
11 for pg L^{-1} concentrations without the utilization of cold vapor technique. In addition, this is
12 the first instance where thiol functionalized material, use-case and functionality have been
13 combined with SLS 3D printing.

14 **2. Experimental Section**

15 **2.1. Reagents**

16 All reagents were of analytical grade and all aqueous solutions were prepared in high purity
17 water ($18.2 \text{ M}\Omega \text{ cm}$) produced by Elga Purelab Ultra water purification system
18 (Buckinghamshire, U.K.). Single-element standard of Hg as Hg^{2+} (10 mg L^{-1} , PurePlus), Cd,
19 Co, Cr, Fe, Ir, K, Mg, Mn, Na, Ni, and Pb (1000 mg L^{-1} , Pure) were supplied by PerkinElmer.
20 1000 mg L^{-1} stock solution of Ca was prepared from CaCO_3 (Merck, $\geq 99.0\%$). The ICP-MS
21 performance was checked daily with a NexION Setup Solution ($1 \mu\text{g L}^{-1}$ Be, Ce, Fe, In, Li,
22 Mg, Pb, and U in 1% HNO_3 , PerkinElmer). Thiourea (99.8%) was obtained from VWR
23 Chemicals and SiliaMetS Thiol was purchased from SiliCycle (Quebec City, Canada).
24 Polyamide-12 (PA12), thermoplastic polyurethane and polypropylene printing powders were

1 purchased from BASF (Ludwigshafen am Rhein, Germany). Certified reference material
2 (ERM-CA615, groundwater) was produced and certified by the Joint Research Centre,
3 Institute for Reference Materials and Measurements (JRC-IRMM). Ultra-pure hydrochloric
4 acid (34–37%) was purchased from ANALYTIKA, spol. s r.o. (Prague, Czech Republic).
5 Puriss. p.a. hydrochloric acid ($\geq 37\%$) and nitric acid ($\geq 65\%$) were purchased from Merck and
6 were used for wash solutions with ESI prepFAST. All glass containers that could come into
7 contact with samples and standards were filled with 10% (v/v) hydrochloric acid ($\geq 37\%$) for
8 at least 24 h and tested for Hg followed by ICP-MS analysis. Sample bottles were first rinsed
9 thoroughly with ultrapure water and then completely filled with it. A blank sample was
10 prepared by using this water and was then analyzed with ICP-MS. Sample bottles were used
11 if the measured Hg concentration was below the instrument detection limit (IDL). After
12 testing, glass containers were filled with dilute HCl ($\sim 0.5\%$) and were thoroughly rinsed with
13 high purity water before usage. Calibration standards and smaller volume samples were
14 prepared into 50 mL polypropylene centrifuge tubes (VWR International).

15 **2.2. Sampling and sample pretreatment**

16 A total of seven different water samples were collected in 2L glass bottles from central
17 Finland. Tap water was collected from the research laboratory of the Department of
18 Chemistry (Jyväskylä), groundwater from Heinäahonlähde (Laukaa), Lake water 1 from lake
19 Päijänne (Korpilahti), Lake water 2 from lake Jyväsjärvi (Jyväskylä), River water from river
20 Tourujoki (Jyväskylä), Pond water 1 from Myllylampi (Tikkakoski) and pond water 2 from
21 Mustalampi (Jyväskylä). Samples were filtered (Whatman No. 41) only if they contained
22 solid material. Then they were acidified with ultra-pure HCl to obtain 3% (v/v) acid
23 concentration and stored at 4 °C in a refrigerator overnight if not analyzed immediately after
24 the sample collection.

1 2.3. Instrumentation

2 All mercury analyses were performed on a PerkinElmer (Massachusetts, U.S.A.) NexION
3 350D inductively coupled plasma mass spectrometer (ICP-MS) in standard mode. The ICP-
4 MS was interfaced with an Elemental Scientific (Nebraska, U.S.A.) prepFAST 4DX
5 sample/standard autodilution system. A NexION Setup Solution was used for the daily
6 optimization procedures for the ICP-MS according to manufacturer's instructions. Data were
7 acquired in counts per second (cps) for Hg (the sum of the counts at m/z 200, 201 and 202),
8 and Ir (m/z 193) was used as the internal standard. The most abundant W isotope ^{184}W was
9 monitored during all measurements, and it was concluded that WO did not interfere with Hg
10 in the studied water samples when the 3D-Thiol method was used. Calibration standards were
11 prepared in the range of 0–500 ng L^{-1} (blank, 50, 250 and 500 ng L^{-1}) Hg and the standards
12 and samples were adjusted to contain 3% (v/v) HCl. For analysis of elution solutions
13 containing thiourea and HCl, the standards were matrix-matched with 0.1125% (m/v)
14 thiourea and 3% (v/v) HCl. The system was first rinsed with a mixture of 0.75% (v/v) HNO_3
15 and 2.25% (v/v) HCl and then with 3% (v/v) hydrochloric acid between samples. The
16 optimum operating conditions and measurement parameters for ICP-MS are listed in Table 1.
17 Elements present in the selectivity study were analyzed on Avio 500 ICP-OES (PerkinElmer,
18 Massachusetts, U.S.A.) or with ICP-MS. More detailed information about the analytical
19 procedure is presented in Tables S1 and S2. Black/white 3.18 mm i.d. PVC peristaltic pump
20 tubing (PerkinElmer, Massachusetts, U.S.A.) was used for the preconcentration process and
21 the flow rate of the samples was adjusted using an ISCO (Nebraska, U.S.A.) WIZ peristaltic
22 pump. Helium imaging was performed with Carl Zeiss (Oberkochen, Germany) ORION
23 NanoFab helium ion microscope (HIM). The beam energy was set to around 30 kV while the
24 beam current varied between 0.211 and 0.234 pA. Scan dwell time of 0.2 μs with a viewing
25 distance of 5.101 to 7.833 millimeters was utilized for the imaging. Flood gun was used

