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Author(s): Fei, Wenwen; Antonello, Sabrina; Dainese, Tiziano; Dolmella, Alessandro; Lahtinen, Manu; Rissanen, Kari; Venzo, Alfonso; Maran, Flavio

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Article

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Metal Doping of Au₂₅(SR)₁₈ Clusters: Insights and Hindsights

Wenwen Fei,[†] Sabrina Antonello,[†] Tiziano Dainese,[†] Alessandro Dolmella,[≠] Manu Lahtinen,[‡] Kari Rissanen,[‡] Alfonso Venzo,[‡] and Flavio Maran^{†,§},*

Keywords: atomically precise gold nanoclusters, Au₂₅(SR)₁₈, metal doping, NMR of doped gold nanoclusters, electrochemistry of doped gold nanoclusters, single crystal X-ray crystallography

E-mail: flavio.maran@unipd.it

Abstract. The study of the structures and properties of atomically precise gold nanoclusters is the object of active research worldwide. Recently, research has been also focusing on the doping of metal nanoclusters through introduction of noble metals, such as platinum, and less noble metals, such as cadmium and mercury. Previous studies, which relied extensively on the use of mass spectrometry and single crystal X-ray crystallography, led to assign the location where each of these foreign-metal atoms go. Our study provides new insights into this topic and, particularly, compelling evidence about the actual position of the selected metal atoms M = Pt, Pd, Hg, and Cd in the structure of Au₂₄M(SR)₁₈⁰. To make sure that the results were not dependent on the thiolate, for SR we used both butanethiolate and phenylethanethiolate. The clusters were prepared according to different literature procedures that were supposed to lead to different doping positions. Use of NMR spectroscopy and isotope effects, with the support of mass spectrometry, electrochemistry, and single crystal X-ray crystallography, led us to confirm that noble metals indeed dope the cluster at its central position, whereas no matter how the doping reaction is conducted and the nature of the ligand, the position of both Cd and Hg is always on the icosahedron shell, rather than at the central or staple position, as often reported. Our results not only provide a reassessment of previous conclusions, but also highlight the importance of NMR spectroscopy studies and cast doubts on drawing conclusions mostly based on single crystal X-ray crystallography.

INTRODUCTION

Many thiolate-protected gold nanoclusters, especially those sufficiently small (typically, less than ca. 144 atoms) to display electrochemical, optical, and magnetic molecular properties, can be prepared with atomic precision. Executly, research has also been focusing on the selective doping of metal nanoclusters through introduction of foreign-metal atoms. It is a very important area for both fundamental and applied (e.g., catalysis) purposes. Modification of the metal composition has been studied for several clusters, but most research has focused on Au₂₅(SR)₁₈, which is an atomically precise cluster that has been long considered a convenient benchmark system for understanding properties and devising applications of gold nanoclusters. ^{10,11}

Controlled doping of Au₂₅(SR)₁₈⁻ has been carried out with the noble metals platinum¹²⁻¹⁶ and palladium, ^{14,16-19} and less noble metals, such as cadmium²⁰⁻²² and mercury, ^{15,20-23} also because of the ease by which monodoping could be achieved with these metals as opposed to, say, copper and silver. ⁹ Mass spectrometry and single crystal X-ray crystallography were extensively employed to interpret the doping results and, particularly, assign the specific locations where these single foreign-metal atoms go. The platinum-doped clusters were prepared by direct synthesis, *i.e.*, by reacting a mixture of

tetrachloroauric and hexachloroplatinic acids with a given thiol, followed by sodium borohydride reduction. 12-15 The same procedure was applied to palladium^{14,16-19} and mercury.¹⁵ Cadmium²⁰⁻²² and mercury²⁰⁻²³ were introduced into preformed Au₂₅(SR)₁₈, and studied from several viewpoints. Regarding cadmium, Wu and co-workers reported that when the metal source is a salt, Cd(NO₃)₂, doping occurs on the icosahedron.²¹ The analysis of the data pertaining to Au₂₄Cd(SC2Ph)₁₈⁰ (SC2Ph = phenylethanethiolate; hereafter, we will indicate the number of carbon atoms of the alkyl chain simply as Cn) relied on X-ray crystallography and theoretical calculations of the experimental UV-vis-NIR spectrum, in comparison with corresponding mercury monodoped Au₂₄Hg(SC2Ph)₁₈⁰ and the pertinent matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectrometry fragmentation patterns. The latter also was obtained using a salt as the metal source, Hg(NO₃)₂.²³ Interestingly, the two very similar syntheses led to doping at different positions: whereas Cd would go on the icosahedron,21 for Hg the X-ray single-crystal diffraction results were interpreted to indicate that one of the staple Au atoms is replaced by Hg. Theoretical simulations of the experimental UV-vis-NIR spectrum, and the MALDI-TOF mass-spectrometry, thermogravimetric analysis,

[†] Department of Chemistry, University of Padova, via Marzolo 1, 35131 Padova, Italy

[‡] Department of Pharmaceutical and Pharmacological Sciences, University of Padova, via Marzolo 5, 35131 Padova, Italy

[#]University of Jyvaskyla, Department of Chemistry, P.O. Box 35, 40014 Jyväskylä, Finland

^{*} National Research Council, CNR-ICMATE, Department of Chemistry, University of Padova, via Marzolo 1, 35131 Padova

[§] Department of Chemistry, University of Connecticut, 55 North Eagleville Road, Storrs, Connecticut 06269, United States

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electrochemical results provided further support to this conclusion. ²³ Further work used this conclusion to understand what happens when Ag is used to dope a preformed Au₂₄Hg(SC2Ph)₁₈⁰ cluster. ²⁴ For both Au₂₄Hg(SC2Ph)₁₈⁰ and the so-formed trimetallic cluster, the NMR results were taken as a further indication that Hg atom most probably occupies a staple position.

Cadmium and mercury were introduced by the Zhu group into preformed Au₂₅(SC2Ph)₁₈ by using a different approach in which the metal is added to the cluster solution as a thiolate, Cd(SC2Ph)₂ or Hg(SC2Ph)₂.²⁰ For Cd, single-crystal X-ray crystallographic data indicated that doping occurred at the central position. As to Hg, the cluster was concluded to have the same structure due to the same valence, NMR spectrum, and the loss of the same M₁Au₄ fragment in MALDI-TOF mass spectrometry. NMR was used only to rule out the possible presence of the tetraoctylammonium countercation, present in the native Au₂₅(SC2Ph)₁₈ solution. As for the metal-salt studies, ^{21,23,24} the NMR spectrum showed a complex pattern. The outcome of the stepwise formation of trimetallic clusters, MAg_xAu_{24-x}(SC2Ph)₁₈, was interpreted²⁵ on the basis of the aforementioned conclusions on the central position eventually occupied by Cd or Hg.²⁰ Some doped clusters were prepared with ligands other than phenylethanethiol. In particular, Thanthirige et al. prepared $Au_{24}M(SC6)_{18}^{0}$ (M = Pt, Hg) clusters by direct synthesis. Analysis of the MALDI-TOF mass spectrometry and X-ray photoelectron (XPS) spectra led to conclude that for both metals the doping occurred at the central position.¹⁵ The preparation and other properties of $Au_{24}M(SC6)_{18}^{0}$ (M = Pt, Pd) were described by the Lee and Jiang groups in a previous publication. ¹⁴ Negishi et al. used dodecanethiol to prepare Au₂₄Pd(SC12)₁₈⁰ and concluded, on the basis of experiments and density-functional theory (DFT) calculations that Pd occupies the center of the core. 17,18

In a recent review article,8 Zhu and co-workers concluded that "Doping specific number of heterometal atoms into specific positions of the nanocluster template is still one of the most challenging tasks in the nanofield." We could not agree more. Indeed, now the question is: How can we assign the specific position where these foreign-metal atoms actually go to? This is not just a problem per se, but also has far-reaching consequences because the results described above are consistently taken as the starting point for other investigations, whether related to different clusters or applications, as discussed in several review articles.^{5,7-9,11,25-28} Here we address this problem by specifically focusing on the doping with Pt, Pd, Cd, and Hg atoms to form the corresponding $Au_{24}M(SR)_{18}^{0}$ clusters. As aforementioned, conclusions on the specific location of the foreign-metal atom have been drawn mostly on the basis of the interpretation of single-crystal Xray crystallography and MALDI-TOF mass spectrometry data, sometimes with the support of DFT calculations, and results from XPS and UV-vis absorption spectroscopy. The structure of Au₂₅(SR)₁₈, whether in the anionic or neutral form, ²⁹⁻³¹ is maintained in $Au_{24}M(SR)_{18}{}^0$, 16,19,20,21,23 and shows that there are three possible positions for the M atom: center (c), icosahedron (i), and staples (s) (Figure 1a); whereas there is only one central atom, the other positions are of 12-fold equivalency.

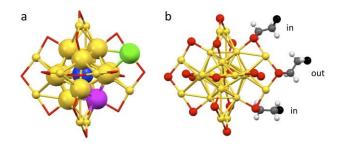


Figure 1. (a) Structure of $Au_{25}(SR)_{18}^{0/-}$ showing the three position types that can be occupied upon metal monodoping: central (blue), icosahedron (pink), and staple (green). The gold (yellow) and sulfur (red) atoms are shown, whereas the carbonaceous part of the ligands is omitted for clarity. (b) Structure of $Au_{25}(SR)_{18}^{0/-}$ showing the carbons (gray), hydrogens (white), and terminal groups (black) for both ligand types of one of the six staples.

