

**This is an electronic reprint of the original article.  
This reprint *may differ* from the original in pagination and typographic detail.**

**Author(s):** Daza Secco, Emmanuella; Haapalehto, T.; Haimi, Jari; Meissner, K.; Tahvanainen, T.

**Title:** Do testate amoebae communities recover in concordance with vegetation after restoration of drained peatlands?

**Year:** 2016

**Version:**

**Please cite the original version:**

Daza Secco, E., Haapalehto, T., Haimi, J., Meissner, K., & Tahvanainen, T. (2016). Do testate amoebae communities recover in concordance with vegetation after restoration of drained peatlands?. *Mires and Peat*, 18, Article 12.  
<https://doi.org/10.19189/MaP.2016.OMB.231>

All material supplied via JYX is protected by copyright and other intellectual property rights, and duplication or sale of all or part of any of the repository collections is not permitted, except that material may be duplicated by you for your research use or educational purposes in electronic or print form. You must obtain permission for any other use. Electronic or print copies may not be offered, whether for sale or otherwise to anyone who is not an authorised user.

1 DO TESTATE AMOEBAE COMMUNITIES RECOVER IN CONCORDANCE WITH VEGETATION  
2 AFTER RESTORATION OF DRAINED PEATLANDS?

3

4 E Daza Secco. University of Jyväskylä, Department of Biological and Environmental Science. Jyväskylä,  
5 Finland. Finnish Environment Institute. Jyväskylä, Finland.

6 T Haapalehto. Metsähallitus Parks and Wildlife Finland. Jyväskylä, Finland.

7 J Haimi. University of Jyväskylä. Department of Biological and Environmental Science. Jyväskylä, Finland.

8 K Meissner. Finnish Environment Institute. Jyväskylä, Finland.

9 T Tahvanainen. Department of Biology, University of Eastern Finland. Joensuu, Finland.

10

11 ABSTRACT

12 The environmental importance of peatlands has raised efforts to restore their specific ecosystem functions  
13 and structure. Monitoring and assessment of peatland ecological state is fundamental in restoration programs.  
14 However, most studies have focused on the responses of vegetation and, to a lesser extent, on testate  
15 amoebae (TA). To our knowledge, none have addressed whether these two groups show concordance with  
16 respect to drained peatland restoration. Here we assessed community concordance between TA and  
17 vegetation among boreal peatlands of four different landuse management classes (natural, drained, restored  
18 3–7 and 9–12 years ago). TA and vegetation communities were concordant when comparing all sites studied.  
19 However, there was no concordance within management classes except for the sites restored 3–7 years ago.  
20 We found that TA and vegetation communities are not surrogates of each other when measuring restoration  
21 success and that a holistic understanding of the changes during restoration from an ecosystem perspective  
22 requires thorough studies of both communities. TA seemed to respond faster to changes caused by  
23 restoration hence, could be a better early indicator of restoration success than plants. Further, studies of TA  
24 and plant communities' relationships could provide important insights into understanding the link between  
25 the recovery of ecosystem structures and functions.

26 Key words: Assessment, ecological state, monitoring, indicators of restoration, taxa surrogates.

27

28

29

## 30 INTRODUCTION

31 Peatlands of the boreal and subarctic regions cover only 3% of the global land area. Due to the accumulation  
32 of organic matter as peat during hundreds to thousands of years, they store ca. 500Gt of C corresponding to  
33 one third of the global soil C (Yu 2011, Yu 2012). Like many other ecosystems, peatlands have been heavily  
34 exploited. In Finland, more than 50% of the total peatland area has been drained for forestry, 2.6% for  
35 agriculture and 0.6% for peat mining (Lappalainen 1996, Vasander *et al.* 2003). Peatland use is mainly  
36 concentrated to central and southern Finland where pristine peatlands now correspond to less than 25% of  
37 their original area (Aapala *et al.* 1996). The increasing threat of environmental degradation has raised  
38 awareness of ecological restoration as part of conservation programs (Dobson *et al.* 1997). The  
39 environmental importance of peatlands has led to a growing number of attempts to restore or partially  
40 rehabilitate the surface water retention, carbon sink, and specific flora and fauna (Lunn & Burlton 2013).

41 Monitoring and assessment of ecological state is a fundamental part of peatland conservation and  
42 restoration programs (Trepel 2007). Studies based on the assessment of the plant community composition  
43 after restoration practices have shown promising results (e.g. Haapalehto *et al.* 2011, Laine *et al.* 2011,  
44 Hedberg *et al.* 2012, Poulin *et al.* 2013). However, little is known about changes in the microorganism  
45 communities of restored peatlands. Testate amoebae (TA) are a polyphyletic group of shell-building,  
46 unicellular protists (Meisterfeld 2002), commonly associated with peatland plants, and especially abundant  
47 in *Sphagnum* mosses (Tolonen 1986). In peatlands, the number of TA can be as high as  $16 \times 10^6$  individuals  
48 per m<sup>2</sup>, making them a significant component of peatland heterotrophic soil community (Sleigh 1989). TA  
49 diversity and distribution in bogs is mainly controlled by hydrological variables (e.g. moisture content and  
50 water table depth), and in fens by water pH, oxygen concentration, and peat composition (Charman 1997,  
51 Bobrov *et al.* 1999). Depending on taxa and environmental conditions, their generation time ranges from  
52 days to weeks, while that of bryophytes and vascular plants is much longer (Schönborn 1986). Under natural  
53 conditions, TA communities are stable between seasons (Warner *et al.* 2007) but when conditions change,  
54 they can change within months (Marcisz *et al.* 2014).

55 Koenig *et al.* (2015) found that TA provide more accurate information on microenvironmental conditions  
56 compared to vegetation, and knowing all the TA taxa is not necessary to obtain valuable ecological  
57 information. Thus, TA communities have been proposed as a tool for monitoring and assessment of peatland

58 conditions, but to date TA have been employed in relatively few studies (e.g. Jauhiainen 2002, Davis &  
59 Wilkinson 2004, Raabe & Lamentowicz 2012, Turner & Swindles 2012, Koenig *et al.* 2015).

60 Well-known taxonomic groups are usually those used as biological indicators in assessment and  
61 monitoring of ecosystem health, i.e. the ecosystem's ability to maintain its structure and function when  
62 facing external stress (Costanza & Mageau 1999). However, the degree to which certain taxonomic groups  
63 actually mirror trends among other groups and can be used as surrogates is still unknown, and studies based  
64 on the surrogate-taxa approach have repeatedly been questioned (e.g. Paavola *et al.* 2003, Bilton *et al.* 2006).  
65 Most bioassessment studies focus on single taxonomic group responses to environmental factors while few  
66 have addressed parallel group responses (i.e. concordance). Community concordance describes the degree of  
67 similarity in distributions and abundances of different taxonomic groups across a region and emerges when  
68 different communities show similar responses to environmental changes (Infante *et al.* 2009). Studies that  
69 have investigated concordance have mostly focused on lakes and fluvial systems (e.g. Heino 2002, Mykrä *et*  
70 *al.* 2008, Infante *et al.* 2009, Jyväsjärvi *et al.* 2014) and very few have examined such patterns for peatland  
71 ecosystems (Mitchell *et al.* 2000a, Lamentowicz *et al.* 2010, Hájek *et al.* 2014, Koenig *et al.* 2015, Hunter Jr  
72 *et al.* 2016).

73 Aspects driving community concordance include strong species interactions within and between  
74 communities (Heino 2002) and co-losses of species in response to environmental stress, including  
75 anthropogenic stressors (Mykrä *et al.* 2008, Yates & Bailey 2010). Life histories and reproductive and  
76 dispersal capabilities of taxa could influence the extent to which different environmental factors and biotic  
77 interactions drive spatial patterns of community composition and subsequently, community concordance  
78 (Shurin *et al.* 2009). Being based on species identities instead of only number of species, community  
79 concordance provides a wider understanding of similarities between communities than richness measures  
80 alone (Pawar *et al.* 2007). Further, when concordance does not occur among taxonomic groups at a specific  
81 geographical scale, the use of a single group as an indicator for the state of the ecosystem may not be  
82 appropriate (Paavola *et al.* 2006).

83 Studies examining peatland restoration efforts generally focus on the recovery of vegetation, but whether  
84 important microorganisms such as TA are concordant in their responses to restoration and how these  
85 responses are linked to general ecosystem functions remains an open question. Because TA are especially  
86 common in *Sphagnum* and other mosses, they may potentially be concordant with peatland vegetation. We

87 therefore assessed concordance of the changes in TA and plant community structures among natural, ditched,  
88 and restored boreal peatlands. We specifically aimed i) to analyse changes and concordance between TA and  
89 plant communities in response to restoration processes, and ii) to identify which environmental variables  
90 drive the changes in these two communities.

91

## 92 METHODS

### 93 Study sites

94 Study sites were located in the south-boreal climatic-phytogeographical zone of Southern Finland between  
95 61° 53'–62°51' N and 22°53'–25°26' E. In this region, the larger peatland formations are mainly raised bogs,  
96 while small weakly minerotrophic *Sphagnum*-dominated mires are typically found in a mosaic landscape  
97 pattern with coniferous forests. Sampling sites are located ca. 150m above sea level with a mean annual  
98 temperature of +4°C and precipitation ca. 650mm. We selected 19 study sites divided in four land use  
99 management classes: i) natural ( $n=5$ ), ii) drained ( $n=4$ ), iii) drained and restored 3–7 years before the  
100 sampling ( $n=5$ ), and iv) drained and restored 9–12 years before the sampling ( $n= 5$ ). For brevity,  
101 management classes will hereafter be referred to as Natural, Ditched, Res05, and Res10.

