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**Title:** Bioinspired Mo, W and V complexes bearing a highly hydroxyl-functionalized Schiff base ligand

**Year:** 2020

**Version:** Accepted version (Final draft)

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**Please cite the original version:**

Salonen, P., & Lehtonen, A. (2020). Bioinspired Mo, W and V complexes bearing a highly hydroxyl-functionalized Schiff base ligand. *Inorganica Chimica Acta*, 503, Article 119414. <https://doi.org/10.1016/j.ica.2020.119414>

# 1 **Bioinspired Mo, W and V complexes bearing a highly hydroxyl-functionalized** 2 **Schiff base ligand**

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9

## 10 **Abstract**

11 A series of bioinspired dioxidomolybdenum(VI), dioxidotungsten(VI) and oxidovanadium(V) complexes [MoO<sub>2</sub>(H<sub>2</sub>L<sup>Saltris</sup>)], [WO<sub>2</sub>(H<sub>2</sub>L<sup>Saltris</sup>)] and [VO(HL<sup>Saltris</sup>)]<sub>2</sub> were prepared  
12 by the reaction of a hydroxyl-rich Schiff base proligand *N*-(1,3-dihydroxy-2-(hydroxymethyl)propan-2-yl)-3,5-di-*tert*-butylsalicylaldimine (H<sub>4</sub>L<sup>Saltris</sup>) with metal  
13 precursors in methanol solutions. Molybdenum and tungsten complexes crystallize as  
14 mononuclear molecules, whereas the vanadium complex forms dinuclear units. From the  
15 complexes, [VO(HL<sup>Saltris</sup>)]<sub>2</sub> shows activity in the oxidation of 4-*tert*-butylcatechol and 3,5-  
16 di-*tert*-butylcatechol, mimicking the action of the dicopper enzyme catechol oxidase.\*  
17  
18

## 19 **Introduction**

20 The easy exploitation of aerial dioxygen in the context of oxidation catalysis represents  
21 one of the most desired goals in synthetic organic chemistry because of its ready  
22 availability, low cost and environmental compatibility.[1] Nature has perfected the use of  
23 the relatively inert dioxygen molecule in a wide variety of metabolic oxidation reactions  
24 through metalloenzymes. Efforts to mimic various metalloenzyme active sites by  
25 (bio)inorganic chemists can be divided into biomimetic (structural) and bioinspired  
26 (functional) modelling.[2,3] A central idea in coordination chemistry is to use ligands  
27 with diverse electronic and steric features to modulate the properties of central metals  
28 in various model complexes, and thus achieve some properties related to natural enzyme  
29 active sites.

30 The enzymes from the molybdenum oxotransferase superfamily catalyze oxygen atom  
31 transfer (OAT) reactions, whereby an oxygen atom is transferred from a high-valent,  
32 catalytically active (di)oxidomolybdenum species to a suitable donor/acceptor  
33 molecule.[4,5] The superfamily can be divided into three subfamilies, namely xanthine  
34 oxidase, sulfite oxidase and DMSO reductase.[6] The active sites of the eponymous  
35 enzymes DMSO reductase and sulfite oxidase consist of a monooxido- and dioxidomolybdenum(VI) centers, respectively.[6] They, together with many other molybdoenzymes

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List of abbreviations: 3,5-DTBC (3,5-di-*tert*-butylcatechol), 3,5-DTBSQ (3,5-di-*tert*-butylsemiquinone), 3,5-DTBQ (3,5-di-*tert*-butyl-1,2-benzoquinone), 4-TBC (4-*tert*-butylcatechol), 4-TBQ (4-*tert*-butyl-1,2-benzoquinone), OAT (oxygen atom transfer).

