

This is a self-archived version of an original article. This version may differ from the original in pagination and typographic details.

Author(s): Daza Secco, Emmanuella; Haimi, Jari; Högmander, Harri; Taskinen, Sara; Niku, Jenni; Meissner, Kristian

Title: Testate amoebae community analysis as a tool to assess biological impacts of peatland use

Year: 2018

Version: Accepted version (Final draft)

Copyright: © Springer Science+Business Media B.V., part of Springer Nature 2018.

Rights: In Copyright

Rights url: <http://rightsstatements.org/page/InC/1.0/?language=en>

Please cite the original version:

Daza Secco, E., Haimi, J., Högmander, H., Taskinen, S., Niku, J., & Meissner, K. (2018). Testate amoebae community analysis as a tool to assess biological impacts of peatland use. *Wetlands Ecology and Management*, 26(4), 597-611. <https://doi.org/10.1007/s11273-018-9594-y>

1 TESTATE AMOEBAE COMMUNITY ANALYSIS AS A TOOL TO ASSESS BIOLOGICAL
2 IMPACTS OF PEATLAND USE

3

4 Daza Secco, Emmanuela. Department of Biological and Environmental Science, P.O.Box 35, FI-40014
5 University of Jyväskylä, Finland. Finnish Environment Institute. Survontie 9 A, Jyväskylä, FI-40500
6 Finland.

7 Email: emitadaza@gmail.com, emdazase@student.jyu.fi

8 Phone: (+57) 3502718987

9

10 Haimi, Jari. Department of Biological and Environmental Science, P.O.Box 35, FI-40014 University of
11 Jyväskylä, Finland.

12

13 Högmander, Harri. Department of Mathematics and Statistics, P.O.Box 35 (MaD), FI-40014 University
14 of Jyväskylä, Finland.

15

16 Taskinen, Sara. Department of Mathematics and Statistics, P.O.Box 35 (MaD), FI-40014 University of
17 Jyväskylä, Finland.

18

19 Niku, Jenni. Department of Mathematics and Statistics, P.O.Box 35 (MaD), FI-40014 University of
20 Jyväskylä, Finland.

21

22 Meissner, Kristian. Finnish Environment Institute. Survontie 9 A, Jyväskylä, FI-40500 Finland.

23

24

25

26

27

28

29

30

31 ABSTRACT

32 As most ecosystems, peatlands have been heavily exploited for different human purposes. For
33 example, in Finland the majority is under forestry, agriculture or peat mining use. Peatlands play an
34 important role in carbon storage, water cycle, and are a unique habitat for rare organisms. Such properties
35 highlight their environmental importance and the need for their restoration. To monitor the success of
36 peatland restoration sensitive indicators are needed. Here we test whether testate amoebae can be used
37 as a reliable bioindicator for assessing peatland condition. To qualify as reliable indicators, responses in
38 testate amoebae community structure to ecological changes must be stronger than random spatial and
39 temporal variation.

40 In this study, we simultaneously assessed differences between the effects of seasonality,
41 intermediate scale spatial variation and land uses on living testate amoebae assemblages in natural,
42 forested and restored peatlands. We expected the effects of seasonality on testate amoebae communities
43 to be less pronounced than those of land use and within site variation.

44 On average, natural sites harboured the highest richness and density, while the lowest numbers
45 were found at forestry sites. Despite small changes observed in taxa dominance and differences in TA
46 community structure between seasons and years at some sites, spatial heterogeneity, temperature, pH,
47 nor water table depth seemed to significantly affect testate amoebae communities. Instead, observed
48 differences were related to type of land use, which explained 75% of the community variation. Our results
49 showed that testate amoebae community monitoring is a useful tool to evaluate impacts of human land
50 use on boreal peatlands.

51 Keywords: Bioindicators, boreal peatlands, forestry, land uses, peatland restoration.

52

53 ACKNOWLEDGEMENTS

54 We thank Yuri Mazei for his guidance on testate amoebae taxonomy and Merle Pöntynen for her
55 valuable work in the field. Emmanuela Daza Secco thanks the Maj and Tor Nessling foundation for the
56 financial support.

57

58 INTRODUCTION

59 While peatlands of the boreal and subarctic regions cover only 3% of the global area, they store
60 ca. 500Gt of C corresponding to one third of the terrestrial carbon storage (Yu 2011, 2012). Roughly 50

61 % of the total Finnish peatland area has been used for forestry, followed by 2.6% for agriculture, and
62 0.6% for peat mining (Lappalainen 1996; Vasander et al. 2003). Peatland use in Finland is mainly
63 concentrated to the central and southern parts of the country where less than 25% of the peatland area is
64 pristine (Aapala et al. 1996).

65 Besides their importance as carbon storage, peatlands play an important role in balancing the
66 water cycle. They are also a unique habitat for many organisms including many rare and endangered
67 species and form repositories of paleontological information through the accumulation and storage of
68 remains of flora, fauna, and atmospheric particles (Gorham 1991; Barber 1993). Thanks to these
69 properties, there is an increased attention on the environmental importance of peatlands and the need for
70 restoration of impacted areas to regain lost ecosystem services (e. g. specific biodiversity, carbon sink,
71 etc.; Lunn and Burlton 2013). To accurately determine whether ecosystem structure and functioning are
72 indeed moving towards near-pristine state following restoration attempts, sensitive, yet robust indicators
73 are needed.

74 To date, there is no evidence that any specific indicator (be it biological, chemical, or physical)
75 outperforms others in indicating changes in peatlands or their restoration success (Chapman et al. 2003).
76 Indeed, in the case of biological indicators, it is well known that different taxa respond differently to
77 ecosystem dynamics and environmental gradients (Francez et al. 2000). However, assessments of
78 peatland plant community composition to study restoration success have shown some promising results
79 (e. g. Haapalehto et al. 2011; Laine et al. 2011; Hedberg et al. 2012; Poulin et al. 2013) but little is known
80 about responses of other important components of peatland ecosystems. Recently, the search for efficient
81 indicators has directed the focus on testate amoebae because they possess several beneficial qualities (e.
82 g. Mitchell et al. 1999; Charman 2001; Koenig et al. 2015; Daza Secco et al. 2016). First, testate amoebae
83 (TA) are shell-building protists (Charman 1999). found in a wide range of habitats e.g. soils, lakes, rivers,
84 they are very strongly associated with peatland plants and especially abundant in *Sphagnum* mosses
85 (Tolonen 1986). Second, TA density can be as high as 16×10^6 individuals per m^2 and even in boreal
86 environments TA can produce several generations per year (Sleigh 1989). Third, TA are a vital
87 component of the microorganism community in *Sphagnum*-dominated peatlands where they account for
88 almost half of the community in terms of biomass (Gilbert et al. 1998). Lastly seasonality is generally
89 not expected to significantly affect the TA communities since they can survive throughout the year
90 through their encysting capacity (Gilbert and Mitchell 2006). These factors could make TA a valuable

91 tool in the assessment of short -and long- term responses of peatlands to land use changes (e. g. Koenig
 92 et al. 2015; Daza Secco et al. 2016). However peatland microorganisms can exhibit microtopographic
 93 transitions at scales of few centimetres (Mitchell et al. 2000b) and previous studies on temporal (Warner
 94 et al. 2007) and spatial TA community variation (Mitchell et al. 2000b) highlight the need to account for
 95 spatial variation when trying to assess seasonal community changes. Thus, to which degree spatial or
 96 temporal variation in community structure may confound the use of TA in routine monitoring of
 97 peatlands is yet unknown and warrants study.

98 In this study, we simultaneously assess differences between the effects of seasonality,
 99 intermediate scale spatial variation and land uses on living TA assemblages in i) natural peatlands, ii)
 100 forested peatlands and iii) restored peatlands. We expected the effects of seasonality on TA communities
 101 to be less pronounced than those of land use and within site variation.

102

103 MATERIALS AND METHODS

104 Study sites

105 Study sites were chosen to represent three stages of peatland use: pristine, forested, and restored.
 106 We assumed that if restoration was successful, sites restored many decades ago were likely to resemble
 107 natural sites more than sites actively forested ones.

108 All the studied peatlands are raised bogs, which represent the prevalent peatland type within a
 109 mosaic of *Sphagnum*-dominated peatlands, coniferous forests and lakes in the boreal zone of central
 110 Finland. Sites were chosen based on their similar characteristics such as elevation, mean annual
 111 temperature, and mean annual precipitation (Table 1). Each land use was represented by two peatlands:
 112 Riihineva and Aittosuo (natural), Lahnanen and Ruuskanlampi (forestry), and Aitoneva60 and
 113 Aitoneva80 (restored).