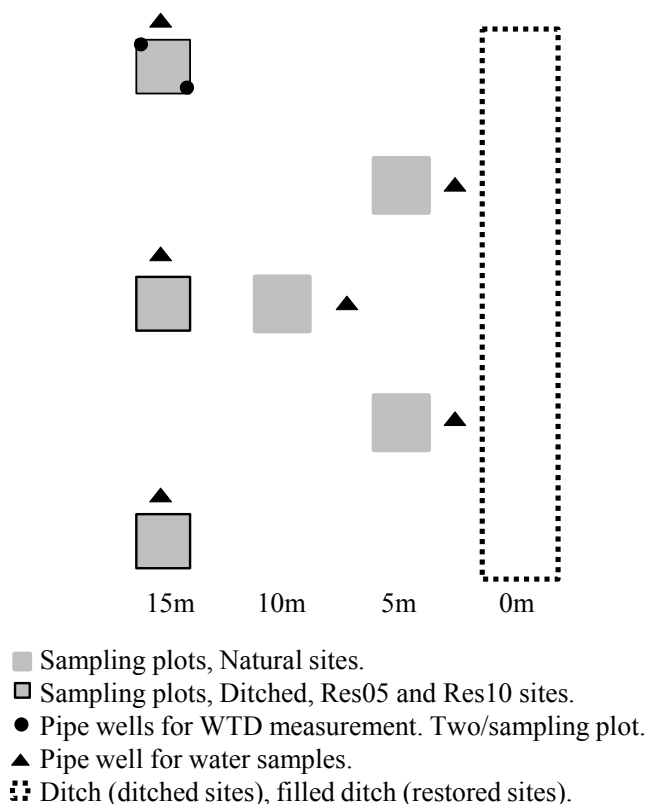
102 Selection of pine fen sites of low minerotrophy was based on field observations and historic aerial  
103 photographs ensured that original tree stands of the drained and restored sites were similar. Sites consisting  
104 of naturally similar type of vegetation were located in a mosaic of ombro-mesotrophic peatland vegetation,  
105 where *Sphagnum* was the main peat-forming plant genus. Based on topographic data and field observations,  
106 surface water flow was considered independent for each management class. Peat depth ranged from 95cm to  
107 more than 200cm with underlying soil of till or sand.

108 Sites without altered hydrology by ditches or other direct measures represent the least impacted  
109 management class and are subsequently referred to as “Natural”. Even though we acknowledge that forestry  
110 measures in distant parts of Natural site catchments may have slight impacts on their hydrology  
111 (Tahvanainen 2011), such effects were not obvious. Ditched and Restored sites were drained for forestry  
112 purposes during the 1960s and 1970s. During the 1980s, peatlands were restored by filling in the ditches and  
113 by removing trees in areas where drainage had significantly increased tree growth. More detailed information  
114 about the studied peatlands is given in Haapalehto *et al.* (2014).

115

116 Field sampling and sample processing

117 A grid of 20 1m<sup>2</sup> plots was established at each site (Figure 1). At the Ditched and Restored sites, the plots  
 118 were situated in five parallel transects, spaced four meters apart and running perpendicular to the ditch. At  
 119 each transect, there was a plot at 5, 10 and 15 meters distance from the ditch. A 15m x 20m grid of plots was  
 120 used at the Natural reference sites. Within each of the study sites, the location of the first plot was  
 121 randomised.



123 Figure 1. Study sampling design for: testate amoebae, plants and environmental variables. Distances (m)  
 124 refer to distance of the 1m<sup>2</sup> plots to the ditches at the Ditched and Restored sites. At Natural sites, a similar  
 125 arrangement of plots was used. Res05: peatlands restored 3-7 years ago; Res10: peatlands restored 9-12  
 126 years ago.

127 For water chemistry analyses at the Natural sites, three 32mm diameter polypropylene pipe wells with  
 128 2mm slits spaced at 2–3cm intervals and polypropylene filter gauges were distributed in the central part of  
 129 the sampling grid. At the Ditched and Restored sites filter gauges were set at a distance of 15m to the ditch  
 130 and 10m apart (Figure 1). Samples were collected in August 2007. Redox potential (Eh<sub>7</sub>), electrical  
 131 conductivity (EC) and pH were measured between two to five hours after sample collection using a Consort

132 SP50X meter with SP50X, SK10T and SP10B electrodes, respectively. Samples were then stored in darkness  
133 at +4°C before further analysis. Cation concentrations (Al, Ca, Fe, K, Mg, Na) were obtained after filtration  
134 with 0.45 µm pore size filters and analysed with a PerkinElmer Optima 4300 DV inductively coupled plasma  
135 optical emission spectrometer. Water table depth (WTD) was measured five times (May, June, July, August,  
136 and September) in 2008 in selected plots from two permanent polypropylene pipe wells. WTD values were  
137 corrected for the natural slope (see Haapalehto *et al.* 2014) and averaged to absolute levels throughout the  
138 site. WTD data are usually recorded as negative values, but to facilitate the readability, inverse, positive  
139 values are used here throughout the text.

140 Plants and TA were sampled from the plots located next to the polypropylene pipe wells (three plots at  
141 each site). We estimated the relative abundance (percentage of coverage) of each taxon of vascular plants,  
142 bryophytes, and some lichens for each sampled plot. Sampling was done during July and August 2007. To  
143 obtain TA, approximately 10cm<sup>3</sup> of bryophyte mass was extracted (see also Booth *et al.* 2010). Samples  
144 were stored in plastic Ziploc bags, frozen, defrosted and oven dried, stored in paper bags and analysed in  
145 2013 using the protocol proposed by Booth *et al.* (2010). To retain TA, each sample was boiled for 10  
146 minutes in distilled water with one tablet of *Lycopodium clavatum* spores (batch 1031) standard preparation  
147 from Lund University (Sweden), and sieved it through a 300µm mesh to remove coarse materials and onto a  
148 7µm mesh. Retained TA were centrifuged at 3000 rpm for five minutes and stored in distilled water. A  
149 minimum of 150 TA was counted and identified using 40X magnification (Olympus BX41 microscope). As  
150 the standing plant community is the result of several years of development, non-living TA were also included  
151 in the analysis to integrate the accumulation of tests over the entire time period. TA identification was based  
152 on characteristics of the shell following a number of different taxonomic keys (e.g. Charman *et al.* 2000 with  
153 modifications by Booth 2008, Meisterfeld 2002, Clark 2003, Mitchell 2003a, Mitchell 2003b, Mazei &  
154 Tsyganov 2006).

155

#### 156 Data analysis

157 We calculated species richness, Shannon diversity index, and relative abundances of TA and vegetation  
158 communities for each site (using plot averages), and averaged by management classes. For TA communities,  
159 density was estimated using the *Lycopodium* counts as an external marker (see Stockmarr 1971 for details)  
160 and relative abundances were calculated as a percentage of the total counted. ANOVA and Tukey's (HSD)

161 post hoc test were performed to assess differences in species richness and Shannon diversity indices between  
162 management classes. Due to the unbalanced design, permutational MANOVA (PerMANOVA) was used to  
163 test the significance of the differences in structure and composition among the communities in the four  
164 management classes. Differences in water chemistry and WTD were analysed using ANOVA or Kruskal-  
165 Wallis depending on whether the data met the normality and homoscedasticity assumptions. To summarise  
166 variability in the communities, a Non-metric Multidimensional Scaling (NMDS) based on Sørensen's (Bray-  
167 Curtis) distance was performed. We used NMDS to examine the degree of concordance of water chemistry  
168 variables with community ordinations. WTD was not included in the community ordinations, as these data  
169 were not taken at exactly the same time as the biological samples. A two dimensional NMDS solution  
170 achieved a moderate stress level for plant communities but not for TA communities; however, for simplicity  
171 of comparison we retained the two dimensional TA solution. Concordance between TA and plant  
172 communities was tested using the PROTEST analysis (Peres-Neto & Jackson 2001). All calculations were  
173 done using R (version 3.0.2) and the vegan package for NMDS and PROTEST analyses (Oksanen *et al.*  
174 2015).

175

## 176 RESULTS

### 177 Community data

178 Altogether 44 TA and 45 plant taxa were found in our study. TA taxon richness at Natural sites ( $\bar{X}$ = 17.7  
179 range 14–24, SE= 0.71) was higher than at Ditched, Res05, and Res10 sites (ANOVA  $F_{[1,3]}= 15.79$ ,  $p <$   
180 0.001) whereas no differences were found among Ditched, Res05, and Res10 sites (Table 1; Figure 2a). TA  
181 Shannon diversity index was generally higher at Natural sites, where the highest value was 2.56, while the  
182 lowest values were found at Res05 sites (Figure 2c). We observed differences in TA diversity only between  
183 Natural-Res05 and Natural-Res10 sites (Table 1). The highest cell density was found at Res10 sites ( $\bar{X}$ =  
184 24953 cells/10cm<sup>3</sup>) and the lowest at Ditched sites ( $\bar{X}$ = 6767 cells/10cm<sup>3</sup>).