1 have been structurally and functionally modelled by a number of monooxido- and  
2 dioxidomolybdenum(VI)- and tungsten(VI) complexes, which find applications in OAT  
3 catalysis research.[7]

4 Vanadates and other vanadium-containing compounds have been studied for decades  
5 due to their biological relevance and catalytic activity in a variety of oxidation  
6 reactions.[8–11] Especially, the bioinorganic chemistry of vanadium, highlighted by the  
7 three vanadium haloperoxidases and V-nitrogenase, as well as the anti-diabetic effects of  
8 many vanadium compounds have motivated synthetic chemists to prepare model  
9 compounds for use in oxidation catalysis, as well as in structural studies, and in medicinal  
10 research.[12–14] Catechol oxidase is an dioxygen-activating dicopper enzyme[15–17]  
11 which catalyzes the oxidation of catechols to the corresponding *o*-benzoquinones,  
12 producing water as a by-product. A number of vanadium(IV) and vanadium(V) complexes  
13 have also been discovered to efficiently catalyze the same reaction.[18–23]

14 Although compounds that display catechol oxidase mimetic activities are usually  
15 reserved for of Cu coordination model compounds[24,25], recently many other metals  
16 *e.g.* Co, Mn, Fe, Ni and Zn, as substitutes for copper, have also been used to reach moderate  
17 to high conversions of various catechols to *o*-benzoquinones.[26–28] There are also some  
18 examples on the catechol oxidase mimicking activity of molecular vanadium(IV) and  
19 vanadium(V) complexes.[18–23] It is well established that various catechols readily form  
20 redox non-innocent complexes with vanadium, where the vanadium center may be  
21 assigned +III, +IV and +V oxidation states.[29–33] There are also some reports for  
22 catecholate-containing molybdenum and even tungsten complexes.[34] In the case of  
23 vanadium(V) complexation of catechols may lead to a reduction in the metal center (V(V)  
24 → V(IV)), sometimes accompanied by cleavage of the oxido ligand.[30,31,33]

25 The mechanism(s) of vanadium-catalyzed oxidation of catechols are still not fully  
26 understood. For example, many vanadium systems apparently demonstrate catechol  
27 oxidase- and catechol dioxygenase-like activities simultaneously, leading to a number of  
28 oxidation products ranging from intra- and extradiol cleavage products to autoxidation  
29 products.[35,36] Furthermore, there is no definitive consensus on the nature of the active  
30 catalyst, with some proponents in support of a “common catalyst hypothesis”, which  
31 states that a specific vanadium-containing, catalytically active species is always formed  
32 *in-situ* by vanadium-leaching regardless of used vanadium catalyst precursor.[36] This is  
33 evidenced by very similar product distributions forming by the action of very different  
34 precatalysts, as well as from indicative spectroscopic and spectrometric signatures.

35 In light of available data[37], the common catalyst has been suggested to be [VO(3,5-  
36 DTBC)(3,5-DTBSQ)], which is formed during catalytic turnover conditions from its  
37 catalytic resting-state [VO(3,5-DTBC)(3,5-DTBSQ)]<sub>2</sub>, also known as Pierpont’s  
38 complex.[29] The Pierpont’s complex itself is derived from H<sub>2</sub>O<sub>2</sub>-assisted leaching of  
39 virtually any vanadium catalyst precursor, provided that 3,5-DTBC is present, in a so-  
40 called autoxidation-product-initiated reaction pathway.[36,38] However, there are also  
41 some reports of discrete oxidovanadium(V) catalysts where the supposed active catalyst,

1 allegedly not formed from metal leaching, has been identified using mass  
2 spectrometry.[20,23]

3 Herein we describe the synthesis and characterization of dioxidomolybdenum(VI) and -  
4 tungsten(VI) complexes (**1**, **2**) as well as the oxidovanadium(V) complex (**3**) and their  
5 application as bioinspired synthetic analogues to molybdenum oxotransferases and  
6 catechol oxidase. The syntheses of **1-3** involve complexation to a bulky Schiff base  
7 proligand *N*-(1,3-dihydroxy-2-(hydroxymethyl)propan-2-yl)-3,5-di-*tert*-butylsalicylal-  
8 imine ( $H_4L^{Saltris}$ ) featuring several alcohol side-arms. The presence of non-coordinating  
9 hydrogen bond/donor acceptor moieties such as alcohols or amides in the outer  
10 coordination spheres of various complexes has been associated with heightened catalytic  
11 activities of the complexes in some oxidation reactions.[39,40] However, only **3** showed  
12 catechol oxidase-like activities, forming 3,5-di-*tert*-butyl-1,2-benzoquinone (3,5-DTBQ)  
13 and 4-*tert*-butyl-1,2-benzoquinone (4-TBQ), respectively. Moreover, mechanistic studies  
14 with regard to vanadium-catalyzed catechol oxidation were performed to further  
15 elaborate on the nature of the active catalyst.