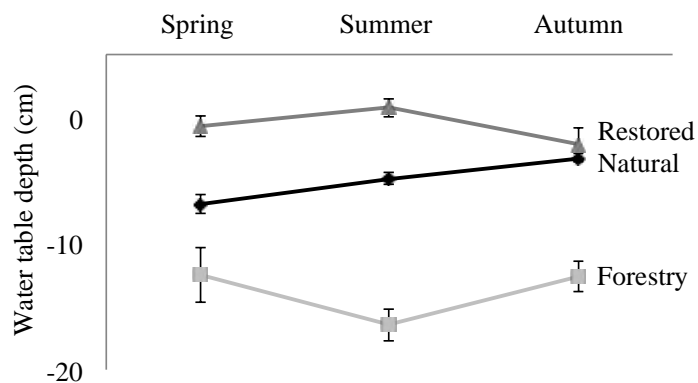
114

115 Table 1. Sampling sites coordinates and elevation by land use. Elevation is given in meters above sea
 116 level. Land use refers to: Natural: peatlands not under direct human influence, Forestry: peatlands used
 117 for forestry, Restored: peatlands previously under human use but restored either 60 or 80 years ago.

| Land use | Coordinates | | Elevation (m.a.s.l.) | Temperature (mean annual °C) | Precipitation (mean annual mm) |
|----------|-------------------------|---------------------------|-------------------------|---------------------------------|-----------------------------------|
| | Longitude | Latitude | | | |
| Natural | 25° 28' 9-24° 37' 53" E | 61° 50' 43"-62° 45' 15" N | ca. 150 | 3 | 600 |
| Forestry | 25° 28' 9-24° 37' 53" E | 61° 50' 43"-62° 45' 15" N | ca. 150 | 3 | 600 |
| Restored | 23° 18' 19.3248" E | 62° 10' 52.1544" N | ca. 100 | 4 | 650 |

118

119 While *Sphagnum* mosses mainly dominated at natural sites, Aittosuo had a higher coverage of
 120 shrubs and trees whereas mosses almost exclusively covered Riihineva. At the forestry sites Lahnanen
 121 and Ruuskanlampi, vegetation was mainly composed of brown and *Sphagnum* mosses with high presence
 122 of trees and bushes. Lahnanen was mainly a dry site surrounded by ditches while Ruuskanlampi
 123 displayed a patchwork of flooded and very dry spots and highly diversified microhabitats. Restored sites
 124 Aitoneva were previously used for peat extraction and were restored either 60 (Aitoneva60) or 80
 125 (Aitoneva80) years ago by blocking ditches and rewetting the sites to restore previous hydrological
 126 conditions in order to allow natural peatland species to recolonise. Aitoneva60 particularly, displayed
 127 the highest water table of all sites, and it was flooded most of the year with small streams crossing the
 128 site. On average, the highest water table depths were found at restored sites while lowest at forestry sites
 129 (Fig. 1).



130
 131 **Fig. 1** Average water table depth by season for the three land uses. Values correspond to measurements
 132 during three years (2013-2015) at each site, during spring, summer and autumn for a total of 15 sampling
 133 plots/site at each sampling time. Restored: peatlands previously under human use but restored either 60
 134 or 80 years ago, Natural: peatlands not under human use, Forestry: peatlands used for forestry. Y-axis:
 135 water table depth given in centimetres below ground level. Points represent mean values; bars represent
 136 SE (± 1).

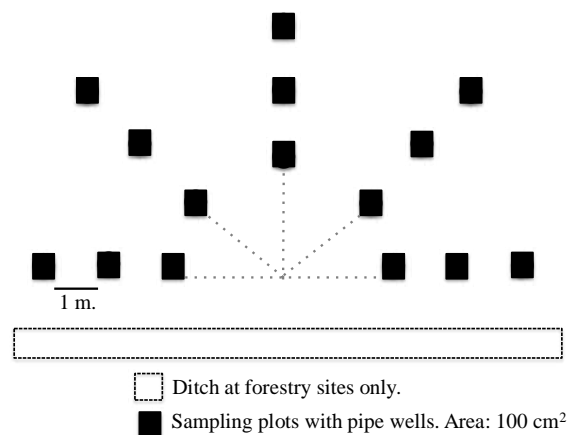
137

138 Field sampling and sample processing

139 Starting from a randomly selected point, fifteen 100 cm²-sampling plots were placed at each site
 140 and distributed in three concentric semicircles of five plots each with ca. 1m spacing (Fig. 2). For water
 141 table depth measurements (WTD), 2cm diameter polypropylene pipe wells with 2mm slits at every 3cm
 142 were placed at the top-left corner of each sampling plot. Water table depth was recorded as negative

143 values when the water level was below ground (top of moss layer), and as positive values when it was
 144 above ground (flooded plots). Water temperature and water pH were measured in the middle of each
 145 sampling plot using a VWR pH meter 1000H. For TA samples, bryophyte mass was collected using a
 146 knife, including only the stem and capitulum of the mosses (ca. 10 cm) in order to obtain mainly living
 147 TA (see Booth et al. 2010). Samples were stored in Ziploc bags at 5°C before further analysis. TA
 148 samples were taken during spring (May), summer (July), and autumn (September) in three consecutive
 149 years (2013, 2014 and 2015). WTD, pH and water temperature from the peat layer were measured during
 150 each TA sampling.

151



153

Fig. 2 Schema of the sampling design for testate amoebae, pH, temperature and water table depth.

154 TA samples were analysed during the following days immediately after sampling in order to
 155 account only for the living TA. Sample processing was carried using the protocol proposed by Booth et
 156 al. (2010). Each sample was boiled for ca. 10 minutes in distilled water with one tablet of *Lycopodium*
 157 *clavatum* spores (batch 1031) standard preparation from Lund University (Sweden). To remove coarse
 158 materials, samples were sieved through 300µm mesh and further filtered onto a 7µm mesh to retain TA
 159 and transferred into a centrifuge tube. TA were centrifuged at 3000rpm for five minutes and stored in
 160 distilled water. Counting and identification of living TA (empty shells were not taken into account) was
 161 done using a 40X magnification (Olympus BX41 microscope). TA were identified mainly to species or
 162 species groups based on characteristics of the shell following a number of different taxonomic keys (e.
 163 g. Charman et al. 2000; Meisterfeld 2002; Clark 2003; Mazei and Tsyganov 2006).

164

165 Data analysis

166 TA number of taxa and relative abundances were calculated for each site (using plot averages),
167 and averaged by land use. Concentrations of TA in 10 cm³ of fresh *Sphagnum* were estimated using the
168 *Lycopodium* counts as an external marker (Stockmarr 1971). The Shannon Wiener diversity index
169 (Shannon and Weaver 1963) was used to estimate TA diversity and a non-parametric Kruskal-Wallis H
170 analysis (Van Hecke 2013) was used to check for temporal differences in TA richness and diversity. TA
171 taxa not frequently found were excluded from ordination analysis to avoid noise. We applied a model-
172 based ordination method as suggested in Hui et al. (2015) and Warton et al. (2015) to visualize the main
173 patterns between different sampling sites in terms of the taxa composition. A model-based ordination
174 approach offers several advantages over traditional distance-based ordination methods such as non-
175 metric multidimensional scaling (NMDS). The main advantage is that models can be used to account for
176 important features such as the mean-variance relationship. For a thorough comparison of model-based
177 and traditional ordination methods, see Hui et al. (2015). A latent variable model with two latent
178 variables was fitted to the data assuming a negative binomial distribution for the TA density. An
179 ordination plot was then produced based on the bivariate latent variables. As the samples were collected
180 in three seasons, we first fitted latent variable models to data sets corresponding to different seasons
181 separately. Lastly, a latent variable model with covariates was fitted in order to identify drivers of amoeba
182 community composition.

183

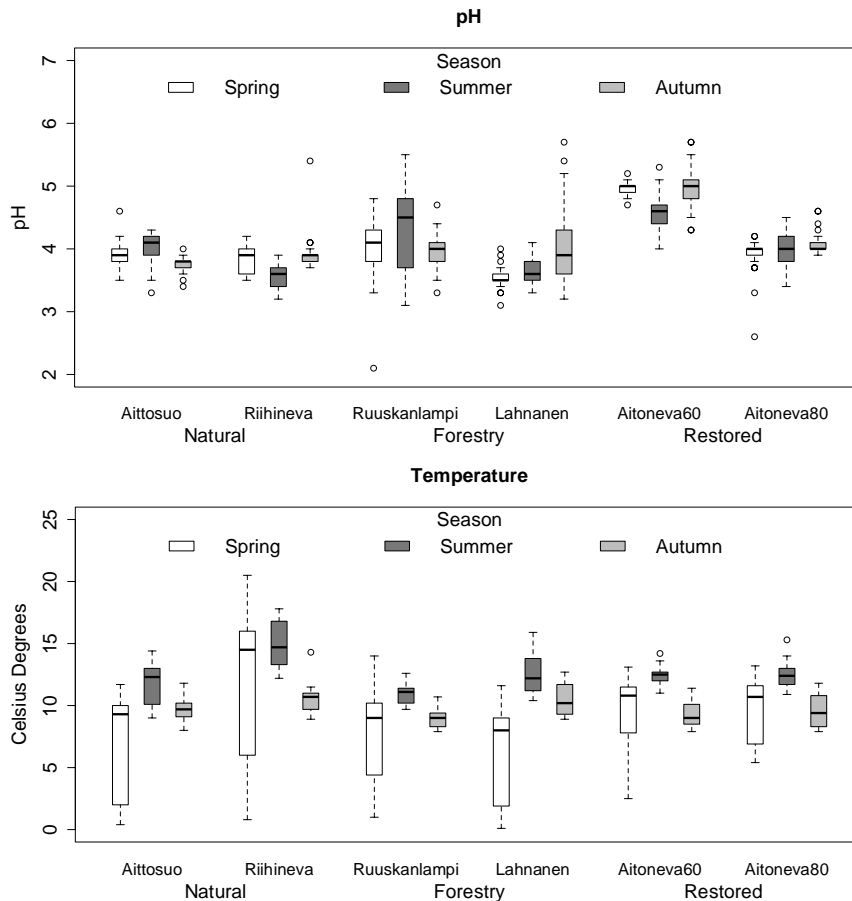
184 RESULTS

185 Environmental variables

186 Environmental variables generally displayed clear patterns during all study years. On average,
187 pH was highest at restored sites, especially at Aitoneva60. Surface water temperature of the peatlands
188 was less variable between sites, but values were slightly higher at the natural site Riihineva (Fig. 3).