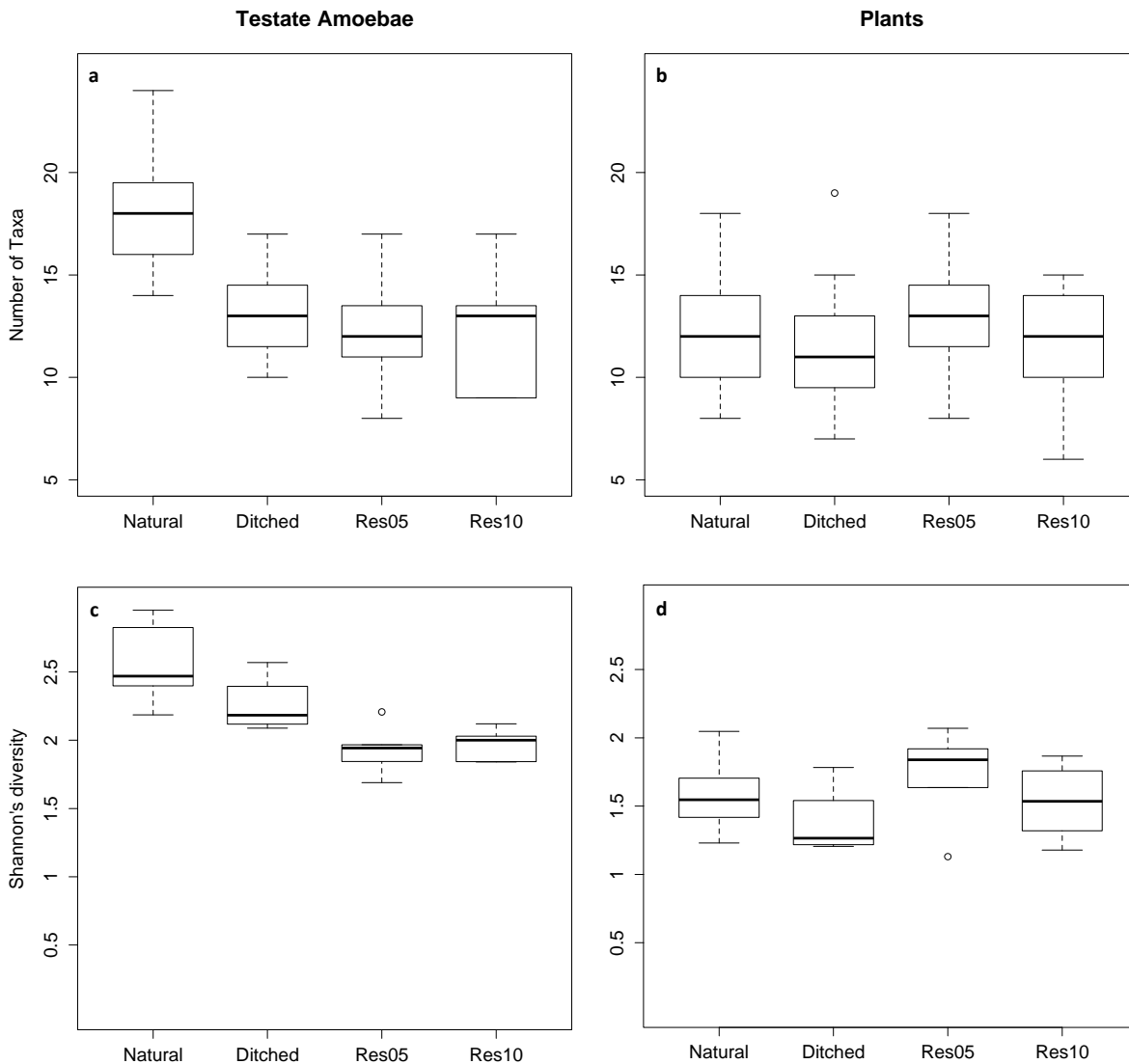
185

186 Table 1. Tukey's (HDS) results for TA taxon richness and Shannon diversity. Res05: peatlands restored 3-7  
187 years ago; Res10: peatlands restored 9-12 years ago.



		Richness				Shannon Diversity			
		Difference	lower bound	Upper bound	<i>p</i>	Difference	lower bound	Upper bound	<i>p</i>
Natural	Res05	5.47	3.00	7.94	<0.001	0.64	0.23	1.04	0.002
	Res10	-5.47	-7.94	-3.00	<0.001	-0.60	-1.00	-0.19	0.003
	Ditched	4.64	1.96	7.33	<0.001	0.31	-0.12	0.74	0.205
Res05	Res10	0.00	-2.47	2.47	1.000	0.04	-0.37	0.44	0.993
	Ditched	-0.82	-3.51	1.86	0.847	-0.33	-0.76	0.10	0.171
Res10	Ditched	-0.82	-3.51	1.86	0.847	-0.29	-0.72	0.14	0.253

188



189

190 Figure 2. Taxon richness boxplots for testate amoebae (a) and vegetation (b). Shannon's diversity boxplots  
 191 for testate amoebae (c) and vegetation (d) for each management class. Circles: outliers, upper whisker:  
 192 maximum value excluding outliers, upper box line: upper quartile, middle line inside box: median, lower box  
 193 line: lower quartile, lower whisker: minimum value excluding outliers. Res05: peatlands restored 3-7 years  
 194 ago; Res10: peatlands restored 9-12 years ago.

195

196 No differences in the mean number of plant taxa were found between management classes (Figure 2b). In  
 197 general, plant diversity tended to be higher at Res05 sites but, overall, there were no significant differences  
 198 between classes ( $F= 0.9, p= 0.46$ ; Table 1; Figure 2d). TA community structure and composition differed  
 199 between classes except between Res05 and Res10 (Table 2). For vegetation communities, only Natural sites  
 200 differed from all other classes (Table 2, see also Haapalehto 2014).

201

202 Table 2. PerMANOVA pairwise comparisons for TA and plant community structure and composition.

203 Res05: peatlands restored 3-7 years ago; Res10: peatlands restored 9-12 years ago.

		TA						Plants					
		<i>df</i>	<i>SS</i>	<i>MS</i>	<i>Pseudo-F</i>	<i>r</i> <sup>2</sup>	<i>p</i>	<i>df</i>	<i>SS</i>	<i>MS</i>	<i>Pseudo-F</i>	<i>r</i> <sup>2</sup>	<i>p</i>
Natural	Res05	1	0.28	0.28	2.74	0.25	0.021	1	0.44	0.44	3.23	0.29	0.042
	Res10	1	0.29	0.29	4.02	0.33	0.016	1	0.15	0.15	1.83	0.19	0.047
	Ditched	1	0.48	0.48	4.90	0.41	0.006	1	0.46	0.46	4.11	0.37	0.016
Res05	Res10	1	0.09	0.09	1.36	0.15	0.259	1	0.33	0.33	2.35	0.23	0.104
	Ditched	1	0.36	0.36	3.90	0.36	0.036	1	0.12	0.12	0.66	0.09	0.499
204 Res10	Ditched	1	0.59	0.59	10.17	0.59	0.010	1	0.35	0.35	2.98	0.30	0.093

205 On average, TA communities in Natural, Res05, and Res10 sites were mainly dominated by *Arcella*  
 206 *catinus* and *Bullinularia indica*. At Ditched sites, *Trigonopyxis arcula* was the most widely dominant taxon  
 207 followed by *A. catinus*. More than 50% of the TA species were found over all sites regardless of  
 208 management class, but some species displayed a more restricted distribution. For example, *Arcella artocrea*,  
 209 *Centropyxis ecornis*, *Diffflugia leidyi*, *Diffflugia lithophila*, *Hyalosphenia minuta* and *Pseudodiffflugia*  
 210 *fascicularis* were present just at Natural sites while *Hyalosphenia elegans* and *Physochila griseola* were  
 211 absent from Ditched and Res05 sites (Table 3). Vegetation communities were dominated by *Sphagnum*  
 212 *angustifolium* at all sites regardless of management class. However, densities were highly variable.  
 213 *Pleurozium schreberi* was the second most dominant species at Res05 and Ditched sites, while at Res10 and  
 214 Natural sites *S. magellanicum* and *S. fuscum* were more abundant (see Haapalehto 2014).

215

216 Table 3. TA average relative abundances in percentage and standard deviations for each management class.

217 (\*) Taxa found just at natural sites; (\*\*) taxa found at Natural and in sites restored 10 years ago; (\*\*\*) taxa  
 218 found at all but Ditched sites; (\*\*\*\*) taxa found just at Ditched sites. Res05: peatlands restored 3-7 years  
 219 ago; Res10: peatlands restored 9-12 years ago.