## 16 **Experimental**

17 All syntheses, experiments and manipulations were run in an ambient atmosphere.  
18  $[MoO_2(acac)_2]$  (*acac* = acetylacetonato) and  $[W(eg)_3]$  (*eg* = ethane-1,2-diolato = ethylene  
19 glycolato) were prepared according to known methods.[41,42] Other reagents and  
20 solvents were purchased from commercial sources and used as received. IR spectra were  
21 recorded using a Bruker VERTEX 71 FTIR spectrophotometer equipped with a RT-  
22 DLaDTGS detector. 64 scans were performed in the ATR mode for individual  
23 measurements using a Harrick VideoMVP accessory. In this setup, the sample is pressed  
24 against a diamond anvil. All data were recorded in transmittance mode with a resolution  
25 of  $4\text{ cm}^{-1}$ . The UV-Vis measurements were performed in MeOH, MeCN and  $CHCl_3$  using a  
26  $\varnothing$  1 cm quartz cuvette on an Agilent CARY60 spectrophotometer. High-resolution mass  
27 spectra were recorded on a Bruker Daltonics MicroTOF-Q II electrospray ionization time-  
28 of-flight (ESI-TOF) mass spectrometer using both negative and positive polarization as  
29 well as in MSMS mode. Full UV-Vis, NMR, IR and ESI-MS spectra are given in the  
30 supplementary material. The  $^1H$ ,  $^{13}C$ ,  $^{19}F$  and  $^{51}V$  spectra were recorded on either Bruker  
31 Avance III 600 MHz ( $^1H$ : 600.16 MHz,  $^{13}C$ : 150.91 MHz) equipped with a CryoProbe  
32 Prodigy triple resonance inverse (TCI) probe or Bruker Avance 500 MHz ( $^1H$ : 500.08 MHz  
33  $^{13}C$ : 125.75 MHz,  $^{19}F$ : 470.55 MHz,  $^{51}V$ : 131.54 MHz) equipped with a broad-band observe  
34 probe (Bruker BBO-5 mm-Zgrad). All  $^1H$  and  $^{13}C$  NMR spectra are reported in ppm  
35 downfield relative to tetramethylsilane (TMS,  $\delta$  = 0.00 ppm) and referenced to residual  
36 solvent signals  $CHCl_3$ -H ( $^1H$ :  $\delta$  7.26,  $^{13}C$ :  $\delta$  77.16), DMSO-*H*<sub>6</sub> ( $^1H$ :  $\delta$  2.50,  $^{13}C$ :  $\delta$  39.52),  
37 acetone-*H*<sub>6</sub> ( $^1H$ :  $\delta$  2.05,  $^{13}C$ :  $\delta$  206.26, 29.84) if TMS is not present.[43] The 0 ppm fluorine  
38 and vanadium reference frequencies were calculated from the TMS  $^1H$  frequency using  
39 the unified chemical shift scale by IUPAC ( $E(^{19}F, CCl_3F)$  = 94.094011) and ( $E(^{51}V, VOCl_3)$   
40 = 26.302948), respectively.[44]