Theoretical calculations have been performed to predict or explain the position of the heteroatom upon monometal doping, even ahead of substantial experimental work. Earlier DFT calculations by Jiang and Dai pointed to Cd and Hg as stable when in the center position.³² On the other hand, other DFT calculations carried out by Walter and Moseler predicted that Pd should be more stable when at the center, whereas for Cd the lowest energy isomer is at the icosahedron, rather than elsewhere.³³ In a recent study, Taylor and Mpourmpakis used³⁴ their thermodynamic stability model (TSM), which attributes structure stability to a balance between the chemical potentials of the metal atoms in the core and the protecting shell,³⁵ to describe doping effects on nanoclusters. The TSM predictions were concluded to be in excellent agreement with experiments. The case of Hg is particularly interesting. As we saw, Hg has been described as being at the center^{15,20} or in a staple position,²³ with the latter considered³⁴ as more likely and in agreement with the TSM. Although the icosahedral position, Hg(i), resulted close to the 95% prediction interval, a better proximity of the Hg(s) to the parity line (in a plot between shell-to-core bond energy and the metal-core cohesive energy) was seen as providing the first theoretical rationalization for the experimental observation of the Hg(s) position in Au₂₄Hg(SR)₁₈⁰. Regarding Cd, which was also described that, depending of the synthetic method, could occupy two positions, center²⁰ or icosahedron,²¹ the TSM results pointed to Cd(c) doping as being closer to the parity line than Cd(i) doping; thus, the authors also suggested that the latter could be potentially transformed into the former under proper experimental stimulus. Very recently, the Aikens group used DFT to study the doping process in a few clusters, including Au₂₄M(SR)₁₈. ³⁶ Whereas group X dopants (Pd, Pt) resulted stable when at the central position, for dopants in groups XI- XIII (and thus also for Cd and Hg, group XII) the icosahedral position was found thermodynamically preferable mainly due to group theory and relativistic effects. As to the staple position compared to the central position, whereas for Cd the former has a slightly lower energy, for Hg the results point to Hg(c) as being quite more stable than Hg(s).

With the exception of Pt and Pd, for which there is no doubt that the direct synthesis yields a cluster monodoped at its central position, it is thus clear that for Cd and Hg available experimental data provide a number of opposite conclusions, possibly related to the specific synthetic method. DFT

calculations have provided hints on this topic, though some analyses appear to be in contrast at least to some experimental Here we use **NMR** conclusions. spectroscopy, electrochemistry, MALDI-TOF, and single-crystal X-ray crystallography of each sample to demonstrate that in several cases the conclusions reached on the actual position of Cd and Hg atoms need to be drastically revised. Our study includes: (i) clusters protected by SC4 (Pt, Pd, Cd, Hg) and the SC2Ph (Cd, Hg) as the ligands; (ii) direct synthesis (Pt, Pd) and indirect synthetic methods, that is, metal exchange on both $\text{Au}_{25}(SR)_{18}^{-}$ (Cd, Hg) and $\text{Au}_{24}\text{Cd}(SR)_{18}^{0}$ (Hg); (iii) for Cd and Hg we used both the metal salt and the metal thiolate methods. This study is meant to provide new insights and perspectives into this general problem, and describe a possible experimental methodology to understand the actual doping location. The power of NMR spectroscopy and associated isotopic effects are especially highlighted. These results call for a warning about the reliability of conclusions based on mass-spectrometry fragmentation patterns and, especially, Xray crystallography of doped clusters.

RESULTS AND DISCUSSION

Synthesis. Au₂₄Pt(SC4)₁₈⁰ was prepared by reacting a solution of HAuCl₄ and H₂PtCl₆ with the given thiol, followed by addition of NaBH₄.¹² Au₂₄Pt(SC4)₁₈⁰ could be purified from the main co-product, Au₂₅(SR)₁₈, according to the procedure described by Qian et al., 12 in which H₂O₂ is used to cause degradation of the undoped cluster through multiple oxidation processes. For reasons that will be discussed in the X-ray crystallography section, we used *n*-butanethiol. The synthesis of Au₂₄Pd(SC4)₁₈0 was carried out according to a very similar protocol, but for the use of Na₂PdCl₆ in place of H_2PtCl_6 . Two $Au_{25}(SR)_{18}$ clusters (R = C4, C2Ph), which were prepared as already described, 37-39 were allowed to react with Cd(NO₃)₂ or Hg(NO₃)₂, as described by the Wu group, 21,23 and Cd(SR)2 or Hg(SR)2, as described by the Zhu group.20 These reactions are described in detail in the Experimental Section; we found that the same protocol works well for both C4 and C2Ph. In addition to using Cd(NO₃)₂ for making the thiolate, we used CdCl2, and the reactions went equally well. This check was expedient to then carry out the exchange reactions on Au₂₅(SR)₁₈ clusters with ¹¹³Cd(SR)₂. Finally, on the two Au₂₄Cd(SR)₁₈⁰ clusters we exchanged Cd with Hg, as described by the Wu group for $R = C2Ph.^{21}$ The clusters were carefully purified, recrystallized, and only afterward each sample batch was used for the NMR spectroscopy, UV-vis absorption spectroscopy (Figure 2 shows the SC4 series, whereas Figure S1 shows the SC2Ph series), MALDI-TOF mass spectroscopy, cyclic voltammetry pulse differential voltammetry and measurements. The synthetic methods used only yielded monodoped clusters. Most of them were also studied by single crystal X-ray crystallography. To check the quality of the results further, some of the crystals studied at the University of Jyväskylä were also studied at the University of Padova.

 $Au_{24}Pt(SC4)_{18}^0$ and $Au_{24}Pd(SC4)_{18}^0$. For both platinum and palladium, the direct synthesis of $Au_{24}M(SR)_{18}^0$ (R = C2Ph, C6, C12) has been consistently described to yield clusters doped at the center. ¹²⁻¹⁹ For R = C4, we followed the same synthetic and purification protocol described for platinum by Qian et al. ¹² and adapted for palladium by Kwak et al.. ¹⁴ MALDI-TOF mass spectrometry clearly indicated that the purified clusters only contain one foreign-metal atom, as

shown in Figure S2 for $Au_{24}Pt(SC4)_{18}^{0}$, and no contamination from residual $Au_{25}(SC4)_{18}^{-}$ or $Au_{25}(SC4)_{18}^{0}$.

The NMR behaviors of $Au_{24}Pt(SC4)_{18}{}^0$ and $Au_{24}Pd(SC4)_{18}{}^0$ were studied in C_6D_6 , at 1.5-2.1 mM concentration of the cluster, and the chemical shifts (δ) are referred to tetramethylsilane; these conditions were the same also for all other clusters studied. Beside the monodimensional 1H NMR spectra, the clusters were studied by 1H , 1H -homonuclear correlation spectroscopy (COSY) and 1H , 1S C-heteronuclear multiple quantum coherence (HMQC) spectroscopy. The 2D spectra, whose analysis allowed assigning all resonances, are provided in Figures S3-S5, whereas Table S1 gathers all chemical shift values. $Au_{24}Pt(SC4)_{18}{}^0$ and $Au_{24}Pd(SC4)_{18}{}^0$ are diamagnetic, neutral species 16 and are thus directly comparable with the diamagnetic anion $[Au_{25}(SC4)_{18}]$ (n-Oct₄N) $^+$ (where n-Oct stands for norm-octyl), whose NMR data 37,38 are also provided in Table S1.

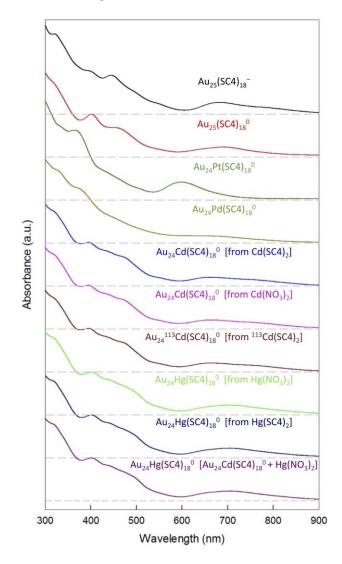


Figure 2. UV-vis absorption spectra of all SC4 samples (0.2 mM, 1 mm cuvette) in CH₂Cl₂. For the sake of better comparison, the curves have been shifted vertically. The dashed lines mark the corresponding zero absorbance.

 Au_{25} and monodoped clusters are known to share the same structural features: a central Au atom, 12 Au atoms forming an icosahedron, and an external shell composed of six $-(SR)_{in}$ -

Au-(SR)_{out}-Au-(SR)_{in}- double staples. Each staple consists of two inner thiolates (in) and one outer thiolate (out) (Figure 1b). The term inner indicates that the two SR groups also bind to the icosahedron Au atoms, whereas outer indicates that the SR group is at the outmost position of the double staple. In $[Au_{25}(SC4)_{18}]$ (n-Oct₄N)⁺, ^{37,38} the two ligand types have different δ values, well-defined signals (corresponding to the methylene groups in positions α , β , and γ with respect to sulfur, and the methyl group in position δ), and for the same resonance the integrals are in the 2:1 ratio expected for the 12 inner and 6 outer ligands. The spectra of Au₂₄Pt(SC4)₁₈⁰ and Au₂₄Pd(SC4)₁₈⁰ exhibit exactly the same general features, but for slightly different chemical-shift values (Table S1). Figure 3 shows the comparison between the Pt-doped and the undoped clusters. This behavior clearly indicates that the symmetry of the ligands of the parent Au₂₅(SC4)₁₈ cluster is preserved upon monodoping, as also discussed for R = C2Ph by Qian et al.¹² and later by Tian et al.¹⁶ For both Pt and Pd, the ¹H NMR spectra of Au₂₄M(SC4)₁₈⁰ can thus be taken as representing the blueprint of the typical "uncomplicated" NMR behavior expected for a Au₂₄M(SC4)₁₈⁰ cluster doped in its central position.