189

190



191

192 **Fig. 3** Environmental variables (surface water pH and temperature) in relation to land use and season of

193 sampling. Values correspond to measurements during three years (2013-2015) at each site, during spring,

194 summer and autumn for a total of 15 sampling plots/site at each sampling time. Natural: peatlands not

195 under human use, Forestry: peatlands used for forestry, Restored: peatlands previously under human use

196 but restored either 60 or 80 years ago. Circles: outliers, upper whisker: maximum value excluding

197 outliers, upper box line: upper quartile, middle line inside box: median, lower box line: lower quartile,

198 lower whisker: minimum value excluding outliers.

199

200 TA community data

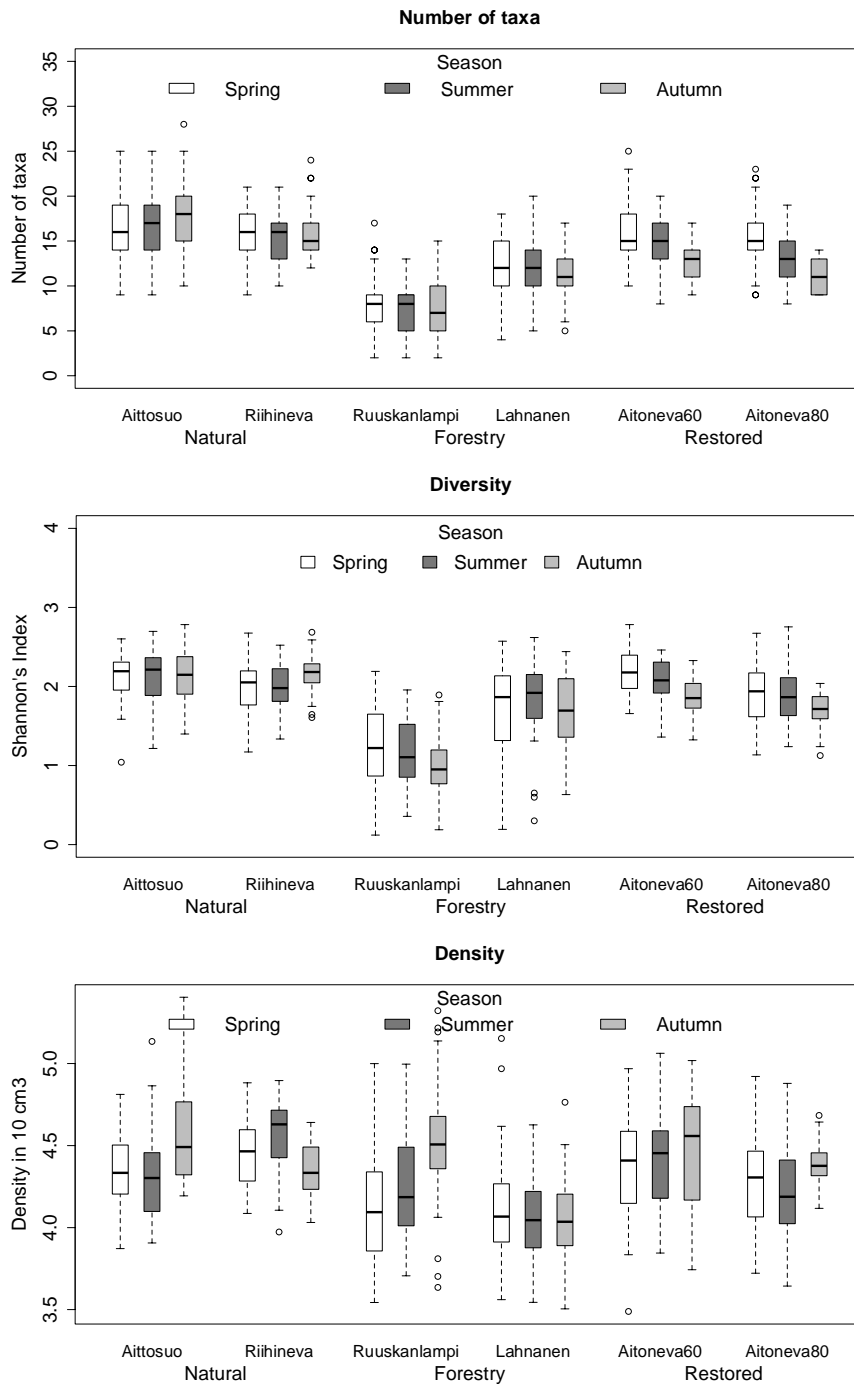
201 We found altogether 62 TA taxa in our study. The highest number of taxa was found at natural

202 sites except in spring 2013 when number of taxa was highest at restored sites (Fig. 4). On average,

203 forestry sites harboured the lowest number of taxa (Fig. 4). TA densities showed a less distinct pattern.

204 Some particularly high densities ($>150,000/10\text{cm}^3$) of TA were recorded in autumn at Aittosuo and

205 Ruuskanlampi (natural and forestry, respectively; Fig. 4).



206

207

Fig. 4 Number of taxa, Shannon Wiener diversity, and density, whisker boxplots for testate amoebae in

208

relation to land use and season of sampling. Values take into account correspond to measurements during

209

three years (2013-2015) at each site, during spring, summer and autumn for a total of 15 sampling

210

plots/site at each sampling time. Natural: peatlands not under human use, Forestry: peatlands used for

211

forestry extraction, Restored: Peatlands previously under human use but restored either 60 or 80 years

212

ago. Circles: outliers, upper whisker: maximum value excluding outliers, upper box line: upper quartile,

213

middle line inside box: median, lower box line: lower quartile, lower whisker: minimum value excluding

214 outliers. Density values were transformed to logarithmic scale, original values ranged between 3081,87
215 and 253383,38 testate amoebae/10 cm³ of fresh *Sphagnum*.

216 Number of TA taxa and diversity were significantly affected by variations in environmental
217 variables between years at Aittosuo (natural) and Lahnanen (forestry) sites, while at both restored sites
218 the differences were related to seasonality. At Riihineva (natural) significant differences were observed
219 just in taxa diversity between seasons, while at Ruuskanlampi (forestry) richness varied between years
220 and diversity between seasons (Table 2).

221 **Table 2** Non-parametric Kruskal-Wallis H tests results of testate amoebae taxa richness and Shannon's
222 diversity index differences between years and seasons ($df=2$ for all comparisons). Significant differences
223 in richness and diversity are marked in bold. Values take into account correspond to measurements during
224 three years (2013-2015) at each site, during spring, summer and autumn for a total of 15 sampling
225 plots/site at each sampling time. The ranges shown in the table refer to the site variation of the annual
226 and seasonal means. Natural: peatlands not under human use, Forestry: peatlands used for forestry
227 extraction, Restored: Peatlands previously under human use but restored either 60 or 80 years ago.