1 throughout the imaging to counteract the charging event present. Samples did not receive any
2 additional pretreatment before the imaging process.

3 **2.4. Designing and preparation of the 3D printed scavengers**

4 The objects were designed using FreeCad v. 0.16 and then sliced with Slic3r v. 1.2.9 into 100
5 μm layers to obtain filters with desired shape and size (5 mm tall disc with 16.5 mm
6 diameter). The starting material for the SLS 3D printing was made by mixing polyamide-12
7 powder ($\sim 50 \mu\text{m}$ diameter) with either 5 or 10 wt% of thiol-functionalized silica (40–63 μm).
8 The material was then mechanically mixed and placed into the 3D printer. The actual printing
9 process was done using Sharebot SnowWhite SLS 3D printer with a laser power of 30–40%
10 (of a maximum of 14 W), a laser speed of 1800–2400 mm s^{-1} and printing temperature of 170
11 $^{\circ}\text{C}$. No post-processing, aside for cleaning the objects of any unsintered powder, was done for
12 the 3D printed objects.

13 **2.5. Adsorption Experiments**

14 The developed 3D-Thiol method was tested with model solutions prior to the separation and
15 preconcentration of Hg from real samples. For this purpose, 3D printed filters were placed
16 into a 10 mL syringe (Fig. 1) and washed from any unsintered powder with ultrapure water.
17 Extraction of Hg from aqueous solutions was investigated in a flow-through process. The
18 sorption was performed in centrifuge tubes containing 100 ng L^{-1} of Hg in 40 mL of 3% (v/v)
19 HCl. Initial tests were carried out using a peristaltic pump with a maximum flow rate (~ 8.3
20 mL min^{-1}). The adsorbed concentration of Hg was determined using ICP-MS. Adsorption
21 percentage was calculated using the following equation:

$$\text{Adsorption\%} = \frac{C_A - C_B}{C_A} \cdot 100\% \quad (1)$$

1 where C_A and C_B are initial and final concentration (ng L^{-1}) in the solution, respectively [44].
2 In the elution step, 2–10 mL of 0.3–2% (w/v) thiourea and 8–40% (v/v) hydrochloric acid
3 solution was passed through 3D-Thiol at a flow rate of $\sim 1\text{--}8.3 \text{ mL min}^{-1}$.

4 **2.6. Determination of adsorption capacity**

5 Equilibrium isotherm studies were carried out with different initial concentrations of Hg in
6 flow-through experiments. The adsorption capacity was determined for both Thiol and 3D-
7 Thiol. The adsorption efficiency of Thiol was performed by adding 10 mg of beads into 40
8 mL of different solutions containing different concentrations of Hg (1, 25, 50, 75 and 100 mg
9 L^{-1}) in 3% (v/v) HCl. The tube was sealed and then agitated using Stuart SF1 at 500 osc min^{-1}
10 for 2 h at room temperature. The thiol-functionalized silica was removed from the Hg
11 solution by filtration (Whatman No. 42) and samples were analyzed by using ICP-MS after
12 appropriate dilution.

13 The adsorption capacity for 3D-Thiol was performed with the same sample volume and Hg
14 concentrations as above. To obtain approximately 10 mg of Thiol in the printed scavenger,
15 the weight fraction of the additive was dropped to 5 wt% in the printed scavenger and 3D-
16 Thiol filters were placed into a 5 mL syringe. 40 mL solutions were passed through one filter
17 five times with a peristaltic pump at a maximum flow rate ($\sim 8.3 \text{ mL min}^{-1}$). After adsorption,
18 the residual concentration of Hg was analyzed by ICP-MS after appropriate dilution. The
19 concentration of Hg adsorbed per unit mass of the adsorbents was calculated from the
20 following equation:

$$q_e = \frac{(C_0 - C_e)V}{m} \quad (2)$$

1 where q_e is the adsorption capacity (mg g^{-1}) of the adsorbent at equilibrium; C_0 and C_e are the
2 initial and final concentration of Hg (mg L^{-1}), respectively; V is the volume of the
3 solution (L) and m is the mass of the adsorbent (g).