## 1 Syntheses

2 ***N*-(1,3-dihydroxy-2-(hydroxymethyl)propan-2-yl)-3,5-di-*tert*-butylsalicylald-**  
 3 **imine ( $H_4L^{Saltris}$ )**. The proligand was synthesized according to a published procedure  
 4 with slight modifications.[28] Detailed synthetic procedure is disclosed in the ESI. Yield:  
 5 3.08 g (91 %).  $^1H$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  14.90 (1H, d,  $J$  = 2.2 Hz, ArOH),  
 6 8.55 (1H, d,  $J$  = 2.2 Hz, Ar—CH=N—CR<sub>3</sub>), 7.27 (1H, d,  $J$  = 2.5 Hz, ArH), 7.22 (1H, d,  $J$  = 2.5  
 7 Hz, ArH), 4.70 (3H, t,  $J$  = 5.4 Hz, NC—(CH<sub>2</sub>OH)<sub>3</sub>), 3.62 (6H, d,  $J$  = 5.4 Hz, NC—(CH<sub>2</sub>OH)<sub>3</sub>),  
 8 1.37 (9H, s, *t*-Bu), 1.27 (9H, s, *t*-Bu).  $^{13}C$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  165.37,  
 9 160.21, 138.12, 136.18, 126.57, 125.98, 117.49, 66.80, 61.41, 34.55, 33.78, 31.32, 29.29.  
 10 IR/cm<sup>-1</sup> ATR mode:  $\nu$  3500–3000vb (ROH, ArOH, R(CH<sub>2</sub>OH)<sub>3</sub>), 2958–2868s (C—H, *t*-Bu),  
 11 1620s (RCH=N), 1593–1361m (arom. C=C), 1252m (C—O, ArOH), 1049vs, 1038vs (C—  
 12 O, R(CH<sub>2</sub>OH)<sub>3</sub>), 882m, 644s.

13 **[MoO<sub>2</sub>(H<sub>2</sub>L<sup>Saltris</sup>)], 1.** 337 mg (1.0 mmol) H<sub>4</sub>L<sup>Saltris</sup> and 326 mg (1.0 mmol) [MoO<sub>2</sub>(acac)<sub>2</sub>]  
 14 were dissolved in 15 mL MeOH in a 50 mL Erlenmeyer flask equipped with a magnetic  
 15 stir-bar. The clear yellow reaction mixture was stirred at RT for 72 hours, after which it  
 16 was transferred to a -25 °C freezer. In a week, single crystals suitable for XRD had formed  
 17 and were subsequently isolated by Büchner filtration, washed with small amounts of ice-  
 18 cold MeOH, and air-dried to afford the analytically pure target complex. Yield: 340 mg (73  
 19 %).  $^1H$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  8.49 (1H, s, Ar—CH=N—CR<sub>3</sub>), 7.47 (1H,  
 20 d,  $J$  = 2.5 Hz, ArH), 7.44 (1H, d,  $J$  = 2.5 Hz, ArH), 4.95 (2H, t,  $J$  = 5.5 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>),  
 21 4.43 (2H, s, NR<sub>2</sub>C—(CH<sub>2</sub>O<sup>-</sup>)), 3.73 (2H, dd,  $J_1$  = 11.3 Hz,  $J_2$  = 5.5 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>), 3.63  
 22 (2H, dd,  $J_1$  = 11.3 Hz,  $J_2$  = 5.5 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>), 1.37 (9H, s, *t*-Bu), 1.29 (9H, s, *t*-Bu).  
 23  $^{13}C$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  164.72, 156.92, 140.49, 137.92, 129.20,  
 24 128.55, 120.60, 76.65, 73.69, 61.28, 48.61, 34.89, 33.97, 31.27, 29.62. IR/cm<sup>-1</sup> ATR mode:  
 25  $\nu$  3500–3000vb (ROH, R(CH<sub>2</sub>OH)<sub>2</sub>), 2956–2866s (C—H, *t*-Bu), 1620s (RCH=N), 1562–  
 26 1361m (arom. C=C), 1038vs (C—O, R(CH<sub>2</sub>OH)<sub>3</sub>), 1253m (C—O, ArOH), 1045s (C—O,  
 27 R(CH<sub>2</sub>OH)<sub>2</sub> and R(CH<sub>2</sub>O<sup>-</sup>)), 916vs  $\nu_s$ (MoO<sub>2</sub>)[45–48] 880vs  $\nu_{as}$ (MoO<sub>2</sub>)[45–48], 761m, 558s  
 28  $\nu$ (Mo—N)[49]. UV-Vis (MeCN;  $\lambda_{max}$ / nm ( $\epsilon$ / M<sup>-1</sup> cm<sup>-1</sup>) 355 (3180). ESI-HRMS<sup>+</sup> [M+Na]<sup>+</sup>  
 29 calculated  $m/z$  = 488.0945, found  $m/z$  = 488.0900. ESI-HRMS<sup>-</sup> [M-H]<sup>-</sup> calculated  $m/z$  =  
 30 464.0980, found  $m/z$  = 464.1056.