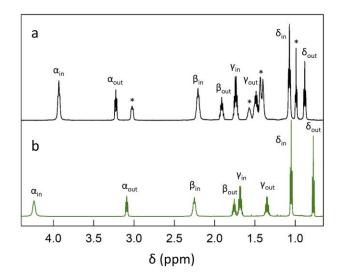


Figure 3. ¹H NMR spectra of (a) $[n\text{-Oct}_4N^+]$ $[Au_{25}(SC4)_{18}^-]$ and (b) $Au_{24}Pt(SC4)_{18}^0$. The peaks marked with a star pertain to $n\text{-Oct}_4N^+$. Both samples were in C_6D_6 at 25 °C.

 $Au_{24}Hg(SC4)_{18}{}^0.$ The mercury-doped clusters were prepared according to three previously published methods (for C2Ph), *i.e.*, by the (i) Au-exchange reaction of $Au_{25}(SC4)_{18}{}^-$ with $Hg(SR)_2$ (Wang et al.) 20 and (ii) $Hg(NO_3)_2$ (Liao et al.), and by the (iii) Cd-exchange reaction of $Au_{24}Cd(SC4)_{18}{}^0$ with $Hg(NO_3)_2$ (Yao et al.). After purification and recrystallization, all three methods led to obtain very pure $Au_{24}Hg(SC4)_{18}{}^0$ samples. The UV-vis absorption spectroscopy and MALDI-TOF spectra are shown in Figures 2 and S6, respectively.

We first consider the Au₂₄Hg(SC4)₁₈⁰ sample prepared from Hg(SR)₂. Figure 4 shows its ¹H NMR spectrum and identification of the signals, as achieved by analysis of the COSY and total correlation spectroscopy (TOCSY) spectra

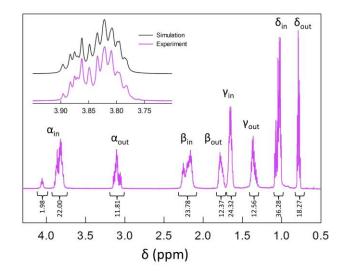


Figure 4. ¹H NMR spectra of 2.1 mM Au₂₄Hg(SC4)₁₈⁰ in C₆D₆ at 25 °C. The Greek symbols have the usual meaning. The inset shows an enlarged part of the spectral region (experiment and simulation) pertaining to 11 α -(CH₂)_{in} resonances. Integrals refer to the number of protons.

(Figures S7 and S8); the integrals, carried out in the ranges indicated, are in a 2:1 ratio, as expected for the inner relative to the outer resonances. Figure 4 clearly shows that for this cluster the ligand symmetry is completely removed. Table S2 gathers all ¹H and ¹³C chemical shifts. Perturbations from the simple pattern exhibited by the undoped cluster (Figure 3a) are seen for all resonances and are especially evident for the inner ligands and the protons nearer to the cluster core. It is noteworthy that signal complexity is observed even for the δ -(CH₃)_{out} proton resonance, which, being the most distant from the core, in all gold nanocluster previously investigated³ has always been the least sensitive to core size, charge, magnetic state, and environmental effects. The α -(CH₂)_{in} and α -(CH₂)_{out} resonances, which are at 4.07-3.78 and 3.15-3.05 ppm, respectively, exhibit particularly complex patterns. In particular, one of the α-(CH₂)_{in} triplets (with an integral value corresponding to one ligand) is clearly separated from the others. It is also worth noticing that this lack of symmetry does not induce diastereotopic effects in the ligands, as opposed to what found for achiral ligands in the presence of interligand interactions and/or when the staple arrangement is chiral $(Au_{38}(SR)_{24}^{0})$ and $Au_{144}(SR)_{60}^{0}$. Regarding the complexity of the proton signals for both the inner and outer ligands (see below), a quite similar behavior is also exhibited by the corresponding ¹³C resonances and ¹³C chemical shift values (HMQC experiments, Figure S9). In particular, for one of α-(CH₂)_{in} carbons, which corresponds to the isolated ¹H triplet at 4.055 ppm, the ¹³C chemical shift value is distinctly smaller (37.25 ppm) than the similar values shown by the other 11 ligands (39.01-38.79 ppm). A few differences are also detected for the β -(CH₂)_{in}, γ -(CH₂)_{in}, and δ -(CH₃)_{in} ¹³C resonances. Small differences are also present in the α- $(CH_2)_{out}$ and γ - $(CH_2)_{out}$ resonances, whereas β - $(CH_2)_{out}$ and δ -(CH₃)_{out} appear isochronous.

Regarding the number of different inner ligands detectable in the 1H NMR spectrum, we took advantage of the net separation of the α -(CH₂)_{in} triplet at 4.055 ppm to simulate the cumulative signal pertaining to the remaining 11 inner

resonances. By using the intensity of the isolated triplet as the starting point, we generated the convoluted signal by assigning and optimizing the chemical shifts of the 11 triplets. The indent in Figure 4 shows the satisfactory outcome of the simulation. The individual chemical shifts (the number of isochronous signals is given in parenthesis) are at 3.883 (1), 3.862 (2), 3.834 (1), 3.823 (4), 3.805 (1), 3.798 (1), and 3.794 (1) ppm.

A very similar set of results was obtained by analysis of the TOCSY spectrum (Figure S8): 3.883 (1), 3.854 (2), 3.818 (3), 3.808 (3), 3.786 (2) ppm. Despite small differences between the two methods, these results show that the perturbation caused by replacing one single Au atom with Hg generates at least 6 subgroups in the α -(CH₂)_{in} resonances. Regarding the α -(CH₂)_{out} triplets, analysis of the TOCSY spectrum shows that the complex signal at ca. 3.1 ppm is composed by 6 distinguishable triplets: 3.147 (1), 3.134 (1), 3.129 (1), 3.109 (1), 3.087 (1), and 3.053 (1) ppm. It is noteworthy that the effect of Hg is so strong that even all other inner and outer resonance types are affected. The ¹H and ¹³C data are collected in Table S2.

Overall, these results clearly show that Hg cannot be located in the center of the cluster, as previously proposed for $R = C2Ph.^{20}$ On the other hand, the above NMR data cannot conclude whether Hg is in one of the icosahedral vertexes or one of the staples.

The second sample investigated was prepared from $Hg(NO_3)_2$, according to the method described by Liao et al..²³ It should be recalled that this method was described to yield an Hg(s) doped cluster, though starting from a different ligand (R = C2Ph). The ¹H NMR spectrum (Figure S10), associated 2D spectra, and integral values of recrystallized cluster are *identical* to those just described for the first sample.

In the third synthetic method, we prepared Au₂₄Cd(SC4)₁₈⁰ (thiolate method, as described in the next section) and then reacted it with Hg(NO₃)₂ according to the protocol described for R = C2Ph by the Wu group.²¹ The reaction proceeded rapidly (<10 min) and efficiently (90% yield). Once again, the ¹H NMR spectrum of the crystalline product turned out to be *identical* to those recorded for the other two samples (Figure S10). The spectrum of this specific sample was also tested for stability and found to be perfectly reproducible after 4 weeks at 10 °C.

It is finally noteworthy that all three samples gave the same fragmentation pattern in MALDI-TOF mass spectrometry (Figure S6) and identical UV-vis absorption spectra (Figure 2). Their electrochemical behavior will be discussed later.

As NMR is an extremely sensitive tool to detect even minor differences in molecular properties and chemical environment effects,³ the results obtained for the three samples allow us to conclude that: (i) the specific synthetic approach, including the indirect method, does not yield clusters doped at different positions; (ii) mercury does not dope the cluster at the central position. At this stage, whether the three identical samples consist of Hg(s) or Hg(i) remains to be understood.

 $Au_{24}Cd(SC4)_{18}^0$. The cadmium-doped clusters were prepared in three ways. The first two methods are based on the use of (i) Cd(SR)₂, obtained upon reaction of Cd(NO₃)₂ with the thiol (Wang et al.),²⁰ or (ii) Cd(NO₃)₂ (Yao et al.).²¹. The third approach consists in using CdCl₂ or ¹¹³CdCl₂ (instead of Cd(NO₃)₂) to make the thiolate. Figure 5a shows the typical

 1 H NMR spectrum of the purified, recrystallized cluster obtained according to the Cd(NO₃)₂ method. Regardless of the synthetic procedure, however, the 1 H NMR spectra are *identical* (Figure S11). As for the Hg case, the 1 H NMR spectra show that the ligand symmetry is removed. The various signal types (position along the ligand chain and ligand type) were attributed through TOCSY analysis. The α-(CH₂)_{in} and α-(CH₂)_{out} resonances at 3.93-3.67 and 3.16-3.03 ppm, respectively, exhibit a complex pattern qualitatively similar to that of the Hg-doped clusters. Main differences are: (i) one of the α-(CH₂)_{in} is separated from the others but is seen at higher fields (at 3.716 ppm); (ii) the separation of one of the α-(CH₂)_{out} resonances (at 3.052 ppm) is more evident than for the Hg case. The integrals (Figure 5a) of the various inner and outer resonances are also in a 2:1 ratio.