Taxa	Treatments							
	Natural		Ditched		Res05		Res10	
	$\bar{X}$	SD	$\bar{X}$	SD	$\bar{X}$	SD	$\bar{X}$	SD
* <i>Arcella artocrea</i>	0.78	1.56	0	0	0	0	0	0
* <i>Centropyxis ecornis</i>	0.09	0.33	0	0	0	0	0	0
* <i>Diffugia leidyi</i>	0.05	0.17	0	0	0	0	0	0
* <i>Diffugia lithophila</i>	0.04	0.17	0	0	0	0	0	0
* <i>Hyalosphenia minuta</i>	0.53	0.85	0	0	0	0	0	0
* <i>Pseudodiffugia fascicularis</i>	0.04	0.16	0	0	0	0	0	0
** <i>Hyalosphenia elegans</i>	3.60	3.93	0	0	0	0	0.22	0.48
** <i>Physochila griseola</i>	0.67	1.03	0	0	0	0	0.09	0.35
*** <i>Heleopera petricola</i>	2.56	3.76	0	0	0.34	0.63	0.12	0.34
**** <i>Arcella vulgaris</i>	0	0	0.11	0.37	0	0	0	0
**** <i>Euglypha compressa</i>	0	0	0.11	0.37	0	0	0	0
**** <i>Hyalosphenia subflava</i>	0	0	0.12	0.41	0	0	0	0
**** <i>Phryganella acropodia</i>	0	0	0.06	0.19	0	0	0	0
**** <i>Trigonopyxis minuta</i>	0	0	1.14	3.79	0	0	0	0
**** <i>Unidentified</i>	0	0	0.34	0.81	0	0	0	0
<i>Archerella flavum</i>	6.02	6.27	0.06	0.19	0	0	0.53	1.71
<i>Arcella catinus</i>	20.96	20.17	14.05	16.46	37.44	17.27	44.95	30.18
<i>Arcella discoides</i>	0.13	0.49	0.41	1.03	0.48	1.17	0.16	0.63
<i>Argynnia vitrea</i>	0.18	0.52	0	0	0.04	0.16	0	0
<i>Assulina muscorum</i>	4.86	4.54	4.92	2.85	3.62	2.62	3.03	3.18
<i>Assulina seminulum</i>	0.56	0.89	0.29	0.62	0.09	0.24	0.04	0.17
<i>Bullinularia indica</i>	8.43	5.41	13.08	10.94	16.81	19.60	10.21	11.62
<i>Centropyxis aculeata</i>	1.18	2.90	2.29	4.92	0.29	1.11	1.93	4.17
<i>Centropyxis cassis</i>	0.05	0.17	6.61	6.38	2.02	5.41	0.24	0.65
<i>Centropyxis platystoma</i>	0	0	0	0	0.04	0.17	0	0
<i>Corythion spp.</i>	1.95	2.93	3.47	3.71	4.23	3.67	3.93	5.70
<i>Cyclopyxis arcelloides</i>	0.48	0.66	6.01	5.82	0	0	0.35	0.66
<i>Diffugia globulosa</i>	0.04	0.16	2.33	5.04	0	0	0.13	0.51
<i>Diffugia lucida</i>	0.11	0.30	0.06	0.20	0.04	0.17	0	0
<i>Euglypha rotunda</i>	3.55	4.16	0.97	1.61	0.83	1.04	3.00	3.39
<i>Euglypha strigosa</i>	3.79	3.30	1.23	1.60	3.01	2.86	2.36	2.31
<i>Euglypha tuberculata</i>	1.23	2.07	3.97	6.65	3.65	3.29	2.07	4.09
<i>Heleopera rosea</i>	1.23	1.81	0.18	0.31	0.45	1.23	0.95	1.36
<i>Heleopera sphagni</i>	5.86	9.24	0.28	0.74	0.68	1.24	0.22	0.48
<i>Heleopera sylvatica</i>	3.02	4.12	0.18	0.43	0.68	1.28	0.47	0.56
<i>Hyalosphenia papilio</i>	7.99	8.01	0.29	0.61	4.43	9.75	8.97	10.84
<i>Nebela bohémica</i>	0	0	0	0	0.04	0.15	0.71	2.06
<i>Nebela flabellulum</i>	0	0	0	0	0.04	0.16	0	0
<i>Nebela militaris</i>	4.21	3.74	2.03	3.49	3.15	3.99	1.07	1.11
<i>Nebela parvula</i>	1.12	1.86	0.17	0.57	0.22	0.86	0.27	0.55
<i>Nebela tinctoria</i>	5.34	3.42	5.75	6.01	5.15	4.61	4.65	3.41
<i>Pseudodiffugia fulva</i>	0.04	0.17	0.43	1.43	0	0	0	0
<i>Trigonopyxis arcuata</i>	5.13	5.80	26.61	15.36	7.24	5.86	3.22	3.63
220 <i>Trinema lineare</i>	4.17	6.44	2.43	3.23	4.99	4.59	6.10	7.17

221

222 Community-environment relationships and community concordance

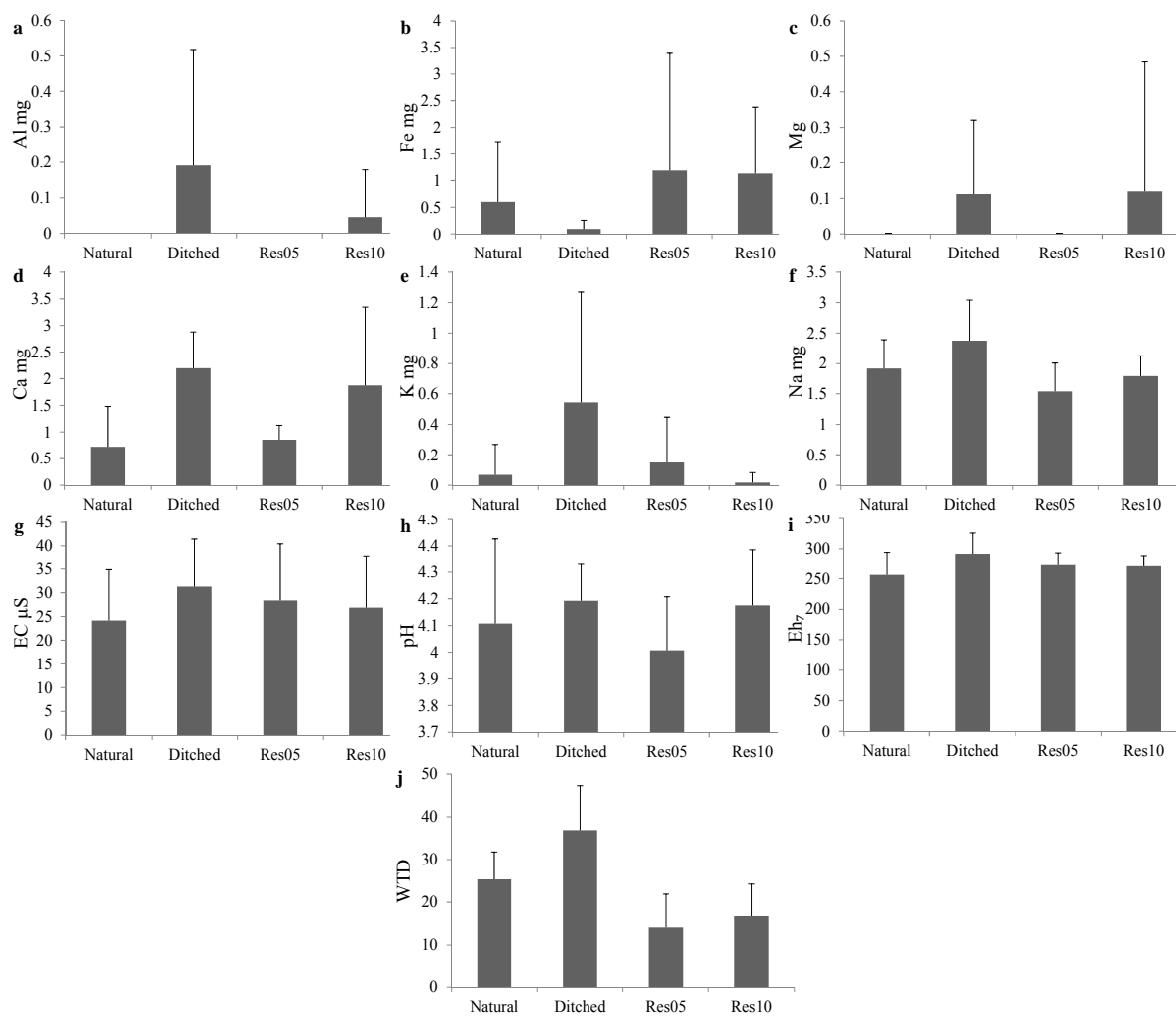
223 Water chemistry did not vary significantly between sites (Table 4). However, in some cases (mainly for Al  
224 and Mg) chemical concentrations were below the detection limits and were recorded as 0. In general, the  
225 highest mean values for studied elements were recorded at Ditched sites except for Fe (Figure 3). WTD  
226 varied between the management classes (Table 4) and between months, with the lowest depths recorded at  
227 the Ditched sites (Figure 4).

228

229 Table 4. ANOVA (\*) and Kruskal-Wallis (\*\*) for water chemistry variables. Calculations are based on  
 230 average values for each site over all management classes.