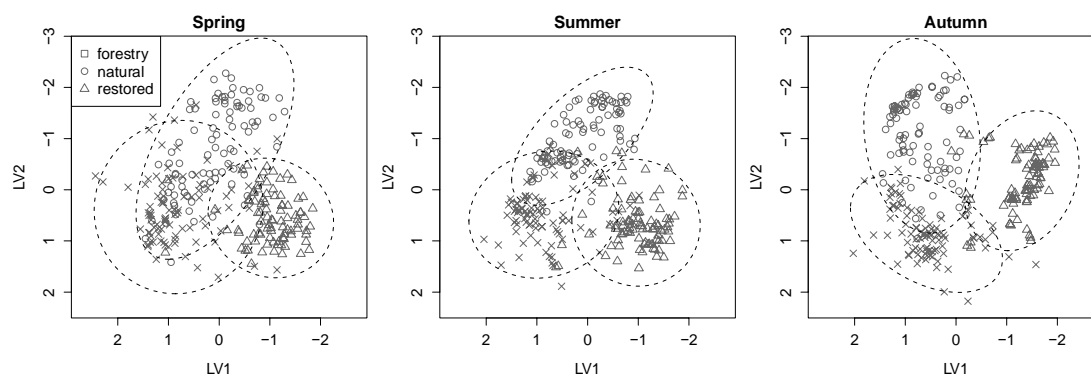
| Land use | site | Variation between years | | | | | | Variation between seasons | | | | | |
|----------|--------------|-------------------------|-----|------------------|--------------------------|-----|------------------|---------------------------|-----|------------------|--------------------------|-----|------------------|
| | | richness | | | Shannon Wiener diversity | | | richness | | | Shannon Wiener diversity | | |
| | | range | H | p | range | H | p | range | H | p | range | H | p |
| Natural | Riihineva | 15.3-15.9 | 1.3 | 0.540 | 1.9-2.1 | 1.2 | 0.540 | 15.5-15.8 | 0.2 | 0.910 | 1.9-2.2 | 11 | 0.005 |
| | Aittosuo | 14.2-18.9 | 41 | <0.001 | 1.9-2.2 | 31 | <0.001 | 16.5-17.6 | 2.3 | 0.310 | 2.1-2.2 | 0.2 | 0.900 |
| | Lahnanen | 10.6-13.2 | 18 | <0.001 | 1.5-1.9 | 9.6 | 0.018 | 11.1-11.8 | 2.2 | 0.330 | 1.7-1.8 | 2.6 | 0.280 |
| Forestry | Ruuskanlampi | 6.7-8.6 | 7.0 | 0.030 | 1.1-1.2 | 1.5 | 0.480 | 7.3-8.1 | 1.1 | 0.560 | 1-1.3 | 6.2 | 0.045 |
| | Aitoneva60 | 14.5-14.8 | 0.1 | 0.960 | 2-2.1 | 1.7 | 0.440 | 12.8-16.1 | 26 | <0.001 | 1.9-2.2 | 27 | <0.001 |
| Restored | Aitoneva80 | 12.6-13.7 | 2.9 | 0.230 | 1.8-1.9 | 4.2 | 0.120 | 11.2-15.6 | 44 | <0.001 | 1.7-1.9 | 11 | 0.003 |

228
229 At natural sites, *Arcella catinus*, *Hyalosphenia papilio* and *Centropyxis aculeata* were the most
230 abundant taxa representing 16.3%, 15.7% and 9.9% of the total average TA numbers, respectively. *A.*
231 *catinus* represented up to 40% of communities at forestry sites. Other common taxa at forestry sites were
232 *C. aculeata* (14%) and *Trigonopyxis arcula* (9.5%). At restored sites dominant taxa differed from those
233 at the other types of peatlands, and showed the highest variability between seasons. Overall *Euglypha*
234 *compressa*, *C. aculeata* and *Diffflugia globulosa* were the most abundant taxa representing on average
235 41% of the communities at restored sites (appendix 1).

236

237 Community-environment relationships and community ordinations

238 In the ordination, the different land uses clustered separately, suggesting that they differed in their
239 TA taxa composition but seasonality did not seem to affect TA communities (Fig. 5).



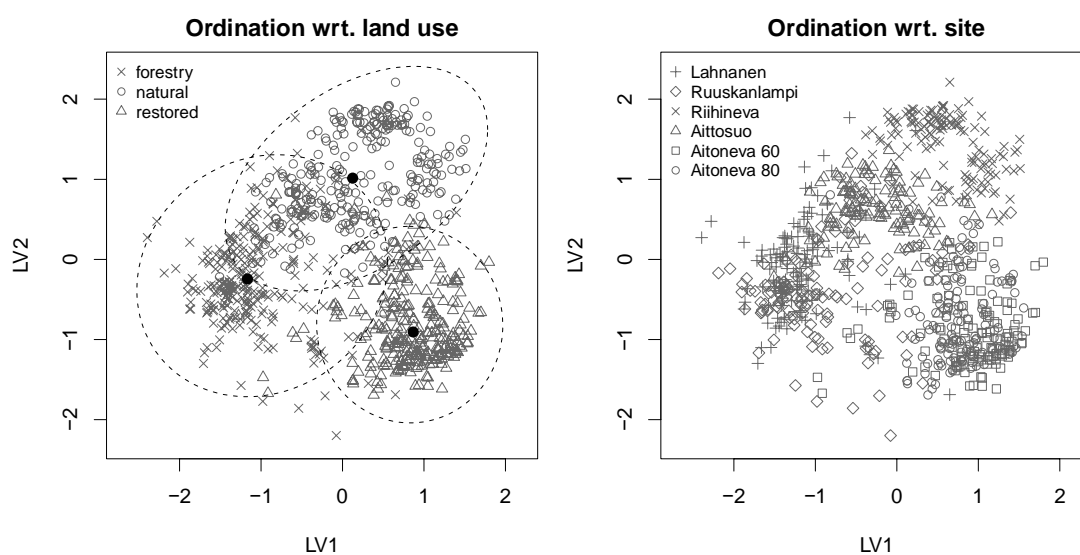
240

241 **Fig. 5** The ordination of $n=270$ sites based on generalized linear latent variable model without any
 242 covariates assuming negative binomial distributed concentrations. The measurements were taken during
 243 the years 2013-2015 and three different seasons. Here the ordinations are shown separately for spring,
 244 summer and autumn. The sites in ordination are labelled according to the land use (Natural: peatlands
 245 not under human use, Forestry: peatlands used for forestry, Restored: Peatlands previously under human
 246 use but restored either 60 or 80 years ago).

247

248 As the seasons did not affect the ordination, we fitted a latent variable model to the whole dataset
 249 (Fig. 6) resulting in a similar pattern as in Fig. 5. When plotting by sites instead of land use, sites still
 250 mainly grouped by land use. The natural site Riihineva displayed a different pattern, where all the plots
 251 grouped separately from the other sites (Fig. 6).

252



253

254 **Fig. 6** The ordination of $n=810$ sites based on generalized linear latent variable model without any
 255 covariates assuming negative binomial distributed concentrations. The measurements were taken during

256 the years 2013-2015 and three different seasons. The sites in left ordination plot are labeled according to
257 the land use (Natural: peatlands not under human use, Forestry: peatlands used for forestry, Restored:
258 Peatlands previously under human use but restored either 60 or 80 years ago), and in right ordination plot
259 according to the sampling site. In left ordination plot, the spatial medians for ordination points
260 corresponding to each land use are shown as black dots.

261

262 To visualize whether the sites align along any of the measured gradients we plotted the sites
263 with shadings corresponding to their respective pH, temperature values, and WTD (Appendix 2).
264 However, none of the variables seemed to affect the ordination i.e. any of the covariates did not affect
265 testate amoebae community structure and composition. To evaluate the amount of variation in testate
266 amoebae taxa caused by different predictors, we used ratios of traces of residual covariance matrices
267 from generalized linear latent variable models as a measure for total variation (Warton et al. 2015).
268 Adding the land use as a covariate to the null model reduced the trace from 497 to 142. Thus, the land
269 use alone explained approximately 71.5% of the covariation across species. Further, the pH alone
270 explained 14.8%, the temperature 2.6% and water table depth 2.3% of the total covariation. To compare
271 the locations of the scatterplots related to different land uses we determined the spatial medians for the
272 sets (see Fig. 6) and tested for differences in spatial medians. The locations differed significantly (Spatial
273 signs test, $p < 0.0001$; Oja and Randles 2004).

274

275

276 DISCUSSION

277 Overall, our results did not show large variations in environmental variables between the land
278 uses except that the highest values of water table depth were recorded at restored and the lowest at
279 forestry sites. Testate amoebae community structure and composition differed between seasons only at
280 the restored sites. However, when all data were analysed together, differences seemed unaffected by
281 seasons or years. Additionally, testate amoebae community structure and composition did not respond to
282 the measured environmental variable gradients (pH, temperature and WTD). Instead, the land use seemed
283 to explain most of the variation between TA communities.

284 On average, natural peatlands Riihineva and Aittosuo harboured the highest number of taxa, and
285 displayed highest diversity and TA density, while the lowest values were found at forestry sites. Wanner

286 and Xylander (2005) found in mineral soils that higher TA taxa richness might be result of a longer time
287 for cumulative colonisation without substantial species replacement, which can also be the explanation
288 for the higher taxa richness found at our natural sites. Additionally, TA commonly occur in highest
289 numbers in wet mosses coverage (Charman 2001), habitat more frequently found at natural sites
290 compared to forestry and restored sites.

291 Forested sites are inherently different from *Sphagnum*-dominated peatlands, thus such differences
292 together with the disturbances caused by forestry practices may have restricted the colonisation and
293 survival of drought intolerant taxa, reducing both the TA taxa richness and their densities. The lowest
294 water tables were in general recorded at forestry sites, where peat was also mainly drier. In *Sphagnum*
295 dominated peatlands, peat moisture is considered the main factor affecting TA communities (Tolonen
296 1986; Booth 2001) and wetter habitats have been found to harbour higher densities of TA (Fournier et
297 al. 2012). Our results generally supported these findings as natural and restored sites showed both higher
298 water table levels and TA densities. However, we found particularly high TA densities in autumn samples
299 at natural and forestry sites. These high TA densities at individual plots were obviously not related to
300 any of the measured environmental variables as these plots were particularly dry (WTD as low as -22cm
301 in some cases) at the time of sampling. While we cannot entirely rule out human errors in sample
302 processing which might have led to such high observed numbers, we feel that observed densities are
303 more likely the result of other favourable environmental factors, such as nutrient concentrations that can
304 either affect TA directly (Lamentowicz et al. 2011) or indirectly by controlling their food sources
305 (Mitchell et al. 2004).