31 **1** could be also synthesized using equimolar amounts of H<sub>4</sub>L<sup>Saltris</sup> and Na<sub>2</sub>MoO<sub>4</sub> · 2 H<sub>2</sub>O.  
 32 However, excess glacial acetic acid was required, as well as overnight refluxing. Yield  
 33 using this methodology was also slightly reduced (277 mg, 60 %).

34 **[WO<sub>2</sub>(H<sub>2</sub>L<sup>Saltris</sup>)], 2.** 337 mg (1.0 mmol) H<sub>4</sub>L<sup>Saltris</sup> and 330 mg (1.0 mmol) Na<sub>2</sub>WO<sub>4</sub> · 2 H<sub>2</sub>O  
 35 were dissolved in 15 mL MeOH in a 100 mL Erlenmeyer flask equipped with a magnetic  
 36 stir-bar. The solution was treated with 1 mL glacial acetic acid, heated to boil in an oil-  
 37 bath for two hours, then left to stir for 72 hours at RT. The pale reaction mixture was  
 38 concentrated to *ca.* half of the original volume, filtered and transferred to +5 °C  
 39 refrigerator. Upon *ca.* two weeks of storage, pale yellow microcrystals suitable for single  
 40 crystal XRD had deposited. They were subsequently isolated by Büchner filtration,  
 41 washed with small amounts of ice-cold MeOH and air-dried to afford the analytically pure

1 target compound. Yield: 258 mg (47 %).  $^1\text{H}$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  8.46  
 2 (1H, s, Ar—CH=N—CR<sub>3</sub>), 7.53 (1H, d, *J* = 2.4 Hz, ArH), 7.47 (1H, d, *J* = 2.4 Hz, ArH), 4.99  
 3 (2H, t, *J* = 5.4 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>), 4.58 (2H, s, NR<sub>2</sub>C—(CH<sub>2</sub>O<sup>-</sup>)), 3.72 (2H, dd, *J*<sub>1</sub> = 11.1  
 4 Hz, *J*<sub>2</sub> = 3.4 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>), 3.63 (2H, dd, *J*<sub>1</sub> = 11.1 Hz, *J*<sub>2</sub> = 3.4 Hz, NRC—(CH<sub>2</sub>OH)<sub>2</sub>),  
 5 1.38 (9H, s, *t*-Bu), 1.30 (9H, s, *t*-Bu).  $^{13}\text{C}$  NMR (DMSO-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  165.51,  
 6 155.81, 141.03, 138.36, 129.37, 129.06, 121.22, 76.76, 73.72, 61.17, 48.61, 34.85, 33.97,  
 7 31.26, 29.60. IR/cm<sup>-1</sup> ATR mode:  $\nu$  3500–3000<sub>vb</sub> (ROH, R(CH<sub>2</sub>OH)<sub>2</sub>), 2958–2850<sub>s</sub> (C–H,  
 8 *t*-Bu), 1621<sub>s</sub> (RCH=N), 1566–1362<sub>m</sub> (arom. C=C), 1300<sub>m</sub> (C—O, ArOH), 1048<sub>vs</sub> (C—O,  
 9 R(CH<sub>2</sub>OH)<sub>2</sub> and R(CH<sub>2</sub>O<sup>-</sup>)), 893<sub>vs</sub>  $\nu_s$ (WO<sub>2</sub>)[45–48] 860<sub>vs</sub>  $\nu_{as}$ (WO<sub>2</sub>)[45–48], 764<sub>m</sub>, 561<sub>s</sub>  
 10  $\nu$ (W—N)[49]. UV-Vis (MeOH;  $\lambda_{\text{max}}$ / nm ( $\epsilon$ / M<sup>-1</sup> cm<sup>-1</sup>) 365 (2440). ESI-HRMS<sup>+</sup> [M+Na]<sup>+</sup>  
 11 calculated *m/z* = 574.1399, found *m/z* = 574.1435. ESI-HRMS<sup>-</sup> [M–H]<sup>-</sup> calculated *m/z* =  
 12 550.1424, found *m/z* = 550.2204.