	<i>df</i>	$\chi^2/F$	<i>p</i>
*Al	3	2.47	0.481
*Fe	3	2.97	0.397
*Mg	3	4.72	0.194
*Ca	3	7.30	0.063
*K	3	5.14	0.162
**Na	3	2.75	0.079
**EC	3	0.27	0.843
**pH	3	0.89	0.470
**Eh <sub>7</sub>	3	2.06	0.148
**WTD	3	4.16	0.025

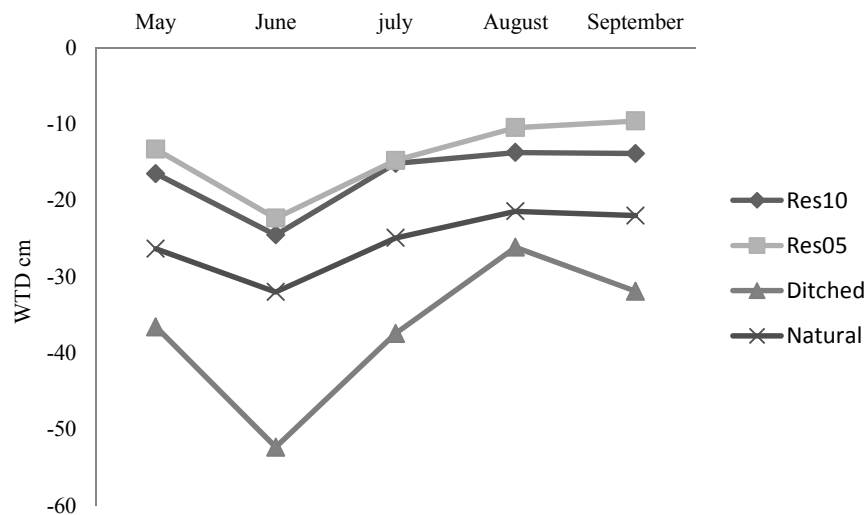
231



232

233 Figure 3. Average values and standard deviations of the environmental parameters measured for each land  
 234 use management class. (a) aluminum, (b) iron, (c) magnesium, (d) calcium, (f) potassium, (g) conductivity,

235 (h) pH, (i) redox potential, (j) water table depth. Res05: peatlands restored 3-7 years ago; Res10: peatlands  
 236 restored 9-12 years ago.

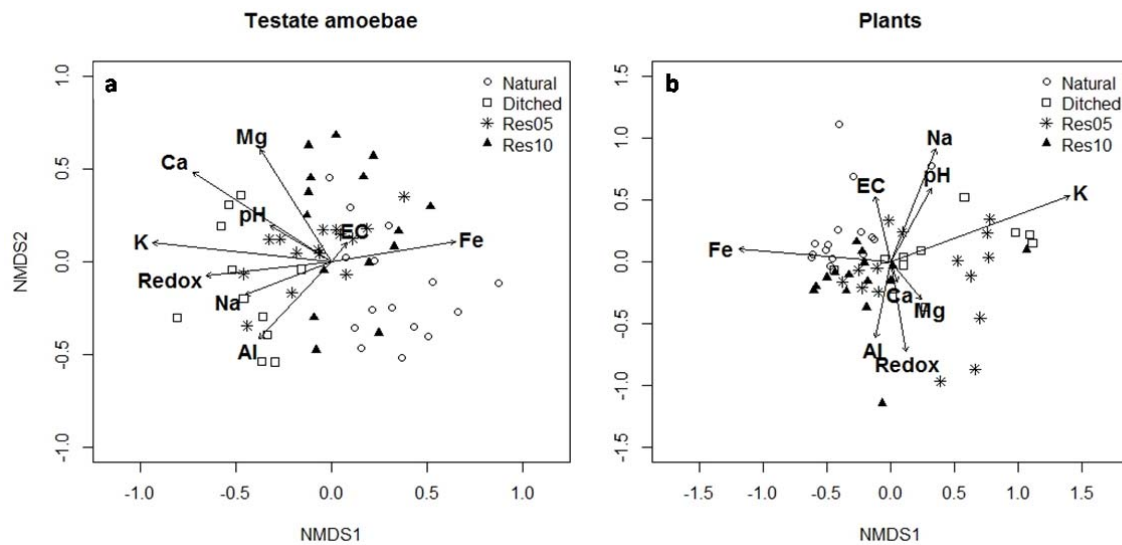


237

238 Figure 4. Monthly water table depth variation in 2007. Res05: peatlands restored 3-7 years ago; Res10:  
 239 peatlands restored 9-12 years ago.

240 A two-dimensional solution NMDS ordination of the TA communities produced the lowest stress at=  
 241 0.207. Ca and K were significantly correlated with TA community variability ( $p= 0.02$  and  $p= 0.01$ ,  
 242 respectively; Figure 5a). The ordination of the plant community resulted in a two-dimensional solution (final  
 243 stress= 0.168) with significant relation to Fe ( $p<0.01$ ), K ( $p<0.01$ ) and Na ( $p= 0.04$ ; Figure 5b).

244 NMDS plots grouped TA communities by management classes. A separation for the communities  
 245 between the Ditched and Natural sites was observed on NMDS1. Res05 sites were mainly grouped in the  
 246 centre of the ordination and overlapped with all other management classes, while Res10 sites were fully  
 247 separated from Ditched sites, and more closely grouped to Natural sites (Figure 5a). Ordination of plant  
 248 communities showed higher variations compared to TA and less clear separation along NMDS1; however,  
 249 group separation was observed for Natural and Ditched sites (upper left and central right in the ordination,  
 250 respectively). Res05 was mostly grouped closer to Ditched sites overlapping only partly with Res10, whereas  
 251 Res10 sites clustered closer to Natural sites (Figure 5b).



252

253 Figure 5. NMDS ordinations of testate amoebae (a) and plant community (a) relationships among  
 254 management classes. Distance measure: Sørensen (Bray-Curtis). Arrow lengths represent the vector loadings  
 255 of environmental variables. Res05: peatlands restored 3-7 years ago; Res10: peatlands restored 9-12 years  
 256 ago.

257 To look for concordance between plant and TA communities we first ran separate analyses to compare  
 258 TA to moss species and TA to vascular plant species (results not shown). However as no differences were  
 259 found, both moss and vascular plants were grouped to form a plant community in all subsequent analyses.  
 260 We found a significant but weak concordance between TA and plant communities (PROTEST  $r=0.44$ ,  
 261  $m^2=0.79$ ,  $p=0.001$ ). To assess whether TA and plant communities were concordant within classes, we ran  
 262 PROTEST separately for each. Results indicated that only communities at Res05 sites were concordant, but  
 263 even this relation was weak (PROTEST  $r=0.54$ ,  $m^2=0.70$ ,  $p=0.01$ ).

264

## 265 DISCUSSION

266 Natural sites showed higher TA richness with more diverse communities than other management classes.  
 267 However, we did not find similar results for plant species richness. Res10 sites showed the lowest TA  
 268 richness and diversity, which could be attributed to the high dominance of *Arcella catinus* at many of the  
 269 sites, where this species accounted for more than 70% of the community. Spatial limitation is less likely to  
 270 occur in soils compared to aboveground environments (Wanner & Xylander 2005). Hence, TA might face  
 271 less competitive pressure as different taxa may simply occupy different ecological niches, whereas plant

272 species tend to compete for the same space and nutrients. As Natural sites have not undergone extreme  
273 environmental changes compared to the other management classes, their higher taxon richness could be  
274 attributed to a longer time for cumulative colonisation without a significant TA taxon replacement. This  
275 supports the observations by Wanner & Xylander (2005) who studied TA colonisation and successional  
276 processes, and found temporal changes in TA species composition but no or very little species replacement.

277 Restoration efforts to rehabilitate peatlands focus mainly on raising the water table to recreate natural  
278 habitat conditions. Substratum moisture is often found to be the main factor controlling TA communities  
279 (Tolonen 1986) particularly in *Sphagnum* dominated peatlands (Booth 2001) with higher densities of TA  
280 cells usually found in soils with a high water holding capacity (Fournier *et al.* 2012). Despite the lack of  
281 corresponding moisture data, we assume that restoration measures increased peat moisture due to elevated  
282 water table levels. As a consequence, increasing TA cell densities should be expected at restored sites over  
283 time. We did observe such patterns, as the highest TA densities were found at Res10 sites and the lowest at  
284 Ditched sites.

285 Testate amoebae communities at the wettest sites (Natural, Res10, and Res05) were mostly dominated by  
286 *A. catinus* and *B. indica* while at Ditched sites, taxon dominance shifted towards *T. arcula*. However, both *A.*  
287 *catinus* and *B. indica* were highly abundant across all sites suggesting that despite these species are  
288 considered indicators of dry conditions, they can display a broad moisture tolerance. Some less abundant  
289 species, such as *A. artocrea*, *C. ecornis*, *D. leidyi*, *D. lithophila*, *H. minuta*, and *P. fascicularis*, were found  
290 exclusively at Natural sites. This suggests that not all species recolonize even within a decade after  
291 restoration. Poor re-colonization of some typical plant species after restoration (Haapalehto *et al.* 2011,  
292 Hedberg *et al.* 2012) highlights the need to avoid degradation of natural habitats, as species that disappear  
293 from a site may be very difficult to reinstate without costly and uncertain re-introduction programs (see also  
294 Moreno-Mateos *et al.* 2015). *H. elegans* and *Ph. griseola* only occurred at Natural sites and sites restored 10  
295 years ago, suggesting that they either have a longer recolonization time, or that disturbed habitats are just not  
296 appropriate for those species. Among plant species, *S. angustifolium* remained dominant across all sites. This  
297 species has a wide ecological niche, and is known to survive in drained peatlands. However, *S. angustifolium*  
298 densities increased rapidly following restoration measures (Haapalehto *et al.* 2011).