306 Taxa dominance was relatively constant among study years and seasons at natural and forestry
307 sites. The high abundance of *A. catinus* at forestry sites and *H. papilio* at natural sites for example, agrees
308 with common findings suggesting that these taxa are potential indicators of dry and wet conditions,
309 respectively (e.g. Charman and Warner 1992; Mitchell et al. 1999; Bobrov et al. 2002; Galka et al. 2012).

310 The highest abundances of *A. catinus* at natural sites were recorded at the driest plots at Aittosuo
311 where they represented more than 50% of the TA communities. Plots in Aittosuo varied in their
312 hydrological characteristics, some plots being very dry (water level as low as 30cm below ground)
313 compared to the average water level found in this study. However, *A. catinus* was commonly found at
314 all sites, even at the flooded restored sites, but in much lower abundances. This suggests that *A. catinus*
315 could display broad moisture tolerance in Finnish peatlands, as also found by Daza Secco et al. (2016).

316 It should be also noted that the taxonomic keys used in this study group some morphologically similar
317 species into one taxon, which may increase its range of environmental tolerance (for more details see
318 Booth 2001). The highest abundances of *H. papilio* were recorded at Riihineva site where they
319 represented more than 40% of the TA community. Riihineva is a site particularly different in its plant
320 composition. The most remarkable characteristic of this site is the absence of vascular plants while
321 mosses are dominating and creating a homogeneous mat. In fact, in a study by Booth and Zygmunt (2005)
322 *H. papilio* was restricted to floating peat mats, suggesting that hydrological stability of such peat mats is
323 of importance for this taxon. These habitat characteristics were unique to Riihineva and might also
324 explain why TA communities from Riihineva sites grouped particularly separated from the other sites.

325 Compared to forestry and natural sites, taxa dominance was more variable between seasons at
326 restored sites where the two dominant taxa were *E. compressa* and *D. globulosa*. These findings agree
327 with other studies (e.g. Bobrov et al. 1999; Booth 2002) that have found the spiny shells of the spined
328 forms of *Euglypha* help them to restrict their sinking and movements during interstitial water flows to in
329 the wet habitats they are commonly associated with (Bobrov et al. 2002). Both *E. compressa* and *D.*
330 *globulosa* were common and generally abundant also at natural sites, but very scarce or totally absent
331 from forestry sites, suggesting a low tolerance of dry conditions. In contrast, *C. aculeata* was abundant
332 at all sampling sites regardless of their hydrological conditions. Taxa such as *C. aculeata*, with an
333 intermediate moisture optimum, are common along a wide range of the moisture gradient (Booth 2001).
334 It should be pointed that also *C. aculeata* may be similar as *A. catinus*, grouping different species
335 together, and thus influencing its observed tolerance range.

336 Although pH, temperature and WTD are often the most important variables driving TA
337 communities, their effect was not able to mask the effect of land use in our study. Hence, it is possible
338 that TA community structure is driven by other variables such as nutrient concentrations in peat water or
339 biotic interactions more directly linked to the land use. Some studies have found an important relationship
340 between TA and nutrients (e. g. Mitchell et al. 2000a; Jauhiainen 2002; Mitchell et al. 2004). Calcium
341 for example, directly affects some TA species due to its importance in the shell building process
342 (Lamentowicz et al. 2011). While the direct role of other nutrients in the regulation of TA communities
343 is still poorly understood, Mitchell et al. (2004) suggested that nutrients indirectly affect TA via affecting
344 the presence and abundance of their prey organisms.

345 After restoration measurements, it is often inherently expected that both environmental
346 variables and biological communities move towards the pristine conditions of a site, recreating
347 ecosystems that preceded human activities (Choi 2004). However, in our ordinations, sites clustered
348 separately by land use, and natural and restored sites did not show higher similarities between them than
349 they did to forestry sites. A combination of random forces such as percolating rainwater, convective
350 transport into atmosphere, burrowing and other moving efforts, may greatly determine the spatial
351 distributions of dormant protists such as TA (Finlay et al. 2001). Hence, the high similarities in taxa
352 composition between natural and forestry sites might be explained by their geographical proximity, as
353 they were more closely located to each other than to restored sites.

354 A commonly expected main outcome of most ecological restoration programs is that the
355 restored ecosystem reverts back to its pristine conditions; however, the validity of this expectation has
356 recently been openly debated (Wortley et al. 2013). In some wetlands and peatland studies that evaluated
357 restoration success, no conclusive evidence for changes towards a target community after restoration was
358 found (e.g. Zedler and Callaway 1999; Moreno-Mateos et al. 2012). Given such results, evaluations of
359 restoration success should account for: i) the unpredictability of ecological succession, ii) the difficulties
360 to determine the pre-disturbance state of the ecosystem, and the fact that iii) ecological change might be
361 irreversible (Choi 2004). Irreversibility of ecological damage has been previously observed in both
362 diversity and ecosystem services that usually remain lower in the restored than in the reference
363 ecosystems (Wortley et al. 2013; Gałka et al 2017). These findings also highlight the importance of
364 studies including peat core sampling in order to recognise the TA community structure and composition
365 before disturbance, and evaluate whether the observed changes following restoration truly move towards
366 the original states of sites (Gałka 2017).

367 The importance of discriminating between the large-scale climatic effects i.e. seasonal variation,
368 and the local-scale changes caused by environmental disturbance (e.g. drainage) on TA communities has
369 been pointed out earlier (e. g. Mitchell et al. 2000b; Warner et al. 2007; Talbot et al. 2010). Here, we
370 found that despite small changes in taxa dominance (especially at restored sites), changes in TA
371 community structure and composition related to land use strongly overrode effects of seasonality and
372 random spatial variation, explaining 75% of the differences between the TA communities. Land use has
373 been previously reported to greatly affect the microorganisms' communities in wetland environments.
374 For example, Hartman et al. 2008 found that wetland restoration significantly influenced the bacterial

375 community composition when comparing restored vs. reference wetlands even when taking into account
376 soil chemistry and wetland type. The lack of influence of seasonal variation has been previously reported
377 for TA in peatlands (e. g. Gilbert et al. 1998) and TA in other habitat types (e.g. Schönborn 1986).

378 Low temporal variation has been attributed to the trophic diversity behaviour of TA and their
379 capacity to resist changes in temperature and water content through encysting (e.g. Gilbert et al. 1998).
380 On the other hand, studies of soil TA communities (e.g. Finlay and Fenchel 2004; Tysganov et al. 2013)
381 have shown that climate-independent drivers of change i.e. local conditions such as soil moisture, either
382 related to topography or geology (Tysganov et al. 2013) greatly affect the taxa number and abundance
383 of soil TA. The differences in the responses of TA communities to large vs. local-scale environmental
384 variation may also be related to the role of vegetation in regulating microclimatic conditions through
385 shading, precipitation interception, etc. (Wookey et al. 2009; Graae et al. 2012). Other studies on the
386 influence of seasonal fluctuations on TA community structure and composition (e.g. Lamentowicz et al.
387 2013; Marcisz et al. 2014) have found differences mainly in TA density between seasons with highest
388 values during spring. However, such a pattern was not observed in our results.

389 In conclusion, our results showed that in boreal peatlands, the magnitude of the response of TA
390 communities to human-induced environmental changes is higher than their response to local spatial and
391 seasonal environmental variation. Additionally, other studies have also shown the advantages of using
392 not only TA community structure and composition (e. g. Koenig et al. 2015; Daza Secco et al. 2016) but
393 also their functional traits (Marcisz et al. 2016) as bioindicators of peatland disturbance. Here, we suggest
394 the use of TA communities as an efficient tool for assessment and monitoring of ecological changes in
395 boreal peatlands caused by human disturbances. Further, TA can and should be used over broad spatial
396 and temporal scales commonly applied in routine monitoring. An especially attractive property of TA is
397 their robustness against short-term climatic events. This greatly expands the timeframe over which
398 samples from the same year can be combined in analyses, which is an especially helpful feature in the
399 boreal region, where weather conditions from spring to autumn can change drastically.