13 **2** could be also synthesized using equimolar amounts of H<sub>4</sub>L<sup>Saltris</sup> and [W(eg)<sub>3</sub>]. The  
 14 isolated yield of a 1 mmol scale synthesis was very low, only 40 mg (9 %).

15 **[VO(HL<sup>Saltris</sup>)<sub>2</sub>, 3**. 337 mg (1.0 mmol) H<sub>4</sub>L<sup>Saltris</sup> and 265 mg (1.0 mmol) [VO(acac)<sub>2</sub>] were  
 16 dissolved in 15 mL MeOH in a 100 mL Erlenmeyer flask equipped with a magnetic stir-  
 17 bar. Upon *ca.* 15 minutes of stirring at RT, an umber-colored solid precipitated out of  
 18 solution. The solid was collected by Büchner filtration, washed with *ca.* 30 mL RT MeOH,  
 19 and air-dried to afford the target compound in a pure non-crystalline form. UMBER-  
 20 colored single crystals suitable for x-ray diffraction were obtained from hot acetonitrile  
 21 solution upon slow cooling. Yield: 357 mg (89 %).  $^1\text{H}$  NMR (acetone-*d*<sub>6</sub>, 298 K, 600 MHz,  
 22 TMS)  $\delta$  9.08 (1H, s, Ar—CH=N—CR<sub>3</sub>), 7.66 (1H, d, *J* = 2.5 Hz, ArH), 7.53 (1H, d, *J* = 2.5 Hz,  
 23 ArH), 5.25 (1H, d, *J* = 14.3 Hz, NR<sub>2</sub>C—(CH<sub>A</sub>H<sub>B</sub>O<sup>-</sup>)<sub>A</sub>), 5.13 (2H, m, NR<sub>2</sub>C—(CH<sub>A</sub>H<sub>B</sub>O<sup>-</sup>)<sub>A</sub> and  
 24 NR<sub>2</sub>C—(CH<sub>A</sub>H<sub>B</sub>O<sup>-</sup>)<sub>B</sub>), 4.70 (1H, d, *J* = 8.9 Hz, NR<sub>2</sub>C—(CH<sub>A</sub>H<sub>B</sub>O<sup>-</sup>)<sub>B</sub>), 4.58 (1H, t, *J* = 5.4 Hz,  
 25 NR<sub>2</sub>C—(CH<sub>2</sub>OH)<sub>C</sub>), 4.04 (2H, dd, *J*<sub>1</sub> = 5.6 Hz, *J*<sub>2</sub> = 3.2 Hz, NR<sub>2</sub>C—(CH<sub>2</sub>OH)<sub>C</sub>), 1.51 (9H, s, *t*-  
 26 Bu), 1.35 (9H, s, *t*-Bu).  $^{13}\text{C}$  NMR (acetone-*d*<sub>6</sub>, 298 K, 600 MHz, TMS)  $\delta$  165.46, 160.98,  
 27 141.26, 137.84, 131.29, 129.74, 121.38, 85.88, 81.87, 79.65, 64.51, 35.99, 34.84, 31.70,  
 28 30.45.  $^{51}\text{V}$  NMR (acetone-*d*<sub>6</sub>, 298 K, 600 MHz, VOCl<sub>3</sub>)  $\delta$  -562. IR/cm<sup>-1</sup> ATR mode:  $\nu$  3571<sub>w</sub>  
 29 (ROH), 2954–2864<sub>s</sub> (C–H, *t*-Bu), 1623<sub>s</sub> (RCH=N), 1556–1360<sub>m</sub> (arom. C=C), 1303<sub>m</sub> (C—  
 30 O, ArOH), 1075<sub>s</sub>, 1030<sub>s</sub> (C—O, R(CH<sub>2</sub>OH)<sub>2</sub> and R(CH<sub>2</sub>O<sup>-</sup>)), 893<sub>vs</sub>  $\nu$ (V=O)[19,22,50].  
 31 853<sub>m</sub>, 764<sub>m</sub>, 552<sub>s</sub>. UV-Vis (MeCN;  $\lambda_{\text{max}}$ / nm ( $\epsilon$ / M<sup>-1</sup> cm<sup>-1</sup>) 520 (550). ESI-HRMS<sup>+</sup> [M+Na]<sup>+</sup>  
 32 calculated *m/z* = 825.2718, found *m/z* = 825.2599. ESI-HRMS<sup>-</sup> [M + Cl]<sup>-</sup> calculated *m/z* =  
 33 837.2508, found *m/z* = 837.2273.