4 **2.7. Selectivity experiments**

5 A variety of 40 mL samples containing spiked lake water or synthetic sample were passed
6 through a 10 ml syringe containing one 3D-Thiol filter with a $\sim 8.3 \text{ mL min}^{-1}$ flow rate.

7 Spiking levels were 5 ng L^{-1} of Hg, 10 mg L^{-1} of Ca, 6 mg L^{-1} of Na, 2 mg L^{-1} of Mg, 1 mg L^{-1}
8 of K, 1 mg L^{-1} of Fe and $50 \text{ } \mu\text{g L}^{-1}$ of Cd, Co, Cr, Mn, Ni, and Pb. Ca, Na, Mg, K, Fe, Cr, and
9 Mn were measured with ICP-OES and Co, Ni, Cd, Pb, and Hg were analyzed with ICP-MS.

10 More detailed information about the analytical procedure is presented in Table S3.

11 **2.8. Spiking studies and the treatment of the real sample**

12 Samples were spiked with 20 ng L^{-1} of Hg and prepared in 45 mL volume of which 5 ml was
13 set aside for direct ICP-MS analysis. The real samples were prepared into 500 mL volume
14 and 5 mL of the sample was saved for direct measurement with ICP-MS. The sample solution
15 was pushed through the 3D-Thiol with $\sim 5 \text{ mL min}^{-1}$ flow rate using a peristaltic pump
16 resulting in the time needed for preconcentration to be about 1 h 40 min. The elution step was
17 done without a peristaltic pump in backflush mode. The retained Hg was eluted with 2 mL of
18 0.3% (w/v) thiourea in 8% (v/v) HCl solution. The syringe plunger was used to gently push
19 the first drop of the eluent through 3D-Thiol. Then the rest of the eluent was allowed to drain
20 through the filter. Finally, the 3D-Thiol was washed with 3.335 mL of ultrapure water to
21 obtain 3% (v/v) HCl concentration in the final solution, which was analyzed by ICP-MS. The
22 elution and washing take approximately 5 minutes.

23

24

1 **3. Results and discussion**

2 **3.1. Analytical performance of ICP-MS analysis**

3 The method showed good linearity over the calibration range of 0–500 ng L⁻¹ (Fig. S1) and
4 the correlation coefficient of linear regression (r^2) was higher than 0.9999. IDL achieved for
5 Hg determination was 1.2 ng L⁻¹ (calculated as three times the standard deviation of a series
6 of 10 replicate measurement of the calibration blank). The method detection limit (MDL)
7 was determined according to U.S Environmental Protection Agency method detection limit
8 procedure [45] and it was calculated to be 3.5 ng L⁻¹. The short and long term precision were
9 estimated by measuring a quality control sample containing 20 ng L⁻¹ of Hg, 20 mg L⁻¹ of C,
10 5 mg L⁻¹ of Ca, 1 mg L⁻¹ of Fe, 2 mg L⁻¹ of K, 2 mg L⁻¹ of Mg and 5 mg L⁻¹ of Na and 2 mg
11 L⁻¹ of S. The precision (relative standard deviation, RSD) within one day was 2.9% (n=7) and
12 during four weeks 4.0% (n=23). In addition, the precision and accuracy of the ICP-MS
13 method were estimated using a certified groundwater reference material (ERM-CA615,
14 certified value 37 ± 4 ng L⁻¹; expanded uncertainty at 95% confidence level). Replicate
15 analyses on two different days gave a mean value of 37.0 ng L⁻¹ (n = 5) with a precision of
16 2.5%.

17 **3.2. Selection of the chemically active component and supporting matrix**

18 Several commercially available metal scavengers and ion exchange materials were examined
19 for the preconcentration of Hg. Thiol-functionalized silica proved to be the most suitable
20 scavenger for Hg so it was chosen for a chemically active component for 3D printed
21 scavenger. Resins containing the thiol functional groups have been found to be able to adsorb
22 Hg [46,47]. Unfortunately, many of these are no longer commercially available (Spheron-
23 Thiol, Duolite GT73, Ambersep GT74). Polyamide-12, thermoplastic polyurethane, and
24 polypropylene printing powders were tested for 3D printing supporting matrix. PA12 proved

1 to be a suitable material since it is a readily available, durable and easily printable polymer.
2 In addition, Hg was not retained by the PA12 after elution with acidic thiourea solution.