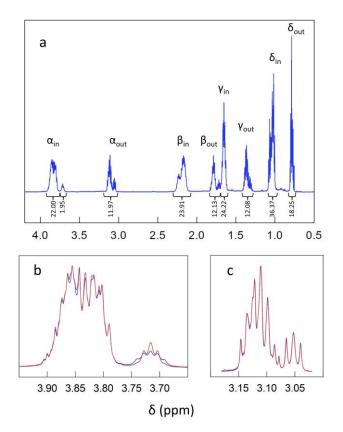


Figure 5. (a) ¹H-NMR spectrum of 2.1 mM $Au_{24}Cd(SC4)_{18}^{0}$ prepared from Cd(NO₃)₂ in C₆D₆ at 25 °C. Integrals refer to the number of protons. Graphs (b) and (c) refer to the α-(CH₂)_{in} and α-(CH₂)_{out} regions, respectively, for 2.1 mM $Au_{24}Cd(SC4)_{18}^{0}$ (red) and 2.1 mM $Au_{24}^{113}Cd(SC4)_{18}^{0}$ (blue); the latter was prepared using the thiolate obtained from ¹¹³CdCl₂.

The 1H and ^{13}C data (Table S3) show very similar patterns as observed for the Hg-doped clusters. According to the TOCSY spectrum, the $12~\alpha$ -(CH₂)_{in} triplets are at 3.865 (2), 3.834 (6), 3.826 (2), 3.803 (1), and 3.716 (1) ppm, whereas the 6 α -(CH₂)_{out} triplets are at 3.126 (1), 3.122 (1), 3.108 (3), and 3.052 (1) ppm. Differences are also seen along the ligand chain for both inner and outer ligands. Regarding ^{13}C , some differences are seen for α -(CH₂)_{in} (whereas 11 ligands are at 38.2 \pm 0.2 ppm, the isolated ligand in at 34.15 ppm), β -(CH₂)_{in}, δ -(CH₃)_{in}, α -(CH₂)_{out}, and β -(CH₂)_{out}; for γ (CH₂)_{in}, γ -

 $(CH_2)_{out}$, and δ - $(CH_3)_{out}$, differences are undetectable or within experimental error.

We also note that the NMR pattern of the Cd-doped clusters does not show any diastereotopic effect, *i.e.*, the protons of each CH₂ in each ligand type and ligand subgroup are equivalent (as already noted for the Hg-doped cluster). It is finally worth stressing that all three samples gave the same fragmentation pattern in MALDI-TOF mass spectrometry (Figure S12) and identical UV-vis absorption spectra (Figure 2). The electrochemical behavior will be discussed later.

To conclude, analysis of the three Au₂₄Cd(SC4)₁₈⁰ samples shows that: (i) independently of the synthetic method doping always occurs at the same position and (ii) this position is not at the center of the cluster.

The question now is: is Cd on the icosahedron, as inferred by Wu and co-workers for SC2Ph,²¹ or in one of the staples? To address this problem, we resorted to carry out the Cd(SR)₂ synthesis by starting from 113CdCl₂. Whereas the natural abundance of ¹¹³Cd in Cd samples is 12.23%, enriched ¹¹³Cd samples contain 95% of this spin 1/2 isotope. A point-by-point comparison between the ¹H NMR spectra of Au₂₄¹¹³Cd(SC4)₁₈⁰ and Au₂₄Cd(SC4)₁₈⁰ (both obtained from the thiolate method) shows (Figure S13) that the effect of the isotopic enrichment is observed only for the signal at 3.716 ppm, while the rest of the spectrum is completely unchanged. Figure 5b shows a detail of the only change detected. In particular, the small bumps around the isolated α-(CH₂)_{in} triplet of Au₂₄Cd(SC4)₁₈⁰ are significantly enhanced in Au₂₄¹¹³Cd(SC4)₁₈⁰, in agreement with the ca. 8-fold isotopic enrichment. Conversely, the isolated and the other 5 convoluted α -(CH₂)_{out} triplets are identical (Figure 5c).

¹H-¹H homodecoupling experiments were carried out by applying a standard pulse sequence. Decoupling was performed at the frequency of the β-(CH₂)_{in} signal (2.173 ppm) that exhibits a scalar correlation with the isolated α-(CH₂)_{in} signal at 3.716 ppm. Figure 6 shows the main details of the effects observed for the Au₂₄¹¹³Cd(SC4)₁₈⁰ and Au₂₄Cd(SC4)₁₈⁰ samples. As expected on the basis of the ¹H-113Cd coupling, whereas the enriched cluster shows a doublet (Figures 6a), the latter yields a singlet (Figure 6b), though accompanied by traces of a doublet (due to the presence of ¹¹³Cd and ¹¹¹Cd, which is another spin 1/2 isotope with a natural abundance of 12.80%). The doublet in Figure 6a allows determining a ³J(¹H-¹¹³Cd) coupling constant of 14.3(0.1) Hz. For both $Au_{24}^{113}Cd(SC4)_{18}^{0}$ and $Au_{24}Cd(SC4)_{18}^{0}$, the same pulse sequence applied to the isolated β-(CH₂)_{out} signal (1.719 ppm), which correlates with the corresponding isolated α-(CH₂)_{out} signal at 3.052 ppm, transforms the latter into a sharp singlet, as could be anticipated on the basis of the uncomplicated shape of this triplet (Figures 6c,d). The slightly different position of the peaks in the decoupled spectra is due to the Bloch-Siegert shift, which causes resonances to move away from the decoupling frequency.

Figure 7a shows two models of the possible positions occupied by Cd. For Cd(s), the bond sequence from Cd to the α -(CH₂)_{in} and α -(CH₂)_{out} protons is the same, H-C-S-Cd, whereas for Cd(i) the distance from the α -(CH₂)_{out} protons is larger by two bonds (H-C-S-Au-S-Cd bond sequence) than for the α -(CH₂)_{in} protons. It is thus conceivable that for Cd(i) doping only the α -(CH₂)_{in} protons are affected, as the experiments indeed indicate. On the other hand, if the

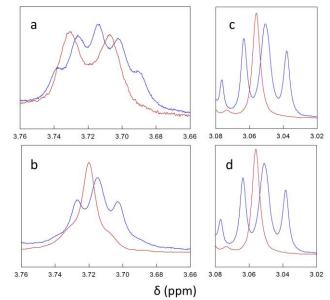


Figure 6. ¹H NMR spectra of 2.1 mM $Au_{24}^{113}Cd(SC4)_{18}^{0}$ (a,c) and 2.1 mM $Au_{24}Cd(SC4)_{18}$ (b,d) focusing on the α-(CH₂)_{in} (a,b) and α-(CH₂)_{out} (c,d) regions. The spectra are shown before (blue traces) and after ¹H-¹H homodecoupling (red traces). C₆D₆, 25 °C.

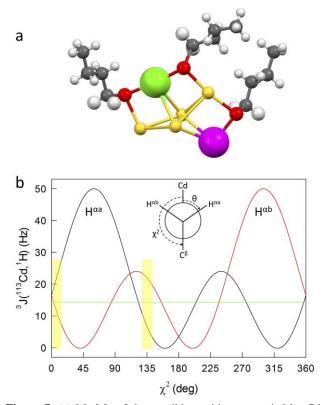


Figure 7. (a) Models of the possible positions occupied by Cd. (b) Karplus-like correlation of the ${}^3J({}^1\mathrm{H}^{-113}\mathrm{Cd})$ coupling constant as a function of the dihedral angle χ^2 . The Newman projection of the investigated bond sequence illustrates the relationship between the χ^2 and θ dihedral angles. The areas highlighted in yellow show the regions where the average ${}^3J({}^1\mathrm{H}^{-113}\mathrm{Cd})$ coupling constant values are compatible with the experimentally determined J value.

exchange yields Cd(s), 113 Cd would have affected *both* the α -(CH₂)_{in} and α -(CH₂)_{out} protons with equal probability, which is in contrast to the experimental outcome.

To gain more quantitative insights into this aspect, we propose the use of a Karplus-type correlation, which was originally developed to describe the dihedral angle dependence of three bond ¹H-¹H coupling.⁴³ In 1994, Vašák and co-workers could demonstrate that a Karplus-type correlation describes nicely also the dihedral angle dependence of the three bond 113Cd-1H coupling, as obtained from HMQC data for Cd-substituted metalloproteins in comparison with the crystal structure data. 44,45 The correlation was observed for the cysteine H-C-S-Cd dihedral angle. The same group could previously demonstrate that the Cdderivative is isostructural with the native protein. 46 Vašák and co-workers concluded that although heteronuclear couplings involving heavy nuclei generally depend on orbital angular momentum, electron-nucleus dipole-dipole interaction, and contact contributions, for Cd-substituted metalloproteins the dihedral angle is the principal determinant of the Fermi contact term and the dominant variable. Figure 7b, which is adapted from the original work, 44,45,47 illustrates the dependence of the ${}^{3}J({}^{1}H-{}^{113}Cd)$ coupling constant on the dihedral angle θ , ${}^{3}J({}^{1}H-{}^{113}Cd) = 36 (\cos^{2}\theta) -13 (\cos\theta) +1$, with $r^2 = 98.7\%$ and confidence limits of ca. 10 Hz; tetrahedral geometry around C^{β} is assumed. More precisely, Figure 7b shows the correlation as a function of χ^2 , which refers to the dihedral angle with respect to the β carbon atom. The inset to Figure 7b shows the Newman projection of the bond sequence and defines the relationship between the dihedral angles θ and χ^2 , where θ relates to H^a (one of the two α-(CH₂)_{in} protons). The correlation shows that a coupling constant should be detected no matter the magnitude of θ or χ^2 , as at least one of the two protons (H^a and H^b) always provides a finite coupling value. Regarding our Cd-doped clusters, the Karplus-type correlation thus confirms that for the hypothetical Cd(s) some coupling with the α -(CH₂)_{out} protons should be observed as well as with the α-(CH₂)_{in} protons. This is not seen.