34 **3** could be also synthesized using equimolar amounts of H<sub>4</sub>L<sup>Saltris</sup> and VOSO<sub>4</sub> · 5 H<sub>2</sub>O, with  
 35 two equivalents of Et<sub>3</sub>N added as a base. Similarly to the synthesis involving [VO(acac)<sub>2</sub>],  
 36 an umber solid was quickly precipitated out of solution, however, with a slightly lowered  
 37 yield; a 1.0 mmol scale synthesis yielded 277 mg of the target compound, corresponding  
 38 to a yield of 69 %.

39

## 1 X-ray crystallography

2 Single crystal X-ray data were collected using Rigaku Oxford diffraction SuperNova  
3 diffractometer equipped with micro-focus single-source (Mo-K $\alpha$  radiation,  $\lambda = 0.71073$   
4 Å) and Eos detector. Data collection and processing were done using CrysAlis<sup>Pro</sup>[51]  
5 software. Crystal structures were solved and refined within Olex<sup>2</sup>[52] GUI using  
6 SHELXS[52] and SHELXL[53] programs, respectively. Atoms heavier than H were located  
7 from the difference density map and refined anisotropically. All C—H atoms were  
8 calculated to their ideal positions and refined using a riding model with the isotropic  
9 displacement parameters fixed to values corresponding to 1.2–1.5 times the  $U_{\text{aniso}}$  of the  
10 respective host atom. O—H hydrogen atoms that participate in hydrogen bond  
11 interactions were located from the difference density map and refined isotropically  
12 without restrictions in displacement parameters. The single crystal X-ray data for **1–3** are  
13 given in ESI table S1. Crystallographic data for the compounds reported in this paper  
14 were deposited with the Cambridge Crystallographic Data Centre, CCDC, 12 Union Road,  
15 Cambridge CB21EZ, UK. These data can be obtained free of charge on quoting the  
16 depository number CCDC 1959437–1959439 (E-Mail: [deposit@ccdc.cam.ac.uk](mailto:deposit@ccdc.cam.ac.uk),  
17 <http://www.ccdc.cam.ac.uk>). The powder x-ray diffraction measurements were  
18 performed using a Huber G670 detector (Cu-K $\alpha$  radiation,  $\lambda = 1.5406$  Å). For each  
19 individual measurement, the exposure time was set to 30 min and with a total of 10 scans.