3 **3.3. Macroscopic structure of the 3D printed filter**

4 The HIM imaging was used to get information about the macroscopic structure of the printed
5 object as well as the distribution and the attachment of the additive. HIM imaging, instead of
6 the more traditional SEM imaging, was conducted for the printed filters to combat the
7 charging effect often when imaging nonconductive samples [48]. It is also evident, that the
8 silica particles have not been encapsulated by the polymer and therefore they remain accessible
9 for chemical reactions (Fig. 2 left). The overall structure of the printed filter can be seen to be
10 highly macroporous, consisting of polymeric beads that have been partially sintered together
11 by the laser during the printing process. Additionally, the distribution of the silica particles
12 throughout the printed object seems to be rather even (Fig. 2 right). However, this was
13 expected as the distribution and the attachment for a similar system has been previously
14 reported [42,43] and thus was not studied more in depth here.

15 **3.4. Adsorption and adsorption isotherm studies**

16 Initial adsorption experiments were conducted using a peristaltic pump. During the
17 experiment, up to 97% of the Hg was adsorbed, which shows the rather satisfactory retention
18 kinetics. Complete adsorption could be achieved with a more moderate flow rate (~5 mL
19 min⁻¹). In this case, the Hg concentration in the remaining solution is very close or below the
20 instrument detection limit (1.2 ng L⁻¹) which indicates complete adsorption. Since the Hg
21 concentration in pristine natural waters can be only a few ng L⁻¹, a high preconcentration
22 factor is desired. The preconcentration factor can be maximized by increasing the sample
23 volume and decreasing the eluent volume. However, due to practical reasons such as ease of
24 sampling and duration of pretreatment, the volume of real natural water sample should be

1 kept reasonable. Taking these considerations into account, a sample volume of 500 mL was
 2 chosen, of which 5 mL was set aside for direct ICP-MS analysis.

3 The equilibrium conditions of the sorption process can be described using sorption isotherms.
 4 The most frequently used adsorption isotherm equations [23,49,50] of the Langmuir (3) and
 5 Freundlich (4) isotherms were applied to utilize the linearized equations indicated below:

$$\frac{C_e}{q_e} = \frac{1}{K_L \cdot q_{max}} + \frac{C_e}{q_{max}} \quad (3)$$

$$\ln q_e = \ln K_F + \frac{1}{n} \ln C_e \quad (4)$$

6 where q_e (mg g^{-1}) is the equilibrium adsorption capacity, C_e (mg L^{-1}) is the Hg concentration
 7 at equilibrium, q_{max} (mg g^{-1}) is the monolayer maximum adsorption capacity, K_L (L mg^{-1}) is
 8 Langmuir equilibrium constant, K_F (L g^{-1}) is a coefficient related to the amount of adsorbed
 9 metal ions and n is Freundlich adsorption constant related to surface heterogeneity. The
 10 Langmuir equation assumes the same energy for all adsorption sites and that monolayer is
 11 formed when the solid surface reaches saturation [51]. The plot of C_e/q_e against C_e gives a
 12 straight line and the values of q_{max} and K_L are obtained from the slope and intercept of the
 13 plot [23]. The Freundlich isotherm, in turn, is an empirical equation and assumes that
 14 adsorption is multilayer and is based on heterogeneous surface adsorption suggesting that
 15 binding sites are not equivalent [23,51].

16 Linear plots of $\ln q_e$ versus $\ln C_e$ revealed that the adsorption obeys well Freundlich isotherm
 17 (Fig. S2). The results showed that the Hg adsorption onto Thiol can be considered to be a
 18 multilayer adsorption system (Table 2). The loading capacity for Thiol is 264.8 mg g^{-1}
 19 according to the manufacturer. The experimental value obtained from the Langmuir isotherm
 20 model for Thiol was 253.5 mg g^{-1} and for 3D-Thiol 222.4 mg g^{-1} . These results indicate that
 21 3D printing decreases the maximum capacity of the resin only slightly. Hg concentrations in

1 natural waters are so low that the maximum loading capacity is not easily reached. For
2 example, Hg concentration of 46 mg L^{-1} with a sample volume of 500 mL is required to
3 saturate all absorption sites of 3D-Thiol.

4 **3.5. Selectivity**

5 In order to study the selectivity of 3D-Thiol towards Hg, competitive adsorption of Ca, Cd,
6 Co, Cr, Fe, K, Mg, Mn, Na, Ni, and Pb from spiked lake water and synthetic sample were
7 investigated. In spite of the high concentrations of other metal ions ($0.05\text{--}10 \text{ mg L}^{-1}$), Hg was
8 retained to 3D-Thiol completely as the Hg concentration of the sample that passed through
9 the filter was below the limit of detection. Moreover, Ca, Na, Mg, K, Cr, and Mn were not
10 retained at all, Fe, Co, Ni, Cd, and Pb showed very low percentage retention values to be \leq
11 1.0% (Table S4). 3D-Thiol proved to be extremely selective for Hg in 3% (v/v) HCl. These
12 results provide further evidence of the great selectivity of 3D-Thiol for Hg.