Let us now focus on the only resonance α -(CH₂)_{in} affected by the presence of 113 Cd. Figure 7b shows that very few χ^2 regions (in yellow; for symmetry, only the range from 0 to 180° needs to be considered), which determine the $^3J(^1\text{H-}^{113}\text{Cd})$ coupling constant values, provide average $^3J(^1\text{H-}^{113}\text{Cd})$ coupling constant values compatible with the experimentally determined J value of 14.3 Hz, at least within a prudential uncertainty of ca. ± 3 Hz: 0-13° ($J = 16.5 \div 17.2$ Hz) and 128-143° ($J = 17.2 \div 11.3$ Hz). For steric reasons, however, the latter is the only plausible region. The reliability of this conclusion will be addressed for Au₂₄Cd(SC2Ph)₁₈°.

Au₂₄Cd(SC2Ph)₁₈⁰ and Au₂₄Hg(SC2Ph)₁₈⁰. The results and conclusions so far reached regard the butanethiolateprotected clusters and thus the question arises as to whether they are extendable to other ligands, also considering that the seminal works carried out by the groups of Zhu and Wu on Cd- and Hg-doping focused on the phenylethanethiolate ligand. Indeed, since Donkers et al. described the first synthesis isolation and of Au_{25} protected phenylethanethiolate ligands⁴⁸ (Note: this cluster originally believed to be Au₃₈(SC2Ph)₂₄), followed some years later by the actual crystallographic structure determination of $[Au_{25}(SC2Ph)_{18}](n-Oct_4N)^+,^{29,30}$ phenylethanethiolate has been adopted by many research groups as sort of a reference ligand. We thus studied the Cd- and Hg-doping of [Au₂₅(SC2Ph)₁₈](*n*-Oct₄N)⁺ according to the same sequence of reactions and tests already described for the SC4 ligand.

Au₂₄Cd(SC2Ph)₁₈⁰ was prepared according to the Cd(SC2Ph)₂,²⁰ Cd(NO₃)₂,²¹ and the CdCl₂ (or ¹¹³CdCl₂) - thiolate methods. As for the SC4 ligand, the corresponding MALDI-TOF (Figure S14) and UV-vis absorption spectroscopy spectra (Figure S1) show no differences. The ¹H NMR spectra of the Au₂₄Cd(SC2Ph)₁₈⁰ sample obtained from Cd(SC2Ph)₂ (blue) and Au₂₄¹¹³Cd(SC2Ph)₁₈ (red) are shown in Figure 8. The perfect overlap of the spectra in Figure 8 (but for the feature magnified in the inset, as discussed below) and Figure S16, which pertain to the three samples, confirms that preparing this doped cluster with the thiolate,²⁰ the salt,²¹ or the CdCl₂ - thiolate method produces the very same result. A previously reported spectrum (CD₂Cl₂, Cd(SR)₂ synthesis) shows similar features (Figure S7 in reference 20), whereas no NMR data were provided in the other report on Cd doping.²¹

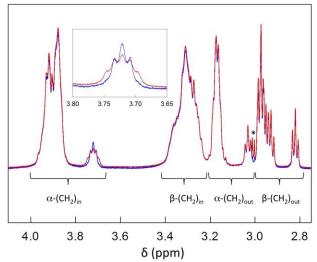


Figure 8. Full overlap of the ^{1}H NMR spectra of 2.2 mM Au₂₄Cd(SC2Ph)₁₈0 (blue) and 2.2 mM Au₂₄¹¹³Cd(SC2Ph)₁₈0 (red) in C₆D₆ at 25 $^{\circ}$ C (region of the aliphatic C-H signals). The inset highlights the effect of isotopic enrichment on the isolated α-(CH₂)_{in} resonance. The star marks a solvent impurity (methanol).

The spectrum shows that the ligand symmetry is disrupted, which, once again, is inconsistent with Cd(c) doping.²⁰ The COSY spectrum (Figure S15) allows attributing (Table S4) the complex signal at 4.0-3.8 ppm (integral corresponding to 11 ligands) to 22 α-(CH₂)_{in} protons, the slightly distorted triplet at 3.722 ppm to the twelfth α-(CH₂)_{in} ligand, and the multiplet at 3.43-3.21 ppm to the 24 β -(CH₂)_{in} protons. The α -(CH₂)_{in} signal at 3.722 ppm correlates with the β-(CH₂)_{in} triplet at 3.287 ppm. The resonances corresponding to the 6 outer ligands appear as a complex multiplet of α-(CH₂)_{out} resonances centered at 3.17 (10 protons) followed by one additional triplet at 3.031 ppm (2 protons), and a series of largely overlapped β-(CH₂)_{out} triplets at 3.0-2.9 ppm (10 protons, five ligands) followed by one additional triplet (2 protons) at 2.820 ppm. The α -(CH₂)_{out} and β -(CH₂)_{out} triplets at 3.031 and 2.820 ppm correlate and thus belong to the same ligand. Careful analysis of the COSY spectrum, which was acquired during a particularly long time frame, allows distinguishing 12 nonisochronous inner ligands and 6 outer ligands (Table S4), which confirms on a quantitative basis the profound effect of Cd-doping on the ligand symmetry. Regarding the ¹³C resonances, the values are pretty much isochronous (for the same position and ligand type), but for small differences for the isolated inner and outer signals.

The observation of an isolated triplet for both the inner and outer ligands is compatible with both Cd(i) and, intuitively, even more for Cd(s), as in this case both the α -(CH₂)_{out} and β -(CH₂)_{out} resonances are significantly affected. Can this be taken as the proof that the staple position is preferable? To clarify the position of the Cd atom, we followed the same procedure used for SC4. First, we checked that the reaction with CdCl₂ proceeds smoothly and then used ¹¹³CdCl₂ to prepare Au₂₄¹¹³Cd(SC2Ph)₁₈⁰. Figure 8 shows that only the isolated α -(CH₂)_{in} is affected by ¹¹³Cd, while the rest of the ¹H NMR spectrum is perfectly superimposable to that of Au₂₄Cd(SC2Ph)₁₈⁰. The inset of Figure 8 shows the detail of this effect. Decoupling was carried out for both $Au_{24}^{113}Cd(SC2Ph)_{18}^{0}$ and $Au_{24}Cd(SC2Ph)_{18}^{0}$, and the effects on both α -(CH₂)_{in} and α -(CH₂)_{out} were tested. As expected on the basis of the ¹H NMR spectra, the only effect is on the isolated α-(CH₂)_{in} resonance (Figure S17). Not only the effect is qualitatively the same as described for SC4, but also the ³J(¹H-¹¹³Cd) coupling constant is quite similar, 13.6(0.2) Hz. As observed for Au₂₄Cd(SC4)₁₈⁰ (Figure 6), a trace of the α-(CH₂)_{in} doublet obtained upon decoupling could be detected also in the Au₂₄Cd(SC2Ph)₁₈⁰ sample. These results confirm that the Cd dopant is on the icosahedron, as demonstrated above for the C4 cluster and originally suggested by Wu and co-workers for the C2Ph cluster.2

According to the correlation between the ${}^{3}J({}^{1}H-{}^{113}Cd)$ coupling constant and the H-C-S-Cd dihedral angle, 43,44 the virtually identical coupling constants determined experimentally for the SC4 and SC2Ph ligands point to very similar H-C-S-Cd dihedral angles. As described later, we could obtain the structures of both Au₂₄Cd(SC4)₁₈⁰ and Au₂₄Cd(SC2Ph)₁₈⁰ by single-crystal X-ray crystallography. Whereas the former is affected by intercluster interactions, the latter refers to unbounded clusters. Au₂₄Cd(SC2Ph)₁₈⁰ thus provides an ideal case to test on quantitative grounds the validity of the ³J(¹H-¹¹³Cd) coupling constant correlation, which was originally described for metalloproteins, also for gold nanoclusters. According to the correlation, the experimentally determined J value corresponds (within ca. 3 Hz) to plausible angle χ^2 values of $130 \div 145^{\circ}$ ($J = 16.6 \div$ 10.6 Hz). In the structure of Au₂₄Cd(SC2Ph)₁₈⁰, we find that the average C^{β} - C^{α} -S-Cd dihedral angle χ^2 is 149°; similar values can be obtained from the structures published by Wang et al., 150°, 20 and Yao et al., 133°. 21 These figures yield an average χ^2 of 144°, which is indeed in excellent agreement with the estimated range, also considering the usual limits of comparing solid- to solution-phase results.