400

401 REFERENCES

402 Aapala K, Heikkilä R, Lindholm T (1996) Protecting the diversity of Finnish mires. In: Vasander H (ed)
403 Peatlands in Finland. Finnish Peatland Society, Jyväskylä, Finland, pp45–57

404

- 405 Barber KE (1993) Peatlands as scientific archives of biodiversity. *Biodivers Conserv* 2: 474–89. doi:
406 10.1007/BF00056743
407
- 408 Bobrov AA, Charman DJ, Warner BG (1999) Ecology of testate amoebae (Protozoa: Rhizopoda) on
409 peatlands in western Russia with special attention to niche separation in closely related taxa. *Protist* 150:
410 125–136. doi: 10.1016/S1434-4610(99)70016-7
411
- 412 Bobrov AA, Charman DJ, Warner BG (2002) Ecology of Testate Amoebae from Oligotrophic Peatlands:
413 Specific Features of Polytypic and Polymorphic Species. *Biol Bull* 29: 605–617. doi:
414 10.1023/A:1021732412503
415
- 416 Booth RK (2001) Ecology of testate amoebae (Protozoa) in two lake superior coastal wetlands:
417 implications for paleoecology and environmental monitoring. *Wetlands* 21: 564–576. doi:
418 10.1672/0277-5212(2001)021[0564:EOTAPI]2.0.CO;2
419
- 420 Booth RK (2002) Testate amoebae as paleoindicators of surface-moisture changes on Michigan
421 peatlands: modern ecology and hydrological calibration. *J Paleolimnol* 28: 329–348. doi:
422 10.1023/A:1021675225099
423
- 424 Booth RK, Zygmunt JR (2005) Biogeography and comparative ecology of testate amoebae inhabiting
425 *Sphagnum*-dominated peatlands in Great Lakes and Rocky Mountain regions of North America.
426 *Diversity Distrib* 11: 577–590, doi: 10.1111/j.1366-9516.2005.00154.x
427
- 428 Booth RK, Lamentowicz M, Charman DJ (2010) Preparation and analysis of testate amoebae in peatland
429 palaeoenvironmental studies. *Mires and peat* 7: 1–7.
430
- 431 Chapman S, Buttler A, Francez AJ, Laggoun-Défarge F, Vasander H, Schloter M, Combe J, Grosvernier
432 P, Harms H, Epron D, Gilbert D, Mitchell E (2003) Exploitation of northern peatlands and biodiversity
433 maintenance: a conflict between economy and ecology. *Front Eco lEnviron* 1: 525–532. doi:
434 10.1890/1540-9295(2003)001[0525:EONPAB]2.0.CO;2

435

436 Charman DJ, Warner BG (1992) Relationship between testate amoebae (Protozoa: Rhizopoda) and
437 microenvironmental parameters on a forested peatland in northeastern Ontario. *Can J Zool* 70: 2474–
438 2482. doi: 10.1139/z92-331

439

440 Charman DJ (1999) Testate amoebae and the fossil record: issues in biodiversity. *J Biogeogr* 26(1): 89–
441 96. doi: 10.1046/j.1365-2699.1999.00286.x

442

443 Charman, D.J., Hendon, D. & Woodland, W.A. (2000) The identification of testate amoebae (Protozoa:
444 Rhizopoda) in peats, technical guide No.9. Quaternary Research Association, U.K

445

446 Charman DJ (2001) Biostratigraphic and palaeoenvironmental applications of testate amoebae. *Quat Sci*
447 *Rev* 20: 1753–64. doi: 10.1016/S0277-3791(01)00036-1

448

449 Choi YD (2004) Theories for ecological restoration in changing environment: toward “futuristic”
450 restoration. *Ecol Res* 19:75–81. doi: 10.1111/j.1440-1703.2003.00594_19_1.x

451

452 Clarke KJ (2003) Guide to the Identification of Soil Protozoa - Testate Amoebae, report No. 4. Soil
453 Biodiversity Programme Research, Fresh Water Biological Association, Ambleside, U.K

454

455 Daza Secco E, Haapalehto T, Haimi J, Meissner K, Tahvanainen T (2016) Do testate amoebae
456 communities recover in concordance with vegetation after restoration of drained peatlands? *Mires and*
457 *Peat* 18(12): 1–14. doi: 0.19189/MaP.2016.OMB.231

458

459 Finlay BJ, Esteban GF, Clarke KJ, Olmo JL (2001) Biodiversity of terrestrial protozoa appears
460 homogeneous across local and global spatial scales. *Protist* 152: 355–366. doi: 10.1078/1434-4610-
461 00073

462

463 Finlay BJ, Fenchel T (2004) Cosmopolitan metapopulations of free-living microbial eukaryotes. *Protist*
464 155(2): 237–244. doi: 10.1078/1434-4610-00179

465

466 Fournier B, Malysheva E, Mazei Y, Moretti M, Mitchell EAD (2012) Toward the use of testate amoebae
467 functional traits as indicator of floodplain restoration success. *Eur J Soil Biol*49: 85–91. doi:
468 10.1016/j.ejsobi.2011.05.008

469

470 Francez AJ, Gogo S, Josselin N (2000) Distribution of potential CO₂ and CH₄ productions,
471 denitrification and microbial biomass C and N in the profile of a restored peatland in Brittany (France).
472 *Eur J Soil Biol* 36: 161–68. doi: 10.1016/S1164-5563(00)01057-8

473

474 Galka M, Miotk-Szpiganowicz G, Goslar T, JeÅLsko M, van der Knaap WO, Lamentowicz M (2012)
475 Palaeohydrology, fires and vegetation succession in the southern Baltic during the last 7500 years
476 reconstructed from a raised bog based on multi-proxy data. *Palaeogeogr Palaeoclimatol Palaeoecol* 370:
477 209-221 doi: 10.1016/j.palaeo.2012.12.011

478

479 Gałka M, Tobolski K, Górski A, Lamentowicz M (2017) Resilience of plant and testate amoeba
480 communities after climatic and anthropogenic disturbances in a Baltic bog in Northern Poland:
481 implications for ecological restoration. *The Holocene* 27: 130-141. doi: 10.1177/0959683616652704

482

483 Gilbert D, Amblard C, Bourdier G, Francez AJ (1998) The microbial loop at the surface of a peatland:
484 structure, function, and impact of nutrient input. *Microb Ecol* 35: 83–93. doi: 10.1007/s002489900062

485

486 Gilbert D, Mitchell EAD (2006) Chapter 13: Microbial diversity in *Sphagnum* peatlands. In: Martini IP,
487 Martinez Cortizas A, Chesworth W (ed) *Developments in earth surface processes*. Elsevier, Paris, pp.
488 287–318

489

490 Gorham E (1991) Northern peatlands: role in the carbon cycle and probable responses to climatic
491 warming. *Ecol Appl* 1: 182–95. doi: 10.2307/1941811

492

493 Graae BJ, De Frenne P, Kolb A, Brunet J, Chabrerie O, Verheyen K, Pepin N, Heinken T, Zobel M,
494 Shevtsova A, Nijs I, Milbau A (2012) On the use of weather data in ecological studies along altitudinal

- 495 and latitudinal gradients. *Oikos* 121: 3–19. doi: 10.1111/j.1600-0706.2011.19694.x
- 496
- 497 Haapalehto T, Vasander H, Jauhianen S, Tahvanainen T, Kotiaho JS (2011) The effects of peatland
498 restoration on water-table depth, elemental concentrations, and vegetation: 10 years of changes. *Restor*
499 *Ecol* 19: 587–598. doi: 10.1111/j.1526-100X.2010.00704.x
- 500
- 501 Hartman WH, Richardson CJ, Vilgalys R, Bruland GL (2008) Environmental and anthropogenic controls
502 over bacterial communities in wetland soils. *PNAS* 105(46): 17842–17847. doi:
503 10.1073/pnas.0808254105
- 504
- 505 Hedberg P, Kotowski W, Saetre P, Mälson K, Rydin H, Sundberg S (2012) Vegetation recovery after
506 multiple-site experimental fen restorations. *Biol Conserv* 147(1): 60–67. doi:
507 10.1016/j.biocon.2012.01.039
- 508
- 509 Hui FKC, Taskinen S, Pledger S, Foster SD, Warton D (2015) Model-based approach to unconstrained
510 ordination. *Methods Ecol Evol* 6: 399–411. doi: 10.1111/2041-210X.12236
- 511
- 512 Jauhianen S (2002) Testacean amoebae in different types of mire following drainage and subsequent
513 restoration. *Eur J Protistol* 38: 59–72. doi: 10.1078/0932-4739-00748
- 514
- 515 Koenig I, Feldmeyer-Christe E, Mitchell EAD (2015) Comparative ecology of vascular plants, bryophyte
516 and testate amoebae communities in four *Sphagnum* peatlands along an altitudinal gradient in
517 Switzerland. *Ecol Indic* 54: 48–59. doi: 10.1016/j.ecolind.2015.01.043
- 518
- 519 Laine AM, Leppälä M, Tarvainen O, Päätaalo ML, Seppänen R, Tolvanen A (2011) Restoration of
520 managed pine fens: effect on hydrology and vegetation. *Appl Veg Sci* 14(3): 340–349. doi:
521 10.1111/j.1654-109X.2011.01123.x
- 522