13 **3.6. The effect of eluent composition and its volume**

14 According to initial testing with different eluents and literature review [1,52,53], thiourea in
15 hydrochloric acid solution was found to be the most promising eluent. HCl has also been
16 reported [54] to have an enhanced performance due to its complexing abilities with mercury
17 and therefore it was of interest to investigate the influence of thiourea and HCl concentration
18 on the elution efficiency for 3D-Thiol. Thiourea was found to affect the intensity level of the
19 internal standard in ICP-MS analysis, hence, the samples with different eluent compositions
20 were analyzed without internal standard correction (Table 3). To enable the use of internal
21 standard, matrix matching was employed in subsequent analysis.

22 Effect of thiourea and HCl concentration on elution efficiency was studied in duplicates with
23 10 mL eluent volumes and $\sim 8.3 \text{ mL min}^{-1}$ flow rate without internal standard correction
24 (Table 3). Higher thiourea and acid concentration results in complete elution of Hg from the

1 3D-Thiol, however, it is desirable to keep the acid concentration and total dissolved solids
2 below 4% and 0.2%, respectively for ICP-MS [55]. Samples have to be diluted before
3 measurement if the acid concentration is above the tolerable limits and hence the higher
4 preconcentration factor would be lost. In addition, at least 4 mL sample volume is needed for
5 measurement. Because of these reasons, 0.3% thiourea in 8% HCl was chosen for further
6 studies.

7 The effect of eluent volume (2, 5 and 10 mL) and eluent flow rate ($\sim 1, 3$ and 5 mL min^{-1})
8 were evaluated using a peristaltic pump with the selected eluent. It was observed, that the
9 recovery was satisfactory (98–110%) with all tested eluent volumes and flow rates. However,
10 these small eluent volumes were not completely recovered when using a peristaltic pump. To
11 obtain quantitative recoveries, the elution step was performed manually without a peristaltic
12 pump as described in section 2.8. Considering the sample volume of 495 mL, final elution
13 volume of 2 mL and a dilution factor of 2.67, the preconcentration factor becomes 92.8. The
14 method detection limit for 3D-Thiol method becomes 0.037 ng L^{-1} when the preconcentration
15 factor is taken into account.

16 **3.7. Reusability of the sorbent**

17 The reusability of 3D-Thiol as a sorbent was examined by assessing the change in the
18 recoveries of the analyte through several sorption-elution cycles at a maximum flow rate of
19 the peristaltic pump following the procedure described in section 2.5. The results revealed
20 that one 3D-Thiol filter could be reused up to 10 times without a loss of adsorption efficiency
21 (Fig. S3). The recovery and relative standard deviation throughout the reusability test for
22 adsorption was 95.2% and 0.7%, respectively ($n=2$). The adsorption was satisfactory even
23 when the highest flow rate was used. As stated previously, the extraction efficiency is easily

1 increased with a more moderate flow rate. Nonetheless, in order to avoid any contamination,
2 new 3D-Thiol filters were employed throughout the study.

3 **3.8. Spiking tests**

4 A total of five spiked real samples were analyzed with the 3D-Thiol method as well as
5 directly without any preconcentration. The results obtained using 3D-Thiol method were
6 significantly closer to the spiked concentration of 20 ng L^{-1} compared with the direct
7 measurement as can be seen in Table 4. This can be seen more clearly in the case of yellow to
8 brown color waters (Lake water 1, River and Pond water) which contain more humic
9 substances than clearer waters (Tap water and Lake water 1). Some spiking tests were also
10 performed at 10 ng L^{-1} level, for which direct measurement from tap water gave 10.2 ng L^{-1}
11 (RSD 7.2%) and 3D-Thiol method 10.2 ng L^{-1} (RSD 1.7%). It is likely that the challenging
12 matrix of the colored waters suppresses the signal of mercury in direct ICP-MS determination
13 and might result in biased low concentrations (Table 4). With the 3D-Thiol method, Hg can
14 be separated from most of the matrix and interferences are reduced when compared to direct
15 determination. It was observed that RSD values could slightly be improved by using two 3D-
16 filters in the same syringe (Tables 4 and 5). As the amount of the functional group is
17 increased it is more likely that Hg interacts with thiol group and different kind of samples
18 behave in the same manner. Although the RSD values were improved with the use of two
19 3D-Thiol filters, RSD values were considered to be sufficient even when only one filter was
20 used. Obtained results further confirm the usefulness of the proposed method.

21 **3.9. Analysis of mercury concentration in real samples**

22 The proposed method was applied for the determination of Hg in different water samples.
23 The samples were analyzed both with the direct determination and with the preconcentration
24 method. With direct determination, Hg concentration of the water samples was below the

1 MDL except for one sample (Table 5). This represents the need for a 3D-Thiol method to
2 enable a reliable analysis of extremely low concentrations of Hg in natural waters. The
3 reproducibility of the method is very good considering the low ng L^{-1} concentrations. In
4 Central Finland, Hg concentrations of natural waters are at a very low level and with the 3D-
5 Thiol method, the recommended 5 ng L^{-1} method detection limit was easily reached.