 $Au_{24}Hg(SC2Ph)_{18}{}^0$ was also prepared in three ways: (i) metal exchange on $Au_{25}(SC2Ph)_{18}{}^-$ with $Hg(SC2Ph)_2{}^{20}$ and $Hg(NO_3)_2,^{23}$ and metal exchange on preformed $Au_{24}Cd(SC2Ph)_{18}{}^0$ with $Hg(NO_3)_2,^{21}$ After purification and recrystallization, the three $Au_{24}Hg(SC2Ph)_{18}$ samples were characterized by UV-vis absorption spectroscopy (Figure S1), electrochemistry (see next section), and MALDI-TOF mass spectrometry, which gave the same fragmentation pattern (Figure S18).

The three $Au_{24}Hg(SC2Ph)_{18}^0$ samples exhibit *identical* 1H NMR spectra, even from the viewpoint of minor features (Figure S19 allows appreciating the perfect correspondence of the three spectra). The typical 1H NMR pattern of $Au_{24}Hg(SC2Ph)_{18}^0$ is exemplified in Figure 9a, which pertains to the sample obtained upon metal exchange on a preformed $Au_{24}Cd(SC2Ph)_{18}^0$ cluster. Please note that whereas the $Hg(NO_3)_2$ synthesis is supposed to yield Hg(s), 23 for the double exchange we purposely used $Au_{24}Cd(SC2Ph)_{18}^0$ obtained using $Cd(SR)_2$, as this sample is that supposed to produce Cd(c).

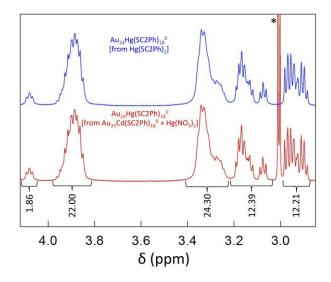


Figure 9. ¹H NMR spectrum of 2.2 mM Au₂₄Hg(SC2Ph)₁₈⁰ (aliphatic C-H signals) obtained from Hg(SC2Ph)₂ (blue trace) or after exchange on a preformed Au₂₄Cd(SC2Ph)₁₈⁰ cluster (red trace). The integrals and ligand types are indicated. The asterisk marks a solvent impurity (methanol).

The ¹H NMR spectrum exhibits four groups of peaks. Assignments were carried out through COSY measurements (Figure S20), and the corresponding ¹H and ¹³C chemical shifts are provided in Table S5. As for Au₂₄Cd(SC2Ph)₁₈⁰, the inner resonances α -(CH₂)_{in} and β -(CH₂)_{in} are clearly separated from the two outer resonances α -(CH₂)_{out} and β -(CH₂)_{out}. For both $\alpha\text{-(CH}_2)_{in}$ and $\alpha\text{-(CH}_2)_{out},$ one of the triplets is well separated from the others. As for SC4, the isolated α -(CH₂)_{in} signal is downfield with respect to the group of the remaining 11 ligands, whereas it is upfield for the outer ligands. The COSY analysis of the α -(CH₂)_{in} and α -(CH₂)_{out} resonances allowed estimating the presence of each of the 12 and 6 ligands, respectively. Nonisochronous signals are also detected for the corresponding β-(CH₂)_{in} and β-(CH₂)_{out} resonances. As to ¹³C, the signals are isochronous, with a few exceptions. Once again, the number of clearly distinguishable resonances and isochronous signals witnesses the significant loss of symmetry undergone upon Hg-doping. It is worth noticing that the ¹H NMR spectra obtained for Au₂₄Hg(SC2Ph)₁₈⁰ (Figure S16 in reference 24 and Figure S7 in reference 20) show very similar features (our data were obtained in C₆D₆, whereas those published data pertain to CD₂Cl₂), though they were not specifically discussed. Overall, the NMR analysis shows that the Hg atom cannot be at the central position of the cluster, as previously hypothesized.²⁰

As to its actual position, we argued that the fast Hg exchange on a preformed Au₂₄Cd(SC2Ph)₁₈⁰ cluster occurs on the icosahedron as well, i.e., directly on the site occupied by Cd(i), rather than involving a complicate molecular rearrangements where first Hg exchanges Cd(i) and then switches position with the nearby Au(s) atom. To gain insights into this problem, we applied the same decoupling sequence used for the Cd-doped clusters. The goal was to detect a possible ${}^{3}J({}^{1}H-{}^{199}Hg)$ coupling by relying on the fact that ¹⁹⁹Hg has a natural abundance of 16.94% and is a spin 1/2 isotope. Assuming that a Karplus-type correlation is valid also for the three-bond system H-C-S-Hg, one would expect to see some ${}^3J({}^1\text{H}-{}^{199}\text{Hg})$ coupling only for α -(CH₂)_{in} or both α -(CH₂)_{in} and α-(CH₂)_{out} for Hg(i) and Hg(s), respectively. Radiating the corresponding β-(CH₂)_{in} signal (assessed via TOCSY) transforms the α -(CH₂)_{in} signal into a singlet accompanied by a doublet (Figure S21a) that allows calculating a ${}^3J({}^1H^{-199}Hg)$ coupling constant of 36 Hz. Conversely, radiating the β-(CH₂)_{out} signal transforms the corresponding isolated α-(CH₂)_{out} signal into uncomplicated singlet (Figure S21b). We also applied the same decoupling analysis to the isolated α-(CH₂)_{in} signal of Au₂₄Hg(SC4)₁₈⁰, obtained the same outcome, and calculated the very similar value of 37 Hz (Figure 10), whereas no effect was detected for the isolated α-(CH₂)_{out} resonance. Although to the best of our knowledge a Karplus-like dependence has never been observed for ¹⁹⁹Hg, it is conceivable that a periodic dependence such as that found for ${}^3J({}^1\mathrm{H}{}^{-113}\mathrm{Cd}){}^{44,45}$ should be qualitatively valid also for of the ${}^{3}J({}^{1}H^{-199}Hg)$ coupling. The virtually identical values determined for C2Ph and C4 would thus point to very similar average dihedral angles, as determined for the Cd-doped clusters. Most important, these results provide compelling evidence that Hg-doping, whether performed directly on Au₂₅(SR)₁₈ or indirectly on $Au_{24}Cd(SR)_{18}^0$, consistently yields Hg(i), rather than Hg(s).²³

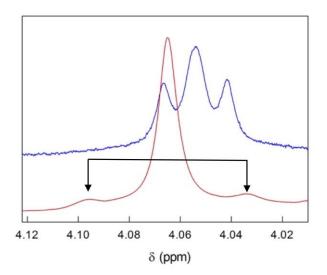


Figure 10. ¹H NMR spectra of 2.1 mM Au₂₄Hg(SC4)₁₈⁰, focusing on the isolated α-(CH₂)_{in} region before (blue) and after (red) ¹H-¹H homodecoupling at the frequency of the corresponding β-(CH₂)_{in} signal. The two spectra are vertically shifted for clarity and the arrows mark the doublet. C_6D_6 , 25 °C.

To conclude, NMR demonstrates that both Cd and Hg are exchanged on the icosahedron, no matter the synthetic method employed or the nature of the ligand. It should be also noted

that the MALDI-TOF mass spectra of the samples obtained for each doped clusters are identical (Figures S6, S12, S14, and S18). As either foreign-metal atom is on the icosahedron, differences in the fragmentation patterns observed between the Hg and Cd doped clusters should not to be taken as indicating a different doping position.²³ Rather, they just reflect the effect of the specific doping element, as also supported by the similar fragmentation patterns exhibited for the same doping metal by the SC4 and SC2Ph protected clusters.

Electrochemistry of Au₂₄**M**(**SR**)₁₈. The electrochemical measurements were carried out in dichloromethane (DCM) containing 0.1 M tetrabutylammonium hexafluorophosphate (TBAH), using a glassy carbon (GC) microdisk electrode. Figure 11a compares the DPV behavior of $Au_{24}M(SC4)_{18}$ for M = Au, Pt, Cd, and Hg. Figure 11b shows the DPV behavior of the $Au_{24}M(SC2Ph)_{18}$ samples (M = Au, Cd, Hg). As expected, for both the Hg- and Cd-doped clusters the various samples exhibit exactly the same DPV pattern and formal potential (E°) values. This is exemplified for both ligands in Figure S22, which shows the DPVs of the Hg samples obtained with the Hg(NO₃)₂, thiolate, and Cd exchange methods.