- 523 Lamentowicz L, Gabka M, Rusinska A, Sobczyński T, Owsiany PM, Lamentowicz M (2011) Testate
524 amoeba (Arcellinida, Euglyphida) Ecology along a poor-rich gradient in fens of western Poland. *Int Rev*
525 *Hydrobiol* 96: 356–380. doi: 10.1002/iroh.201111357
526
- 527 Lamentowicz M, Bragazza L, Buttler A, Jassey VEJ, Mitchell EAD (2013) Seasonal patterns of testate
528 amoeba diversity, community structure and species-environment relationships in four Sphagnum-
529 dominated peatlands along a 1300 m altitudinal gradient in Switzerland. *Soil Biol Biochem* 67: 1-11.
530 doi: 10.1016/j.soilbio.2013.08.002
531
- 532 Lappalainen E (1996) General view on world peatland and peat resources. In: Lappalainen E (ed) *Global*
533 *peat resources*. International Peat Society, Jyväskylä, Finland, pp53–56
534
- 535 Lunn A, Burlton B (2013) The border mires: A completed peatland restoration project. *British Wildlife*
536 24: 153–160
537
- 538 Marcisz K, Lamentowicz L, Slowinska S, Slowinski M, Muszak W, Lamentowicz M (2014) Seasonal
539 changes in Sphagnum peatland testate amoeba communities along a hydrological gradient. *Eur J Protistol*
540 50: 445-55. doi: 10.1016/j.ejop.2014.07.001
541
- 542 Marcisz K, Colombaroli D, Jassey VE, Tinner W, Kolaczek P, Galka M, Karpinska-Kolaczek M,
543 Slowinski M, Lamentowicz M (2016) A novel testate amoebae trait-based approach to infer
544 environmental disturbance in Sphagnum peatlands. *Scientific Reports* 6: 33907. doi: 10.1038/srep33907
545
- 546 Mazei Y, Tsyganov A (2006) *Freshwater testate amoebae*. 300pp. (in Russian).
547
- 548 Meisterfeld R (2002) Order Arcellinida Kent, 1880. In: Lee JJ, Leedale, GF, Bradbury P (eds) *The*
549 *Illustrated guide to the Protozoa Vol. 2*. Society of Protozoologists, Lawrence, pp. 827-860
550
- 551 Mitchell EAD, Buttler A, Grosvernier P, Rydin H, Hoosbeek MR, Greenup A, Saarinen T (1999)
552 *Ecology of testate amoebae (Protozoa) on peatlands in the Jura mountains, Switzerland and France*.

- 553 Ecoscience 6: 565–576
- 554
- 555 Mitchell EAD, Buttler AJ, Grosvernier P, et al (2000a) Relationships among testate amoebae (Protozoa),
556 vegetation and water chemistry in five *Sphagnum*-dominated peatlands in Europe. *New Phytol* 145: 95–
557 106. doi: 10.1046/j.1469-8137.2000.00550.x
- 558
- 559 Mitchell EAD, Borcard D, Buttler AJ, Grosvernier Ph, Gilbert DJM (2000b) Horizontal distribution
560 patterns of testate amoebae (Protozoa) in a *Sphagnum magellanicum* carpet. *Microb Ecol* 39: 290–300.
561 doi: 10.1007/s002489900187
- 562
- 563 Mitchell EAD, Bragazza L, Gerdol R (2004) Testate amoebae (Protista) communities in *Hylocomium*
564 *splendens* (Hedw.) B.S.G. (Bryophyta): relationships with altitude, and moss elemental chemistry. *Protist*
565 155: 423–436. doi: 10.1078/1434461042650334
- 566
- 567 Moreno-Mateos D, Power ME, Comin FA, Yockteng R (2012) Structural and functional loss in restored
568 wetland ecosystems. *PLoS Biol*. doi: 10: e1001247.
- 569
- 570 Oja H, Randles RH (2004) Multivariate nonparametric tests. *Stat Sci* 19: 598–605. doi:
571 10.1214/088342304000000558
- 572
- 573 Poulin M, Andersen R, Rochefort L (2013) A new approach for tracking vegetation change after
574 restoration: a case study with peatlands. *Restor Ecol* 21(3): 363–371. doi: 10.1111/j.1526-
575 100X.2012.00889.x
- 576
- 577 Schönborn W (1986) Population dynamics and production biology of testate amoebae (Rhizopoda,
578 Testacea) in raw humus of two coniferous forest soils. *Arch Protistenkunde* 132: 325–342. doi:
579 10.1016/S0003-9365(86)80027-6
- 580
- 581 Shannon C, Weaver W (1963) *The mathematical theory of communication*. Urbana: University of Illinois
582 Press.

583

584 Sleight MA (1989) Protozoa and other protists 2nd edition. Edward Arnold, London

585

586 Stockmarr J (1971) Tablets with spores used in absolute pollen analysis. *Pollen et Spores* 13: 615–21.

587

588 Talbot J, Richard PJH, Roulet NT, Booth RK (2010) Assessing long-term hydrological and ecological
589 responses to drainage in a raised bog using paleoecology and a hydrosequence. *J Veg Sci* 21: 143–156.

590 doi: 10.1111/j.1654-1103.2009.01128.x

591

592 Tolonen K (1986) Rhizopod analysis In: Berglund BE (ed) Handbook of the holocene paleoecology
593 and paleohydrology. John Wiley & Sons, Chichester, UK, pp 645–666

594

595 Tsyganov AN, Milbau A, Beyenes L (2013) Environmental factors influencing soil testate amoebae in
596 herbaceous and shrubby vegetation along an altitudinal gradient in subarctic tundra (Abisko, Sweden).

597 *Eur J Protistol* 49(2): 238–248. doi: 10.1016/j.ejop.2012.08.004

598

599 Van Hecke (2013) Power study of anova versus Kruskal-Wallis test. *J Stat Manag Syst* 15 (2-3): 241-
600 247. doi: 10.1080/09720510.2012.10701623

601

602 Vasander H, Tuittila E, Lode E et al (2003) Status and restoration of peatlands in northern Europe. *Wet*
603 *Ecol Manag* 11: 5. doi: 10.1023/A:1022061622602

604

605 Wanner M, Xylander WER (2005) Biodiversity development of terrestrial testate amoebae: Is there any
606 succession at all? *Biol Fertil Soils* 41: 428–438. doi: 10.1007/s00374-005-0850-y

607

608 Warner BG, Asada T, Quinn NP (2007) Seasonal influences on the ecology of testate amoebae (Protozoa)
609 in a small Sphagnum peatland in Southern Ontario, Canada. *Microb Ecol* 54: 91–100

610

611 Warton DI, Guillaume Blanchet F, O'Hara R, Ovaskainen O, Taskinen S, Walker SC, Hui FKC (2015)

612 So many variables: Joint modelling in community ecology. *Trends Ecol Evol* 30: 766–779.
613 doi:10.1016/j.tree.2015.09.007

614

615 Wookey PA, Aerts R, Bardgett RD, Baptist F, Bråthen KA, Cornelissen JHC, Gough L, Hartley IP,
616 Hopkins DW, Lavorel S, Shaver GR (2009) Ecosystem feedbacks and cascade processes: understanding
617 their role in the responses of Arctic and alpine ecosystems to environmental change. *Glob. Change Biol.*
618 15: 1153–1172. doi: 10.1111/j.1365-2486.2008.01801.x

619

620 Wortley L, Hero JM, Howes M (2013) Evaluating ecological restoration success: a review of the
621 literature. *Restor Ecol* 21: 537–543. doi: 10.1111/rec.12028

622

623 Yu ZC (2011) Holocene carbon flux histories of the world's peatlands. *Holocene* 21 (5): 761–774. doi:
624 10.1177/0959683610386982

625 Yu ZC (2012) Northern peatland carbon stocks and dynamics: a review. *Biogeosciences* 9: 4071–4085.
626 doi: 10.5194/bg-9-4071-2012

627

628 Zedler JB, Callaway JC (1999) Tracking wetland restoration: Do mitigation sites follow desired
629 trajectories? *Rest Ecol* 7: 69–73. doi: 10.1046/j.1526-100X.1999.07108.x

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647 APPENDIXES

648 Appendix 1. Testate amoebae taxa mean relative abundances. Values correspond to percentage of

649 presence of each taxa relative to the total of testate amoebae found.