6 **4. Conclusions**

7 A SLS 3D printed 3D-Thiol scavenger for Hg preconcentration was prepared and applied for
8 the determination of Hg from water samples with ICP-MS. The proposed method offers a
9 simple and highly selective method for the extraction and determination of ultra-trace Hg.
10 3D-Thiol has an excellent selectivity towards Hg over Ca, Cd, Co, Cr, Fe, K, Mg, Mn, Na,
11 Ni, and Pb. The interfering effects of the matrix are reduced when 3D-Thiol method is
12 applied. In addition, the loading capacity of 222.4 mg g^{-1} for 3D-Thiol is significantly higher
13 than adequate for this application. The captured Hg can be quantitatively eluted from the 3D-
14 Thiol filter with a mixture of 0.3% (w/v) thiourea and 8% (v/v) HCl solution. If desired, the
15 3D-Thiol scavenger can be reused at least 10 times with no decrease in its extraction
16 efficiency. The results indicate that the new 3D-Thiol is an exceptional candidate for Hg
17 sorption from natural waters. Due to the high preconcentration factor of 92.8 and extremely
18 low detection limit of 0.037 ng L^{-1} , ultra-trace amounts of Hg at pg L^{-1} levels in water
19 samples can be quantified by the method developed.

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2 use of ICP-MS and for water samples and Dr. Kimmo Kinnunen for the HIM imaging.

3 **Appendix A. Supplementary data**

4 Supplementary data to this article can be found online at <https://doi.org>.

5 **Author Contributions**

6 The manuscript was written through contributions of all authors. All authors have given
7 approval to the final version of the manuscript. S.K. did the Hg adsorption and elution
8 experiments and carried out the ICP-MS and ICP-OES analysis. E.L. prepared the 3D model
9 of the scavenger and performed 3D printing. S.P. assisted in the designing of the experiments
10 and A.V. supervised the project.

11

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6

Journal Pre-proof

1 **Captions to figures**

2 **Fig. 1.** SLS 3D printed 5 mm (diam. 16.5 mm) 3D-Thiol filter (left) and filter placed in a
3 syringe (right).

4 **Fig. 2.** HIM image showing the attachment of the thiol-functionalized silica particle onto the
5 3D printed PA12 framework (left). HIM image of the overall macroporous structure of the
6 3D printed filter containing 10 wt% of functionalized silica on the PA12 framework. Thiol-
7 functionalized silica particles are highlighted for clarity.