299 Ordinations showed significant relationship of TA communities with Ca and K. Strong relationships  
300 between Ca and TA communities have been reported before (e.g. Lamentowicz *et al.* 2010, Hájková *et al.*

2011, Raabe & Lamentowicz 2012, Jassey *et al.* 2014). These could be due to the importance of Ca for the shell building process of some TA species (Lamentowicz *et al.* 2011). TA relationships with other nutrients have also been observed (e.g. Mitchell *et al.* 2000a, Jauhiainen 2002, Mitchell *et al.* 2004) but their direct effects on TA are not fully understood. Such correlations could arise when water chemistry and nutrient availability indirectly affect TA communities by controlling their prey organisms such as bacteria, fungi, other Protista, and micrometazoa (Mitchell *et al.* 2004).

Plants were strongly related to levels of K and Fe, indicating the vital importance of pore water chemistry. The highest K concentrations were found at Res05 sites, and the lowest in Natural and Res10. As an easily leachable cation, K may be released from dying biomass and rewetted organic material after hydrological restoration. Hence the lower concentrations of K in pore water at Res10 sites might indicate that K is efficiently taken up during the regrowth of *Sphagnum* and other peatland species. Additionally, after ditch filling small ponds, topographically lower areas are formed at restored sites enabling the establishment of anaerobic and reducing conditions that could favour reduction of Fe<sup>3+</sup> to Fe<sup>2+</sup> (Haapalehto *et al.* 2014). This reduced form of Fe is readily available for uptake by plants and facilitates plant regrowth.

In NMDS ordination vegetation communities displayed more within-class variation compared to TA. While TA communities differed between Ditched & Res5 (PerMANOVA) plants did not show differences. These results suggest that TA taxa might be better suited than plants as indicators of early restoration success thanks to their rapid response to hydrological variations. This indeed was observed by Warner & Chmielewski (1992) who found changes in TA taxa composition within 2½ years of peatland drainage. Further, also Talbot *et al.* (2010) who studied peat cores of drained peatlands found that TA species indicative of dry conditions reached their highest percentages around the time of drainage, suggesting a very quick response potential of the TA communities to environmental disturbance. Our results highlight the severity of hydrological disturbance by drainage as TA communities still displayed clear differences from those at Natural sites even a decade after restoration. Observed long-term drainage impacts on TA are paralleled by changes in plant communities after drainage (Laine *et al.* 1995, Kareksela *et al.* 2015). From visual inspection of NMDS, vegetation communities 10 years after restoration appeared more similar to Natural sites. Further, Maanaviilja *et al.* (2014) suggested that only a large rise in the water table level would induce significant changes in vegetation communities. Indeed, in our study plants were sampled 15m from the ditch where water table changes to drainage and restoration are smallest and where plant community



330 compositions still largely overlap (Haapalehto *et al.* 2015, Kareksela *et al.* 2015). Finally, Wardle (2002)  
331 pointed out that aboveground vegetation communities and soil microorganisms differ in their resistance,  
332 resilience, adaptation strategies, and dispersal abilities. These differences might also relate to the observed  
333 time lags in responses of TA and plant communities to restoration measures.

334 When comparing all sites, TA and vegetation communities were concordant although this relationship  
335 was weak. When each management class was separately analysed we found no within management class  
336 community concordance except for Res05 sites. Some studies on stream community concordance (e.g.  
337 Infante *et al.* 2009) have suggested a scale dependency, which could explain the observed patterns. However,  
338 the fact that TA and vegetation community concordance was found to be weak, and the apparent lack of  
339 within-class community concordance suggest that TA and vegetation community composition are  
340 determined in different ways except during early post-restoration. Our results thus suggest that either overall  
341 communities i) respond differently to the same set of environmental factors, or ii) are affected by different  
342 environmental factors over timespans exceeding 5 years, or iii) have different time scales of succession  
343 processes, and/or iv) display low overall interaction. Additionally, Hájek *et al.* (2014) from their comparison  
344 of vascular plants, bryophytes, fungi, diatoms, desmids, and TA suggested that body size and life span play a  
345 key role when determining concordance between communities.

346 In peatland ecosystems, microtopographic transitions can occur at scales of a few centimetres reducing  
347 the ability to avoid ecological gradients when sampling. Even though microtopographic transitions at the  
348 centimetre scale are specifically important when designing the sampling, even smaller gradients may still  
349 exist at the scale of TA (Mitchell *et al.* 2000b). This vertical variation is mainly caused by differences in the  
350 chemical composition of ground water and rainwater and is enhanced by *Sphagnum* cation exchange  
351 (Mitchell *et al.* 2000a). As moss species and TA are dependent on microsite conditions, they might reflect  
352 different conditions than vascular plants, which are rooted at different depths (Mitchell *et al.* 2000a).  
353 However in separate analyses to compare TA to moss species and TA to vascular plant species we found no  
354 differences in concordance patterns (results not shown). Restoration strategies such as rewetting of drained  
355 peatlands aim mainly at restoring the vegetation coverage of sites (e.g. Similä *et al.* 2014). To this end, plant  
356 communities are obvious indicators of restoration success. However, our results suggest that restoration  
357 effects on other components (e.g. TA communities) in peatlands might not be reflected properly by the use of  
358 vegetation-based indicators alone. Gilbert *et al.* (1998) studied the microbial loop in peatlands and revealed

359 that TA can account for almost half of the microbial community using a wide range of organisms as prey.  
360 Through their feeding TA regulate bacterial biomass, contribute to nutrient mineralisation, nutrition of soil  
361 animals, and plant growth. Griffiths (1994) found that, in general, Protista and mainly TA can account for  
362 20–40% of N mineralisation as they excrete bacterial N into the soil. As the TA communities of Res05 sites  
363 were more similar to the targeted original ecosystem than Ditched ones, restoration by rewetting appears to  
364 be efficient in inducing a rapid recovery of an important structural ecosystem component. While the  
365 relationship is still poorly understood, previous studies suggest that recovery of some important ecosystem  
366 functions like surface peat accumulation precedes the recovery of ecosystem structure (Kareksela *et al.*  
367 2015). In accordance with that study, our results suggest that the recovery of TA community structure  
368 precedes that of plants and in fact may even be a prerequisite for the recovery of higher plants and ecosystem  
369 functions. Thus, TA might play a fundamental role in the facilitation of plant succession by providing  
370 nutrients and consolidating the assembly process on newly exposed land surfaces (Hodkinson *et al.* 2002).

371 In conclusion, despite some similarities in early responses, TA and vegetation communities cannot be  
372 used as surrogates of each other when measuring restoration success. To gain a holistic understanding of the  
373 changes in important ecosystem components during restoration from an ecosystem perspective requires the  
374 use of both vegetation and TA communities as also suggested by Raabe and Lamentowicz (2012). However,  
375 the shorter generation time of TA compared to vegetation enable them to respond faster to environmental  
376 changes making them better suited as early indicators of restoration success particularly because post-  
377 restoration successional changes in TA communities appear to be targeted towards the re-establishment of  
378 the site's original community composition. Our results also suggest that further studies on the relationship  
379 between TA and plant communities could provide important insights into understanding the crucial link  
380 between the recovery of ecosystem structure and functions. Better assessment of the dynamics and interplay  
381 of the post-restoration re-colonisation processes of the two communities in the boreal region will require  
382 studies spanning longer time scales than were studied here.

383

#### 384 ACKNOWLEDGMENTS

385 Thanks to Dan Charman and Matthew Amesbury for their guidance on testate amoebae taxonomy and data  
386 analysis, and to Heikki Hämäläinen for his valuable comments on the community concordance  
387 interpretations. Thanks to Santtu Kareksela and Riikka Juutinen who collected the plant community data and

388 to Rose Matilainen who analysed the pore water data at the Department of Chemistry (University of  
389 Jyväskylä). Special thanks to Roger Jones for language corrections and his valuable input into this work.  
390 Emmanuela Daza Secco thanks the Maj and Tor Nessling foundation for financial support.

391

## 392 REFERENCES

393 Aapala, K., Heikkilä, R. & Lindholm T. (1996) Protecting the diversity of Finnish mires. In: Vasander, H  
394 (ed). *Peatlands in Finland*, Finnish Peatland Society, Jyväskylä, Finland, 45–57.

395

396 Bilton, D.T., MacAbendroth, L., Bedford, A. & Ramsay, P.M. (2006) How wide to cast the net? Cross-taxon  
397 congruence of species richness, community similarity and indicator taxa in ponds. *Freshwater Biology*, 51,  
398 578–590.

399

400 Bobrov, A.A., Charman, D.J. & Warner, B.G. (1999) Ecology of testate amoebae (Protozoa: Rhizopoda) on  
401 peatlands in western Russia with special attention to niche separation in closely related taxa. *Protist*, 150,  
402 125–136.