The DPVs of $Au_{24}Hg(SC4)_{18}^{0}$ and $Au_{24}Cd(SC4)_{18}^{0}$ are qualitatively similar to that of Au₂₅(SC4)₁₈. The doped clusters undergo two successive one-electron oxidations (E_1°) and E°_{2}) at 0.364 and 0.684 V, $Au_{24}Hg(SC4)_{18}^{0}$, and 0.332 and 0.636 V, Au₂₄Cd(SC4)₁₈⁰. In the timescale of voltammetry experiments, both processes are reversible. Further oxidation processes are detectable at more positive potentials, though with formation of chemically labile species. The E° for the first peak of these doped clusters is more positive than that of $\text{Au}_{25}(\text{SC4})_{18}^-$ ($E^{\circ}_1 = -0.188$, $E^{\circ}_2 = 0.139$ V, respectively)⁴⁹ by 0.552 and 0.520 V, respectively. For $Au_{24}Hg(SC2Ph)_{18}{}^{0}$ (E°_{1} = 0.451, $E_{2}^{\circ} = 0.703 \text{ V}$) and $\text{Au}_{24}\text{Cd}(\text{SC2Ph})_{18}^{0}$ ($E_{1}^{\circ} = 0.430$, E_{2}° = 0.668 V) similar considerations apply. With respect to $\text{Au}_{24}(\text{SC2Ph})_{18}^-$ ($E^{\circ}_{1} = -0.077$, $E^{\circ}_{2} = 0.226 \text{ V}$), 50 the positive shifts of E_1° are 0.528 and 0.507 V, respectively. For both Hg and Cd, this remarkable positive shift was already observed.23,51

Regarding the first reduction peak, which $Au_{24}Hg(SC4)_{18}^{0}$ and $Au_{24}Cd(SC4)_{18}^{0}$ occurs at -1.23 and -1.39 V, respectively, the formation of the anion is chemically irreversible. For the latter, increasing the CV potential scan rate (v) allows to detect reversibility, and therefore, determine an E° value of -1.38 V. We described this procedure in detail for a series of Au₂₅(SR)₁₈ clusters.^{50,52,53} The electrochemical gap of $Au_{24}Cd(SC4)_{18}^{0}$ can thus be calculated from the E° difference between the +1/0 and 0/-1 redox couples. The corresponding HOMO-LUMO gap can then be estimated by subtracting the charging-energy contribution, obtained from the E° difference between the +2/+1 and +1/0 states.⁵⁴ The value so-obtained, 1.41 eV, is in very good agreement with the HOMO-LUMO gap of 1.37 eV estimated from the onset of optical absorption (Figure 2). For Au₂₄Cd(SC2Ph)₁₈⁰ we observed the same behavior, and this allows calculating an E° value of -1.26 V for the 0/-1 redox couple. The HOMO-LUMO gap is thus estimated to be 1.46 eV, to be compared with that obtained from the optical spectrum (Figure S1), 1.41 eV and the value of 1.4(0.1) eV obtained by time-resolved spectroscopic analysis.²²

For both SC4 and SC2Ph ligands, the analysis of the reduction of the Hg-doped clusters is more complicated

because the voltammetric peak exhibits features that suggest interaction with the electrode surface. Furthermore, for both Hg-doped clusters the peak is irreversible also at high ν values (up to 50 V s⁻¹). However, at low temperature (-45 °C) and high ν some reversibility is detectable, which allows estimating E° . By comparing this result with the corresponding E° determined at the same temperature for Au₂₅(SC2Ph)₁₈⁻, the E° values of the two Au₂₄Hg(SR)₁₈⁰ clusters at 25 °C could be estimated. Calculation of the HOMO-LUMO gap yields 1.28 (SC4) and 1.29 (SC2Ph),

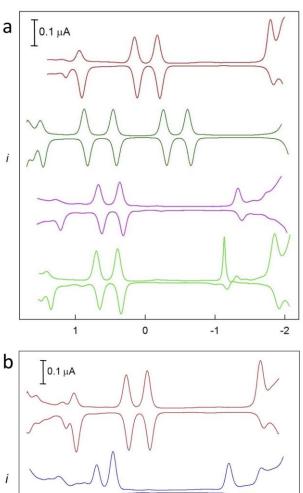


Figure 11. Comparison between the DPV curves for (top to bottom): (a) $Au_{25}(SC4)_{18}^-$, $Au_{24}Pt(SC4)_{18}^0$, $Au_{24}Cd(SC4)_{18}^0$, and $Au_{24}Hg(SC4)_{18}^0$; (b) $Au_{25}(SC2Ph)_{18}^-$, $Au_{24}Cd(SC2Ph)_{18}^0$, and $Au_{24}Hg(SC2Ph)_{18}^0$. Glassy-carbon electrode, DCM/0.1 M TBAH, 25 °C.

respectively; these gaps are thus slightly smaller than those electrochemically determined for $Au_{25}(SC4)_{18}^-$ and $Au_{25}(SC2Ph)_{18}^-$, 1.30 and 1.34 eV, respectively. $^{49.55}$ A very recent time-resolved spectroscopy analysis led for $Au_{24}Hg(SC2Ph)_{18}^0$ and $Au_{25}(SC2Ph)_{18}^-$ to the similar values of 1.2(0.1) and 1.3(0.1) eV, respectively. 22 Very recent calculations provided a similar decrease in the HOMO-LUMO gap energy on going from Cd to Hg, as well as valuable insights into the electronic effects introduced by dopants. 36

The DPV of $Au_{24}Pt(SC4)_{18}^0$ clearly points to a different orbital-energy distribution. It shows two pairs of peaks corresponding to the formation of the mono- and dication on the positive-going scan (E° of 0.475 and 0.853 V), and the mono- and dianion on the negative-going scan (E° of -0.287 and -0.622 V). Each of these charge states is chemically stable. For this cluster, a HOMO-LUMO gap of 0.384 eV can be estimated from the electrochemical data (for the charging energy correction, we used the potential difference calculated for the two oxidation peaks, 0.378 V) is much lower than for the other clusters investigated. A previous electrochemical analysis carried out on $Au_{24}Pt(SC6)_{18}^0$ yielded the similar value of 0.34 eV.¹⁴ With SC4, however, we fail to detect the large potential difference reported for the E°_1 of $Au_{25}(SC6)_{18}^-$ and the first reduction peak of $Au_{24}Pt(SC6)_{18}^{0.14}$

Overall, some of our electrochemical data essentially confirm previous electrochemical conclusions on the position occupied by Pt upon Au_{25} -doping 14 and the effect of Hgdoping. 23 Most important, however, they provide further compelling evidence that Hg- and Cd-doping always occur on the same metal site, no matter the ligand and how metal exchange is carried out.

Single Crystal X-Ray Crystallography. We could solve the structure of most of the clusters, sometimes also as the result of different syntheses and in two laboratories, as specified: Au₂₄Pt(SC4)₁₈⁰ (Padova), Au₂₄Hg(SC4)₁₈⁰ (from Hg(SC4)₂, Jyväskylä), Au₂₄Cd(SC4)₁₈⁰ (from Cd(SC4)₂, Jyväskylä), Au₂₄Hg(SC2Ph)₁₈⁰ (from Hg(NO₃)₂, Padova and Jyväskylä), Au₂₄Hg(SC2Ph)₁₈⁰ (from Hg(NO₃)₂ + Au₂₄Cd(SC2Ph)₁₈⁰, Padova and Jyväskylä), and Au₂₄Cd(SC2Ph)₁₈⁰ (from Cd(SC4)₂, Jyväskylä). Here we will focus on the most salient aspects, whereas full discussion on these results is provided in the Supporting Information.

For both Au₂₄Hg(SC4)₁₈⁰ and Au₂₄Cd(SC4)₁₈⁰, the structure shows the very same features discovered for Au₂₅(SC4)₁₈⁰ (this cluster is a neutral radical)³⁷ and Au₂₅(SC5)₁₈^{0,39} (i) the clusters form linear polymers of interconnected clusters; (ii) the connecting staples form S-Au-Au-S dihedral angles of nearly 90° (for both Hg- and Cd-doping, 81-85°); (iii) the neighboring clusters are connected via aurophilic Au-Au bonds. Formation of the polymers is thus granted by a twistand-lock mechanism³⁷ in which the orientation of the alkyl chains and their van der Waals interaction opens up two opposite sides of the Au-S-Au staples and favor a closer approach between neighboring clusters, thereby causing formation of an intercluster Au-Au aurophilic bond. In the doped clusters, this bond has a similar length, 3.09 Å (Hgdoping) and 3.10 Å (Cd-doping), as found in Au₂₅(SC4)₁₈⁰ $(3.15 \text{ Å})^{37}$ and $\text{Au}_{25}(\text{SC5})_{18}{}^{0} (2.98 \div 3.03 \text{ Å}).^{39}$ These results thus point to the importance of the alkanethiolate ligand, and show that formation of the intercluster aurophilic bond is possible regardless of the magnetic state of the cluster: in fact, as opposed to $Au_{25}(SC4)_{18}^0$, both the Hg- and Cd-doped clusters are diamagnetic, as evinced from the NMR results and previously shown for $Au_{24}Hg(SC2Ph)_{18}^0$ by electron paramagnetic resonance.²³

With that being said, we note that Au₂₄Pt(SC4)₁₈⁰, which was purposely prepared with the SC4 ligand, does not form polymers. Its structure does not show any rotation of the staples, which remains virtually parallel, and shows a relatively large minimum intercluster Au-Au distance of 3.88 Å. This is as previously observed for, say, Au₂₅(SC2)₁₈⁰, which shows a minimum intercluster Au-Au distance of 4.12 Å,⁵⁴ and Au₂₅(SC3)₁₈⁰.³⁸ Overall, this may suggest that electronic factors may also play a role in determining the different behavior observed for Au₂₄Pt(SC4)₁₈⁰.