8

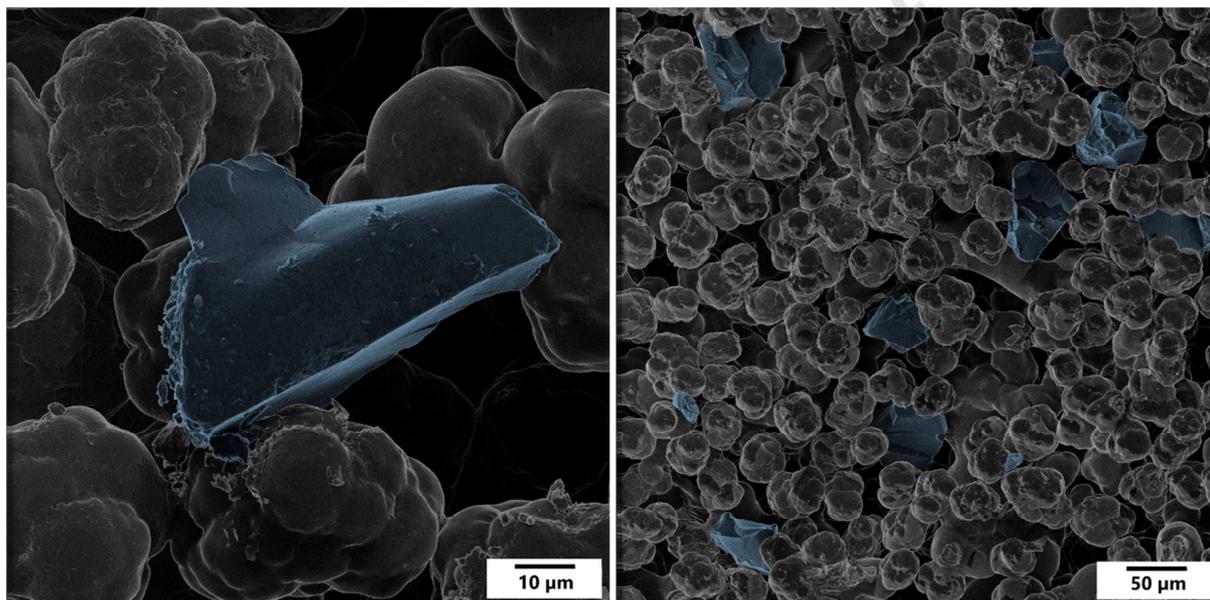
1 Fig. 1



2

3

4 Fig. 2



5

6

1 **Table 1.** ICP-MS instrumental operating parameters and conditions

System/ Operating parameter	Type/Value/Mode
Sample introduction	ESI prepFAST
Nebulizer	PFA-ST nebulizer for prepFAST
Spray chamber	Baffled cyclonic, glass, cooled to 2 °C
RF power	1600 W
Plasma gas flow rate	18 L min ⁻¹
Auxiliary gas flow rate	1.2 L min ⁻¹
Nebulizer gas flow rate	0.89–0.93 L min ⁻¹
Isotope	The sum of ²⁰⁰ Hg, ²⁰¹ Hg, and ²⁰² Hg
Internal standard	¹⁹³ Ir at 100 µg L ⁻¹ with 10 ms dwell time
Detector mode	Dual
Sweeps/reading	50
Replicates	5
Dwell time	100 ms
Scan mode	Peak hopping
Curve type	Linear through zero

2

3

1 **Table 2.** Constants of examined isotherms for Hg adsorption

Isotherm model	Constants	Value for 3D-Thiol (n=1)	The average value for Thiol (n=3)
Langmuir	q_{max} (mg g ⁻¹)	222.4	253.5
	K_L (L mg ⁻¹)	0.160	0.337
	R^2	0.975	0.940
Freundlich	K_F (L g ⁻¹)	23.5	54.2
	n	1.53	2.07
	R^2	0.982	0.980

2

3

1 **Table 3.** Recovery (%) \pm std. deviation for Hg from 3D-Thiol using different eluent
2 compositions

Eluent	Recovery (%)	Required dilution factor for analysis
0.3% thiourea in 8% HCl	90.1 \pm 3.8	2.67
1% thiourea in 8% HCl	99.7 \pm 4.9	5
2% thiourea in 10% HCl	98.8 \pm 2.9	10
2% thiourea in 20% HCl	104.0 \pm 3.7	13.33
0.3% thiourea in 40% HCl	105.1 \pm 2.7	13.33
2% thiourea in 40% HCl	101.3 \pm 4.2	13.33

3

4

1 **Table 4.** Average Hg concentration (ng L^{-1}) \pm std. deviation for 20 ng L^{-1} spike to different
 2 matrices (n=3)

Sample	3D-Thiol method	Direct determination
Tap water ^b	23.27 ± 0.90	19.36 ± 0.45
Lake water 1 ^c	20.75 ± 1.46	15.23 ± 0.40
Lake water 2	20.81 ± 1.88	13.08 ± 0.09
River water	21.24 ± 3.96	12.02 ± 1.01
Pond water 1 ^{a,b}	18.96 ± 1.43	12.67 ± 0.44

3 ^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters, ^cn=2

4

- 1 **Table 5.** Average Hg concentration (ng L^{-1}) \pm std. deviation for different water samples with
 2 the 3D-Thiol method and direct determination (n=3)

Sample	3D-Thiol method	Direct determination
Tap water	0.61 ± 0.06	<MDL
Groundwater	0.34 ± 0.02	<MDL
Lake water 1	0.94 ± 0.09	<MDL
Lake water 2	1.79 ± 0.13	<MDL
River water ^{a,b}	1.39 ± 0.08	<MDL
Pond water 1 ^a	0.61 ± 0.12	<MDL
Pond water 1 ^{a,b}	0.65 ± 0.04	<MDL
Pond water 2	2.94 ± 0.12	4.44 ± 0.12

- 3 ^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters

Table 1. ICP-MS instrumental operating parameters and conditions

System/ Operating parameter	Type/Value/Mode
Sample introduction	ESI prepFAST
Nebulizer	PFA-ST nebulizer for prepFAST
Spray chamber	Baffled cyclonic, glass, cooled to 2 °C
RF power	1600 W
Plasma gas flow rate	18 L min ⁻¹
Auxiliary gas flow rate	1.2 L min ⁻¹
Nebulizer gas flow rate	0.89–0.93 L min ⁻¹
Isotope	The sum of ²⁰⁰ Hg, ²⁰¹ Hg, and ²⁰² Hg
Internal standard	¹⁹³ Ir at 100 µg L ⁻¹ with 10 ms dwell time
Detector mode	Dual
Sweeps/reading	50
Replicates	5
Dwell time	100 ms
Scan mode	Peak hopping
Curve type	Linear through zero

Table 2. Constants of examined isotherms for Hg adsorption

Isotherm model	Constants	Value for 3D-Thiol (n=1)	The average value for Thiol (n=3)
Langmuir	q_{max} (mg g ⁻¹)	222.4	253.5
	K_L (L mg ⁻¹)	0.160	0.337
	R ²	0.975	0.940
Freundlich	K_F (L g ⁻¹)	23.5	54.2
	n	1.53	2.07
	R ²	0.982	0.980