403

404 Bobrov, A.A., Charman, D.J. & Warner, B.G. (2002) Ecology of Testate Amoebae from Oligotrophic  
405 Peatlands: Specific Features of Polytypic and Polymorphic Species. *Biology Bulletin*, 29, 605–617.

406

407 Booth, R.K. (2001) Ecology of testate amoebae (Protozoa) in two lake superior coastal wetlands:  
408 implications for paleoecology and environmental monitoring. *Wetlands*, 21, 564–576.

409

410 Booth, R.K. (2008) Testate amoebae as proxies for mean annual water-table depth in *Sphagnum*-dominated  
411 peatlands of North America. *Journal of Quaternary Science*, 23, 43–57.

412

413 Booth, R.K., Lamentowicz, M. & Charman, D.J. (2010) Preparation and analysis of testate amoebae in  
414 peatland palaeoenvironmental studies. *Mires and peat*, 7, 1–7.

415

- 416 Charman, D.J. (1997) Modelling hydrological relationships of testate amoebae (Protozoa: Rhizopoda) on  
417 New Zealand peatlands. *Journal of the Royal Society of New Zealand*, 27, 465–483.  
418
- 419 Charman, D.J., Hendon, D. & Woodland, W.A. (2000) *The identification of testate amoebae (Protozoa:*  
420 *Rhizopoda) in peats*. Quaternary Research Association, U.K., Technical Guide No. 9, 147 pp.  
421
- 422 Clarke, K.J. (2003) *Guide to the Identification of Soil Protozoa - Testate Amoebae*. Soil Biodiversity  
423 Programme Research, Fresh Water Biological Association, Ambleside, U.K., Report No.4, 40pp.  
424
- 425 Costanza, R. & Mageau M. (1999) What is a healthy ecosystem? *Aquatic Ecology*, 33(1), 105–115.  
426
- 427 Davis, S.R. & Wilkinson, D.M. (2004) The conservation management value of testate amoebae as  
428 “restoration” indicators: speculations based on two damaged raised mires in northwest England. *The*  
429 *Holocene*, 14, 135–143.  
430
- 431 Dobson, A.P., Bradshaw, A.D. & Baker, A.J.M. (1997) Hopes for the Future: Restoration Ecology and  
432 Conservation Biology. *Science*, 277, 515–522.  
433
- 434 Fournier, B., Malysheva, E., Mazei, Y., Moretti, M. & Mitchell, E.A.D. (2012) Toward the use of testate  
435 amoebae functional traits as indicator of floodplain restoration success. *European Journal of Soil Biology*,  
436 49, 85–91.  
437
- 438 Gilbert, D., Amblard, C., Bourdier, G. & Francez, A.J. (1998) The microbial loop at the surface of a  
439 peatland: structure, function, and impact of nutrient input. *Microbial Ecology*, 35, 83–93.  
440
- 441 Griffiths, B.S. (1994) Microbial-feeding nematodes and protozoa in soil: their effects on microbial activity  
442 and nitrogen mineralization in decomposition hotspots and the rhizosphere. *Plant and Soil*, 164, 25–33.  
443

- 444 Haapalehto, T., Vasander, H., Jauhiainen, S., Tahvanainen, T. & Kotiaho, J.S. (2011) The effects of peatland  
445 restoration on water-table depth, elemental concentrations, and vegetation: 10 years of changes. , 19, 587–  
446 598.
- 447
- 448 Haapalehto, T. (2014) Restoring ecosystem structure and functions: results from *Sphagnum* peatlands  
449 degraded by forestry drainage. PhD Dissertation, University of Jyväskylä, Jyväskylä, 30pp.
- 450
- 451 Haapalehto, T., Kotiaho, J.S., Matilainen, R. & Tahvanainen, T. (2014) The effects of long-term drainage  
452 and subsequent restoration on water table level and pore water chemistry in boreal peatlands. *Journal of*  
453 *Hydrology*, 519, 1493–1505.
- 454
- 455 Haapalehto, T., Kotiaho, J.S., Matilainen, R. & Tahvanainen, T. (2015) The effects of long-term drainage  
456 and subsequent restoration on water table level and pore water chemistry in boreal peatlands. *Journal of*  
457 *Hydrology*, 519, 1493–1505.
- 458
- 459 Hájek, M., Pouličková, A., Vašutová, M., Surovátka, V., Jiroušek, M., Štěpánková, J., Opravilová, V. &  
460 Hájková, P. (2014) Small ones and big ones: cross-taxon congruence reflects organism body size in  
461 ombrotrophic bogs. *Hydrobiologia*, 726, 95–107.
- 462
- 463 Hájková, P., Bojková, J., Fránková, M., Opravilová, V., Hájek, M., Kintrová, K. & Horsák, M. (2011)  
464 Disentangling the effects of water chemistry and substratum structure on moss-dwelling unicellular and  
465 multicellular micro-organisms in spring-fens. *Journal of Limnology*, 70(1), 54–64.
- 466
- 467 Hedberg, P., Kotowski, W., Saetre, P., Mälson, K., Rydin, H. & Sundberg, S. (2012) Vegetation recovery  
468 after multiple-site experimental fen restorations. *Biological Conservation*, 147, 60–67.
- 469
- 470 Heino, J. (2002) Concordance of species richness patterns among multiple freshwater taxa: a regional  
471 perspective. *Biodiversity and conservation*, 11,137–147.
- 472

- 473 Hodkinson, I.D., Webb, N.R. & Coulson, S.J. (2002) Primary community assembly on land—the missing  
474 stages: why are the heterotrophic organisms always there first? *Journal of Ecology*, 90, 569–577.  
475
- 476 Hunter Jr, M., Westgate, M., Barton, P., Calhoun, A., Pierson, J., Tulloch, A., Beger, M., Branquinho, C.,  
477 Caro, T., Gross, J., Heino, J., Lane, P., Longo, C., Martin, K., McDowell, W.H., Mellin, C., Salo, H. &  
478 Lindenmayer, D. (2016) Two roles for ecological surrogacy: Indicator surrogates and management  
479 surrogates. *Ecological indicators*, 63, 121–125.  
480
- 481 Infante D.M., Allan, J.D., Linke, S. & Norris, R.H. (2009) Relationship of fish and macroinvertebrate  
482 assemblages to environmental factors: implications for community concordance. *Hydrobiologia*, 623, 87–  
483 103.  
484
- 485 Jassey, V.E.J., Lamentowicz, L., Roborek, B.J.M., Gabka, M., Rusinska, A. & Lamentowicz, M. (2014)  
486 Plant functional diversity drives niche-size-structure of dominant microbial consumers along a poor to  
487 extremely rich fen gradient. *Journal of Ecology*, 102, 1150–1162.  
488
- 489 Jauhiainen, S. (2002) Testacean amoebae in different types of mire following drainage and subsequent  
490 restoration. *European Journal of Protistology*, 38, 59–72.  
491
- 492 Jyväsjärvi, J., Järvinen, M. & Hämäläinen, H. (2014) Spatial community concordance of summer  
493 phytoplankton and profundal macroinvertebrates in boreal lakes. *Canadian Journal of Fisheries and Aquatic  
494 Sciences*, 71, 1776–1783.  
495
- 496 Kareksela, S., Haapalehto, T., Juutinen, R., Matilainen, R., Tahvanainen, T. & Kotiaho, J.S. (2015) Fighting  
497 carbon loss of degraded peatlands by jump-starting ecosystem functioning with ecological restoration.  
498 *Science of the Total Environment*, 537, 268–276.  
499

- 500 Koenig, I., Feldmeyer-Christe, E. & Mitchell, E.A.D. (2015) Comparative ecology of vascular plant,  
501 bryophyte and testate amoeba communities in four *Sphagnum* peatlands along an altitudinal gradient in  
502 Switzerland. *Ecological Indicators*, 45, 48–59.
- 503
- 504 Laine, J., Vasander, H. & Laiho, R. (1995) Long-Term Effects of Water Level Drawdown on the Vegetation  
505 of Drained Pine Mires in Southern Finland. *Journal of Applied Ecology*, 32, 785–802.
- 506
- 507 Laine, A.M., Leppälä, M., Tarvainen, O., Päätaalo, M.L., Seppänen, R. & Tolvanen, A. (2011) Restoration of  
508 managed pine fens: effect on hydrology and vegetation. *Applied Vegetation Science*, 14, 340–349.
- 509
- 510 Lamentowicz, M., Lamentowicz, L., van der Knaap, W.O., Gbka, M. & Mitchell, E.A.D. (2010) Contrasting  
511 species-environment relationships in communities of testate amoebae, bryophytes and vascular plants along  
512 the fen-bog gradient. *Microbial Ecology*, 59, 499–510.
- 513
- 514 Lamentowicz, L., Gabka, M., Rusinska, A., Sobczynsky, T., Owsiany, P.M. & Lamentowicz, M. (2011)  
515 Testate amoeba (Arcellinida, Euglyphida) Ecology along a poor-rich gradient in fens of western Poland.  
516 *International review of hydrobiology*, 96, 356–380.
- 517
- 518 Lappalainen, E. (1996) General review on world peatland and peat resources. In: Lappalainen, E. (ed) *Global*  
519 *peat resources*. International Peat Society, Jyväskylä, Finland, 53–56.
- 520
- 521 Lunn, A. & Burlton, B. (2013) The border Mires: a completed peatland restoration project. *British Wildlife*,  
522 24:153–160.
- 523
- 524 Maanavilja, L., Aapala, K., Haapalehto, T., Kotiaho, J.S. & Tuittila, E.S. (2014) Impact of drainage and  
525 hydrological restoration on vegetation structure in boreal spruce swamp forests. *Forest ecology and*  
526 *Management*, 330, 115–125.
- 527
- 528 Marcisz, K., Fournier, B., Gilbert, D., Lamentowicz, M. & Mitchell, E.A.D. (2014) Response of *Sphagnum*