Crystallographic analysis alone is not distinctive enough for determining the positions of these doping metals with reliable accuracy. This is due to the very small electron-density differences between Au and the Pt, Hg, and Cd metals, especially for Pt and Hg that differ from Au by only one electron. Thus, an Au site substituted by Hg or Pt should show an electron density higher or lower than that of Au by only 1.2%, respectively. This figure will be significantly lowered if the doping metal is disordered over two or more locations (~0.1% difference when all 12 icosahedral sites are partially but evenly occupied; even less if distribution also involves the staples) and/or if the quality of the crystal is less than ideal. This implies that for these clusters small differences in electron densities cannot be determined reliably even with the highest quality data obtained by the modern in house diffractometers. However, thorough analysis of the structure of the Cd doped clusters (the electron-density difference between Cd and Au is ~60%) using several data sets with the highest possible data quality (data redundancy of 5) allowed us to refine the structure of the Cd-doped cluster quite satisfactorily. Consistently with the NMR analysis, refinement indicated that Cd is most likely disorderly located on the icosahedral sites instead of the center or staples. For the Hg and Pt doped Au₂₄M(SC4)₁₈⁰ clusters, on the other hand, the electron-density difference is just too small to draw similar conclusions.

Regarding the SC2Ph-protected clusters, Au₂₄Hg(SC2Ph)₁₈⁰ and Au₂₄Cd(SC2Ph)₁₈⁰ show exactly the same structure, where the orientation of the ligands with respect to plane of the staple is always of the up-down-up type (Figure 12). This is, therefore, identical to the ligand orientation seen in Au₂₅(SC2Ph)₁₈⁰, which is always of the up-down-down type. This is, which is always of the up-down-down type. This is, we checked the two structures of Au₂₄Hg(SC2Ph)₁₈⁰ obtained upon metal exchange in either Au₂₅(SC2Ph)₁₈⁻ or Au₂₄Cd(SC2Ph)₁₈⁰, and found they are identical. Further discussion on the SC4- and SC2Ph-protected clusters is provided in the Supporting Information.

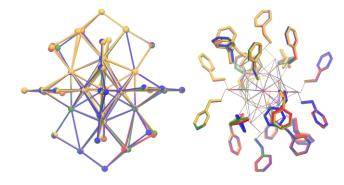


Figure 12. Overlap of the structures of the Hg- and Cd-doped clusters with that of $Au_{25}(SC2Ph)_{18}^{0.39}$ The Au/M-S core units are shown on the left-hand side (ligands removed), whereas the full clusters are shown on the right-hand side (metal core units are faded, for clarity) to evidence full overlapping of the ligands (mixed colors). The color codes are: red = Hg-doped cluster (from $Hg(NO_3)_2$), blue = Hg-doped cluster (from $Hg(SC2Ph)_2$), green = Cd-doped cluster, and yellow = $Au_{25}(SC2Ph)_{18}$.

CONCLUSIONS

This study was meant to obtain insights into the monodoping of Au₂₅(SR)₁₈ clusters with foreign metal atoms. Accurate selection of chemicals, NMR analysis, electrochemical data, and critical analysis of crystallographic data allowed us to highlight hindsights into the challenge of understanding where the foreign-metal atoms are eventually located in the cluster structure, and how to characterize these quite elusive nanosystems. Using the NMR results obtained for $Au_{24}Pt(SC4)_{18}^{0}$ and $Au_{24}Pd(SC4)_{18}^{0}$ as a reference for the behavior expected when the cluster is doped in its central position, we show that Cd- and Hg-doping does not occur at the central position.²⁰ We also show that the Cd-doping mode is not different from that of Hg-doping, as opposed to what previously concluded. 21,23 Rather, we find that both Cd- and Hg-doping occurs in one of the icosahedral positions, independently of the specific ligand. Equally important, we demonstrate that the metal-exchange doping methods so-far developed always yield the very same species. Besides being important from a fundamental viewpoint, these results are also liable to impact applications of doped clusters, e.g., in catalysis, which is a promising growing area of research for atomically precise metal clusters.⁵⁶ This is because proper understanding of the doping site affects the analysis of the catalytic mechanism.⁵¹ Finally, we provide a warning about reaching conclusions on the doping site on the basis of different fragmentation patterns in mass spectra, and especially, single-crystal X-ray crystallography results. We also hope that these new insights will be useful for theoreticians as a sound experimental basis to refine their calculation models.

EXPERIMENTAL SECTION

The $Au_{25}(SC4)_{18}^-$ and $Au_{25}(SC2Ph)_{18}^-$ clusters were prepared and purified as already described. ^{37,42} Full details on chemicals and the preparation of $Au_{24}Pt(SC4)_{18}^0$, $Au_{24}Pd(SC4)_{18}^0$, $Au_{24}Hg(SC4)_{18}^0$, $Au_{24}Hg(SC2Ph)_{18}^0$, and $Au_{24}Cd(SC2Ph)_{18}^0$ are described in the Supporting Information.

The UV-vis absorption spectra of the clusters were obtained in DCM with a Thermo Scientific Evolution 60S spectrophotometer. The spectra were recorded with a spectral

resolution of 0.5 nm. The samples were at 0.2 mM concentration in 1 mm cuvettes. MALDI-TOF mass spectrometry experiments were carried out with an Applied Biosystems 4800 MALDI-TOF/TOF spectrometer equipped with a Nd:YAG laser operating at 355 nm. The laser-firing rate was 200 Hz and the accelerating voltage was 25 kV. trans-2-[3-(4-tert-butylphenyl)-2-methyl-2-propenylidene] malononitrile (DCTB) was used as the matrix. Depending on the experiment, the instrument was calibrated with $Au_{25}(SC4)_{25}{}^0$ or $Au_{25}(SC2Ph)_{25}{}^0$. The clusters were dissolved in DCM containing DCTB to obtain 0.1 mM solutions with a 1:400 nanocluster/matrix ratio. A 5 μ l solution was drop cast onto the sample plate and air-dried. All spectra were recorded using the reflector positive-ion mode.

The electrochemical experiments were carried out under an Ar atmosphere, in a glass cell at room temperature, unless otherwise stated. The solvent-electrolyte system was DCM containing 0.1 M TBAH. The working electrode was a glassy carbon microdisk (9.1 × 10⁻⁴ cm²), prepared and activated as already described.⁵⁷ As a quasi-reference electrode, we used a silver wire, which was kept in a tube filled with the same electrolyte solution and separated from the main compartment by a Vycor frit. Its calibration was performed by addition of ferrocene at the end of the experiments; in DCM/0.1 M TBAH, the ferricenium/ferrocene redox couple has $E^{\circ} = 0.460$ V against the KCl saturated calomel electrode (SCE). All potential values are reported against SCE. The counterelectrode was a Pt wire. We used a CHI 660c electrochemical workstation. In CV, we used the positive feedback correction to minimize the ohmic drop between the working and the reference electrodes. For DPV, we used peak amplitude of 50 mV, pulse width of 0.05 s, 2 mV increments per cycle, and pulse period of 0.1 s.

¹H and ¹³C NMR spectra were obtained on a Bruker Avance DMX-600 MHz spectrometer operating at 599.90 and 150.61 MHz, respectively, and equipped with a 5 mm TX-1 inverse probe powered by field gradients along the x,y,z-axes. The probe temperature was controlled (±0.1 °C) with a Bruker BVT3000 temperature controller. The chemical shift (δ) values are given as ppm downfield from internal tetramethylsilane, for both ¹H and ¹³C nuclei. To ensure a complete relaxation for all the resonances, the integrals of the proton spectra were obtained using a pre-scan delay of 10 s. All measurements were carried out in benzene- d_6 . The proton assignments were performed by COSY or TOCSY, whereas the ¹³C chemical shift values were obtained from HMQC experiments. (¹H-¹H) homodecoupling experiments were performed with the standard zghd pulse sequence provided in the Bruker library.

Single-crystal X-ray data for the metal doped ${\rm Au}_{24}M({\rm SR})_{18}{}^0$ clusters were collected either with a Rigaku Oxford Diffraction SuperNova dual-source X-ray diffractometer using hi-flux Mo and Cu micro-focus sources (Mo ${\rm K}_\alpha$; $\lambda=0.71073$ Å and Cu ${\rm K}_\alpha$; $\lambda=1.54184$ Å) and an Atlas CCD detector (University of Jyväskylä), and/or with an Oxford Diffraction Xcalibur Gemini diffractometer with Mo-radiation and Eos CCD detector (University of Padova). Data collection, reduction processes, and analytical numeric absorption corrections by multifaceted crystal models and/or empirical absorption correction using spherical harmonics, were all carried out using the program CrysAlisPro (v. 39.46). Structures were solved by direct methods with program

SHELXT⁵⁹ and refined by full-matrix least-squares on F^2 by SHELXL⁶⁰ in the OLEX² (v. 1.2.10) program.⁶¹

ASSOCIATED CONTENT

Supporting Information. Chemicals, syntheses, NMR tables, further figures (UV-vis absorption spectroscopy spectra, NMR spectroscopy spectra, MALDI-TOF mass spectra, DPV experiments), and X-ray crystallographic analysis of the single-crystal structures of Au₂₄M(SR)₁₈⁰ samples. The CCDC entries 1938192-1938198 contain the supplementary crystallographic data for this paper. CIFs can obtained free of charge www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk. The Supporting Information is ACS Publication website available on the http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*flavio.maran@unipd.it

ORCID

Sabrina Antonello: 0000-0002-0090-9922 Tiziano Dainese: 0000-0002-5771-7307 Alessandro Dolmella: 0000-0003-1287-6635 Wenwen Fei: 0000-0003-0965-496X Manu Lahtinen: 0000-0001-5561-3259 Flavio Maran: 0000-0002-8627-6491 Kari Rissanen: 0000-0002-7282-8419 Alfonso Venzo: 0000-0002-3211-0242

Notes

The authors declare no competing financial interest.

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