Table 3. Recovery (%) \pm std. deviation for Hg from 3D-Thiol using different eluent compositions

Eluent	Recovery (%)	Required dilution factor for analysis
0.3% thiourea in 8% HCl	90.1 \pm 3.8	2.67
1% thiourea in 8% HCl	99.7 \pm 4.9	5
2% thiourea in 10% HCl	98.8 \pm 2.9	10
2% thiourea in 20% HCl	104.0 \pm 3.7	13.33
0.3% thiourea in 40% HCl	105.1 \pm 2.7	13.33
2% thiourea in 40% HCl	101.3 \pm 4.2	13.33

Table 4. Average Hg concentration (ng L^{-1}) \pm std. deviation for 20 ng L^{-1} spike to different matrices (n=3)

Sample	3D-Thiol method	Direct determination
Tap water ^b	23.27 ± 0.90	19.36 ± 0.45
Lake water 1 ^c	20.75 ± 1.46	15.23 ± 0.40
Lake water 2	20.81 ± 1.88	13.08 ± 0.09
River water	21.24 ± 3.96	12.02 ± 1.01
Pond water 1 ^{a,b}	18.96 ± 1.43	12.67 ± 0.44

^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters, ^cn=2

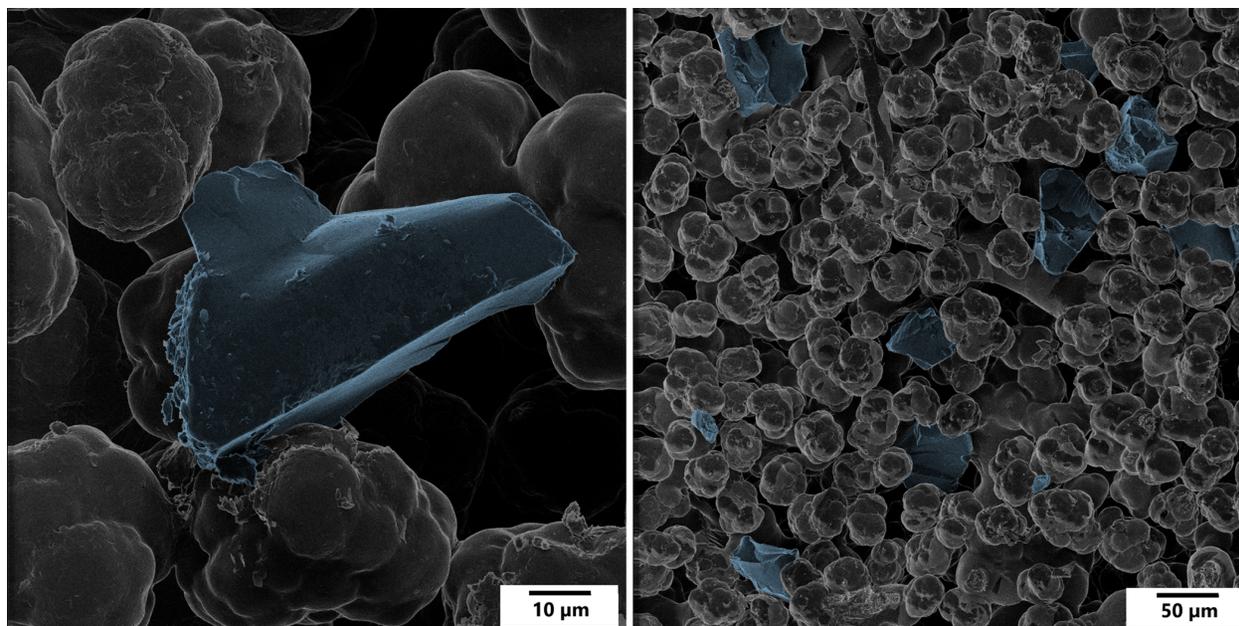
Table 5. Average Hg concentration (ng L^{-1}) \pm std. deviation for different water samples with the 3D-Thiol method and direct determination (n=3)

Sample	3D-Thiol method	Direct determination
Tap water	0.61 ± 0.06	<MDL
Groundwater	0.34 ± 0.02	<MDL
Lake water 1	0.94 ± 0.09	<MDL
Lake water 2	1.79 ± 0.13	<MDL
River water ^{a,b}	1.39 ± 0.08	<MDL
Pond water 1 ^a	0.61 ± 0.12	<MDL
Pond water 1 ^{a,b}	0.65 ± 0.04	<MDL
Pond water 2	2.94 ± 0.12	4.44 ± 0.12

^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters



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Highlights

- Highly selective and accurate preconcentration method for Hg
- Hg adsorption onto 3D printed metal scavenger and elution with acidic thiourea solution
- Accurate Hg determination at picogram level in natural waters with a preconcentration factor of 92.8
- Determination of background concentrations in natural water with ICP-MS

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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