| Species | Natural | | | Forestry | | | Restored | | |
|------------------------------------|---------|--------|--------|----------|--------|--------|----------|--------|--------|
| | Spring | Summer | Autumn | Spring | Summer | Autumn | Spring | Summer | Autumn |
| <i>Amphitrema flavum</i> | 2.00 | 1.43 | 1.98 | 0.23 | 0.10 | 0.00 | 0.08 | 0.06 | 0.02 |
| <i>A. wrightianum</i> | 0.01 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 |
| <i>Arcella catinus</i> | 13.52 | 16.94 | 18.48 | 40.65 | 38.06 | 40.92 | 4.92 | 7.29 | 5.40 |
| <i>A. gibbosa</i> | 0.03 | 0.00 | 0.00 | 0.05 | 0.00 | 0.00 | 0.77 | 0.82 | 0.02 |
| <i>A. vulgaris</i> | 0.07 | 0.09 | 0.10 | 0.19 | 0.31 | 0.09 | 4.29 | 6.76 | 11.85 |
| <i>A. discoides</i> | 0.32 | 0.15 | 0.07 | 2.91 | 2.21 | 3.07 | 6.57 | 4.32 | 2.49 |
| <i>Argynnia vitraea</i> | 0.03 | 0.09 | 0.02 | 0.03 | 0.00 | 0.00 | 0.50 | 0.65 | 0.55 |
| <i>Asulina muscorum</i> | 1.87 | 1.35 | 1.55 | 3.65 | 3.75 | 4.56 | 0.43 | 0.54 | 0.34 |
| <i>A. seminulum</i> | 0.32 | 0.31 | 0.41 | 0.11 | 0.23 | 0.34 | 0.00 | 0.00 | 0.00 |
| <i>Bulinularia indica</i> | 3.54 | 2.76 | 2.91 | 2.04 | 1.27 | 1.99 | 0.16 | 0.02 | 0.04 |
| <i>Centropyxis aculeata</i> | 9.49 | 8.79 | 11.31 | 12.27 | 14.85 | 15.96 | 11.12 | 13.45 | 14.32 |
| <i>C. cassis</i> | 1.49 | 0.72 | 0.83 | 4.11 | 4.22 | 6.12 | 1.17 | 0.25 | 0.40 |
| <i>C. ecornis</i> | 0.38 | 0.40 | 0.16 | 0.01 | 0.40 | 0.14 | 0.02 | 0.00 | 0.00 |
| <i>C. platystoma</i> | 0.01 | 0.07 | 0.00 | 0.06 | 0.04 | 0.00 | 0.01 | 0.09 | 0.01 |
| <i>Cyclopyxis arcelloides</i> | 4.55 | 2.03 | 3.74 | 2.51 | 2.33 | 1.57 | 5.28 | 4.44 | 2.99 |
| <i>Cryptodiffugia oviformis</i> | 0.03 | 0.08 | 0.06 | 0.17 | 0.61 | 0.20 | 0.08 | 0.09 | 0.00 |
| <i>Cyphoderia ampulla</i> | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.01 |
| <i>Diffugia rubescens</i> | 0.01 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.08 | 0.05 | 0.00 |
| <i>D. bacillifera</i> | 0.24 | 0.29 | 0.68 | 0.80 | 0.06 | 0.20 | 8.01 | 8.34 | 13.88 |
| <i>D. oblonga</i> | 0.06 | 0.20 | 0.23 | 0.23 | 0.39 | 0.02 | 4.37 | 5.15 | 5.32 |
| <i>D. lucida</i> | 0.93 | 0.74 | 0.67 | 0.20 | 0.01 | 0.04 | 2.64 | 1.48 | 0.78 |
| <i>D. pristis</i> | 0.03 | 0.00 | 0.00 | 0.07 | 0.00 | 0.00 | 0.59 | 0.07 | 0.00 |
| <i>D. globulosa</i> | 1.69 | 3.25 | 2.56 | 1.92 | 1.73 | 1.62 | 9.88 | 11.26 | 14.32 |
| <i>D. bacilliarum</i> | 0.07 | 0.00 | 0.01 | 0.03 | 0.00 | 0.00 | 0.79 | 0.25 | 0.04 |
| <i>D. acuminata</i> | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.04 | 0.00 | 0.00 |
| <i>D. lithophila</i> | 0.16 | 0.02 | 0.00 | 0.01 | 0.06 | 0.00 | 1.41 | 0.68 | 0.30 |
| <i>D. leidy</i> | 1.53 | 1.49 | 2.19 | 0.03 | 0.06 | 0.00 | 0.08 | 0.19 | 0.23 |
| <i>Euglypha strigosa</i> | 7.30 | 5.98 | 6.49 | 1.27 | 2.53 | 0.99 | 11.35 | 3.75 | 1.03 |
| <i>E. compressa</i> | 2.83 | 7.95 | 6.94 | 0.54 | 0.16 | 0.64 | 8.81 | 14.23 | 16.79 |
| <i>E. tuberculata</i> | 0.32 | 0.21 | 0.19 | 0.39 | 1.24 | 0.47 | 0.62 | 0.43 | 0.00 |
| <i>E. rotunda</i> | 0.27 | 0.20 | 0.22 | 0.85 | 0.24 | 0.56 | 0.41 | 0.37 | 0.12 |
| <i>E. cristata</i> | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 |
| <i>Heleopera rosea</i> | 0.82 | 0.94 | 0.62 | 0.81 | 0.37 | 1.21 | 0.44 | 0.17 | 0.04 |
| <i>H. sphagni</i> | 10.33 | 12.34 | 6.87 | 0.55 | 0.79 | 0.17 | 0.15 | 0.15 | 0.00 |
| <i>H. petricola</i> | 1.85 | 3.84 | 2.30 | 0.19 | 0.28 | 0.07 | 0.11 | 0.03 | 0.07 |
| <i>H. sylvatica</i> | 0.30 | 0.25 | 0.23 | 0.72 | 0.42 | 0.05 | 0.02 | 0.01 | 0.00 |
| <i>Hyalosphenia minuta</i> | 0.05 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 |
| <i>H. papilio</i> | 18.97 | 14.92 | 13.33 | 1.19 | 1.08 | 0.78 | 2.86 | 2.48 | 0.43 |
| <i>H. subflava</i> | 0.01 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.03 | 0.00 | 0.00 |
| <i>H. elegans</i> | 0.79 | 0.50 | 0.67 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 |
| <i>Lesquereusia spiralis</i> | 0.03 | 0.00 | 0.00 | 0.03 | 0.09 | 0.02 | 0.22 | 0.16 | 0.16 |
| <i>Nebela marginata</i> | 0.46 | 0.59 | 0.46 | 0.13 | 0.05 | 0.00 | 3.68 | 5.92 | 3.71 |
| <i>N. carinata</i> | 2.74 | 2.34 | 3.35 | 0.04 | 0.08 | 0.00 | 0.23 | 0.36 | 0.36 |
| <i>N. tubulosa</i> | 0.02 | 0.01 | 0.00 | 0.00 | 0.00 | 0.00 | 1.34 | 0.07 | 0.00 |
| <i>N. parvula</i> | 1.75 | 1.41 | 0.95 | 1.14 | 0.77 | 0.09 | 2.20 | 2.00 | 1.14 |
| <i>N. tincta</i> | 0.88 | 0.85 | 1.20 | 3.54 | 3.47 | 3.78 | 0.34 | 0.08 | 0.04 |
| <i>N. griseola</i> | 1.27 | 1.31 | 1.13 | 0.51 | 0.00 | 0.00 | 0.03 | 0.01 | 0.00 |
| <i>N. flabellulum</i> | 0.01 | 0.00 | 0.00 | 0.03 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| <i>N. militaris</i> | 0.12 | 0.20 | 0.37 | 0.57 | 1.21 | 0.50 | 0.04 | 0.01 | 0.00 |
| <i>N. bohémica</i> | 0.05 | 0.00 | 0.21 | 0.00 | 0.01 | 0.00 | 0.16 | 0.64 | 0.01 |
| <i>Phryganella acropodia</i> | 0.57 | 0.08 | 0.17 | 0.10 | 0.00 | 0.00 | 0.15 | 0.00 | 0.00 |
| <i>Placosista spinosa</i> | 0.32 | 1.40 | 0.78 | 0.04 | 0.01 | 0.01 | 0.01 | 0.33 | 0.00 |
| <i>Pseudodiffugia fascicularis</i> | 0.30 | 0.00 | 0.06 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.01 |
| <i>Sphenoderia lenta</i> | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.04 | 0.01 | 0.00 |
| <i>Tracheleuglyphyphya dentata</i> | 0.00 | 0.00 | 0.00 | 0.08 | 0.06 | 0.06 | 0.10 | 0.10 | 0.13 |
| <i>Trigonopyxis arcula</i> | 3.12 | 2.40 | 2.70 | 10.81 | 10.62 | 7.31 | 0.49 | 0.57 | 0.50 |
| <i>T. minuta</i> | 0.04 | 0.35 | 0.87 | 0.13 | 1.46 | 2.01 | 0.00 | 0.08 | 0.17 |
| <i>Trinema lineare</i> | 0.76 | 0.50 | 1.50 | 1.44 | 2.34 | 2.38 | 1.51 | 1.61 | 1.62 |
| <i>Trinema sp.</i> | 1.31 | 0.23 | 0.44 | 2.42 | 2.04 | 2.06 | 0.50 | 0.19 | 0.10 |
| <i>Sp. 1</i> | 0.00 | 0.00 | 0.00 | 0.18 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| <i>Sp. 2</i> | 0.01 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 | 0.82 | 0.00 | 0.00 |
| <i>Sp. 3</i> | 0.01 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |

650

651

Appendix 2. The ordination of n=810 sites based on generalized linear latent variable model without any

652

covariates assuming negative binomial distributed concentrations. The measurements were taken during

653

the years 2013-2015 and three different seasons. The sites in ordination plot are labelled according to the

654 land use (Natural: peatlands not under human use, Forestry: peatlands used for forestry, Restored:
655 Peatlands previously under human use but restored either 60 or 80 years ago), and coloured according to
656 pH, temperature and water table depth values.
657