- 529 peatland testate amoebae to a 1-year transplantation experiment along an artificial hydrological gradient.  
530 *Microbial Ecology*, 67, 810–818.
- 531 Mazei, Y. & Tsyganov, A. (2006) Freshwater testate amoebae. 300pp. (in Russian).  
532
- 533 Meisterfeld, R. (2002) Order Arcellinida Kent, 1880. In: Lee, J.J., Leedale, G.F. & Bradbury, P. (eds) *The*  
534 *Illustrated guide to the Protozoa Vol. 2*. Society of Protozoologists, Lawrence, Kansas, 827-860.  
535
- 536 Mitchell, E.A.D., Buttler, A., Grosvernier, Ph., Rydin, H., Albinsson, C., Greenup, A.L., Heijmans,  
537 M.M.P.D., Hoosbeek, M.R. & Saarinen, T. (2000a). Relationships among testate amoebae (Protozoa),  
538 vegetation and water chemistry in five *Sphagnum*-dominated peatlands in Europe. *New Phytologist*, 145, 95–  
539 106.  
540
- 541 Mitchell, E.A.D., Borcard, D., Buttler, A.J., Grosvernier, Ph. & Gilbert, D.J.M. (2000b). Horizontal  
542 distribution patterns of testate amoebae (Protozoa) in a *Sphagnum magellanicum* carpet. *Microbial Ecology*  
543 39: 290–300.  
544
- 545 Mitchell, E.A.D. (2003a) The identification of *Centropyxis*, *Cyclopyxis*, *Trigonopyxis* and similar  
546 *Phryganella* species living in *Sphagnum*. Unpublished.  
547
- 548 Mitchell, E.A.D. (2003b) The identification of *Nebela* and similar species with indications on their ecology  
549 and distribution. Unpublished.  
550
- 551 Mitchell, E.A.D., Bragazza, L. & Gerdol, R. (2004) Testate amoebae (Protista) communities in *Hylocomium*  
552 *splendens* (Hedw.) B.S.G. (Bryophyta): relationships with altitude, and moss elemental chemistry. *Protist*  
553 155, 423–436.  
554
- 555 Moreno-Mateos, D., Meli, P., Vara-Rodríguez, M.I. & Aronson, J. (2015) Ecosystem response to  
556 interventions: lessons from restored and created wetland ecosystems. *Journal of Applied Ecology*, 52(6),



557 1528–1535.

558

559 Mykrä, H., Aroviita, J., Hämäläinen, H., Kotanen, J., Vuori, K.M. & Muotka, T. (2008) Assessing stream  
560 condition using macroinvertebrates and macrophytes: concordance of community responses to human  
561 impact. *Fundamental and Applied Limnology*, 172, 191–203.

562

563 Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O’Hara, R.B., Simpson, G.L., Solymos,  
564 P., Stevens, M.H. & Wagner, H. (2015) Vegan: community ecology package. Version 2.3–0.

565 Poulin, M., Andersen, R. & Rochefort, L. (2013) A new approach for tracking vegetation change after  
566 restoration: A case study with peatlands. *Restoration Ecology*, 21, 363–371.

567

568 Paavola, R., Muotka, T., Virtanen, R., Heino, J. & Kreivi, P. (2003) Are biological classifications of  
569 headwater streams concordant across multiple taxonomic groups? *Freshwater Biology*, 48, 1912–1923.

570

571 Paavola, R., Muotka, T., Virtanen, R., Heino, J., Jackson, D. & Maki-Petays, M. (2006) Spatial scale affects  
572 community concordance among fishes, benthic macroinvertebrates, and bryophytes in streams. *Ecological  
573 Applications*, 16, 368–379.

574

575 Pawar, S.S., Birand, A.C., Ahmed, M.F., Sengupta, S. & Raman, T.R.S. (2007) Conservation Biogeography  
576 in North-east India: hierarchical analysis of cross-taxon distributional congruence. *Diversity and  
577 Distributions*, 13, 53–65.

578

579 Peres-Neto, P. & Jackson, D.A. (2001) How well do multivariate data sets match? the advantages of a  
580 Procrustean superimposition approach over the Mantel test. *Oecologia*, 129, 169–178.

581

582 R Core Team (2013) R: A language and environment for statistical computing. R Foundation for Statistical  
583 Computing, Vienna, Austria. ISBN 3-900051-07-0, URL <http://www.R-project.org/>

584

585 Raabe, P. & Lamentowicz, M. (2012) Testate amoebae reflecting present environmental conditions in

- 586 restored cut-over bogs - a new tool for evaluation and monitoring? 14th International Peat Congress, pp. 1-6.  
587
- 588 Schönborn, W. (1986) Population dynamics and production biology of testate amoebae (Rhizopoda,  
589 Testacea) in raw humus of two coniferous forest soils. *Archiv für Protistenkunde*, 132, 325–342.
- 590
- 591 Shurin, J.B., Cottenie, K. & Hillebrand, H. (2009) Spatial autocorrelation and dispersal limitation in  
592 freshwater organisms. *Oecologia*, 159, 151–159.
- 593
- 594 Similä, M., Aapala, K. & Penttinen, J. (2014) *Ecological restoration in drained peatlands-best practices in*  
595 *Finland*. Metsähallitus, Natural Heritage Services, Vantaa, 84 pp.
- 596
- 597 Sleigh, M.A. (1989) *Protozoa and other protists 2nd edition*. Edward Arnold, London. U.K. 343pp.
- 598
- 599 Stockmarr, J. (1971) Tablets with spores used in absolute pollen analysis. *Pollen et Spores* 13, 615–21.  
600
- 601 Tahvanainen, T. (2011) Abrupt ombrotrophication of a boreal aapa mire triggered by hydrological  
602 disturbance in the catchment. *Journal of ecology*, 99(2), 404–415.
- 603
- 604 Tolonen, K. (1986) Rhizopod analysis. In: Berglund, B.E. (ed) *Handbook of the Holocene Paleoecology and*  
605 *Paleohydrology*. J. Wiley Sons, Chichester, U.K, 645–666.
- 606
- 607 Trepel, M. (2007) Evaluation of the implementation of a goal-oriented peatland rehabilitation plan.  
608 *Ecological Engineering*, 30(2), 167–175.
- 609
- 610 Turner, T.E. & Swindles, G.T. (2012) Ecology of testate amoebae in moorland with a complex fire history:  
611 implications for ecosystem monitoring and sustainable land management. *Protist*, 163, 844–855.
- 612

- 613 Vasander, H., Tuittila, E.S., Lode, E., Lundin, L., Llomets, M., Sallantaus, T., Heikkila, R., Pitkanen, M.L. &  
614 Laine, J. (2003) Status and restoration of peatlands in northern Europe. *Wetlands Ecology and Management*,  
615 11, 51–63.
- 616
- 617 Wanner, M. & Xylander, W.E.R. (2005) Biodiversity development of terrestrial testate amoebae: Is there any  
618 succession at all? *Biology of Fertil Soils*, 41, 428–438.
- 619
- 620 Wardle, D.A. (2002) Communities and ecosystems: linking the above-ground and below-ground  
621 components. *Austral Ecology*, 29, 358–362.
- 622
- 623 Warner, B.G. & Chmielewski, J.G. (1992) Testate amoebae (Protozoa) as indicators of drainage in a forested  
624 mire, Northern Ontario, Canada. *Archiv für Protistenkunde*, 141, 179–183.
- 625
- 626 Warner, B.G., Asada, T. & Quinn, N.P. (2007) Seasonal influences on the ecology of testate amoebae  
627 (Protozoa) in a small Sphagnum peatland in Southern Ontario, Canada. *Microbial Ecology*, 54, 91–100.
- 628
- 629 Yates, A.G. & Bailey, R.C. (2010) Covarying patterns of macroinvertebrate and fish assemblages along  
630 natural and human activity gradients: implications for bioassessment. *Hydrobiologia*, 637, 87–100.
- 631
- 632 Yu, Z. (2011) Holocene carbon flux histories of the world's peatlands: Global carbon-cycle implications. *The*  
633 *Holocene*, 21, 761–774.
- 634
- 635 Yu, Z. (2012) Northern peatland carbon stocks and dynamics: a review. *Biogeosciences*, 9, 4071–4085.
- 636
- 637 Author of correspondence:
- 638 M.Sc. Emmanuela Daza Secco, Finnish Environment Institute, Surfontie 9A, Jyväskylä 40500, Finland.
- 639 Email: [emdazase@student.jyu.fi](mailto:emdazase@student.jyu.fi). (+57) 350-2718987.