## 20 Cyclic voltammetry

21 The cyclic voltammetry electrochemical measurements of **1–3** ( $c = 1 \times 10^{-3}$  M) were  
22 performed using a standard three-electrode setup using an Autolab PGSTAT101  
23 potentiostat. The CVs were recorded at RT using  $\varnothing$  1 mm platinum and glassy carbon (GC)  
24 working electrodes. Before use both working electrodes were polished using 6, 3, 1 and  
25 0.25  $\mu\text{m}$  diamond paste, and rinsed with quartz distilled water and technical ethanol. The  
26 quasi-reference electrode was an Ag/AgCl wire, and it was calibrated against the  
27 ferrocene redox-couple (Fc/Fc<sup>+</sup>) ( $E_{1/2}(\text{Fc}/\text{Fc}^+) = 0.55$  V).[54] A coiled platinum wire was  
28 employed as the counter electrode. Measurements were performed using 50, 100 and  
29 200  $\text{mV s}^{-1}$  scanning rates in dry electrochemical grade acetonitrile with 0.1 M tetrabutyl-  
30 ammonium tetrafluoroborate (TBABF<sub>4</sub>) as the supporting electrolyte. Prior to every  
31 measurement, the sample solutions were bubbled with dry N<sub>2</sub> for 10 minutes. The  
32 electrochemical window was  $-2.5 - 2.5$  V for GC working electrode, and  $-0.8 - 2.3$  V for  
33 Pt electrode, respectively.

## 34 Oxygen atom transfer reactions

35 All complexes were preliminarily assessed in the formal oxygen atom transfer (OAT)  
36 reactions of tris(4-fluorophenyl)phosphine (P(*p*-C<sub>6</sub>H<sub>4</sub>F)<sub>3</sub>) to the corresponding tris(4-  
37 fluorophenyl)phosphine oxide (P(*p*-C<sub>6</sub>H<sub>4</sub>F)<sub>3</sub>O).[55] In the OAT experiments, 500  $\mu\text{L}$  of  $5$   
38  $\times 10^{-3}$  M P(*p*-C<sub>6</sub>H<sub>4</sub>F)<sub>3</sub> in DMSO-H<sub>6</sub> solution and 500  $\mu\text{L}$  of  $5 \times 10^{-4}$  M **1–3** in DMSO-H<sub>6</sub>  
39 solution were combined in a screw-capped scintillation vial and maintained for 24 hours  
40 at 60 °C in a thermal oven. Additionally, a control reaction involving 1000  $\mu\text{L}$   $5 \times 10^{-3}$  M

1 P(*p*-C<sub>6</sub>H<sub>4</sub>F)<sub>3</sub> in DMSO-H<sub>6</sub> solution was also run. In all cases, the conversion was assessed  
 2 by <sup>19</sup>F NMR and after the 24-hour period. All measurements were run in duplicate and  
 3 the results obtained are given as the average of the two reactions.

#### 4 **Oxidation of 4-TBC and 3,5-DTBC**

5 The oxidations of 4-TBC and 3,5-DTBC to 4-TBQ and 3,5-DTBQ were monitored by *in-situ*  
 6 UV-Vis spectroscopy on an Agilent Cary 60 UV-Vis spectrophotometer and a Ø 1.00 cm  
 7 quartz cuvette. All measurements were run in duplicate and conducted at room  
 8 temperature (25 °C) under ambient atmosphere. The reaction kinetics were assessed  
 9 using the method of initial rates. For this, a 1.00 × 10<sup>-5</sup> M solution of precatalyst **3** was  
 10 prepared in CHCl<sub>3</sub>. The substrate concentration was varied 1000, 2000, 3000, 4000, 6000,  
 11 8000 and 10000-fold relative to the catalyst to evaluate the substrate dependence of the  
 12 reactions for 4-TBC. In the case of 3,5-DTBC, substrate concentration ratios of 12500,  
 13 15000 and 17500 were additionally used. The reactions were initiated directly in the  
 14 quartz cuvette by combining 1.500 mL precatalyst solutions and 1.500 mL of 1.75 × 10<sup>-1</sup>  
 15 M – 1.00 × 10<sup>-2</sup> M substrate solutions. Accordingly, final concentrations of [cat] = 5.00 ×  
 16 10<sup>-6</sup> M and [S] = 8.75 × 10<sup>-2</sup> M – 5.00 × 10<sup>-3</sup> M, respectively, were obtained during  
 17 catalysis. The progress of the reaction was monitored by following the increase in the  
 18 absorbance at *ca.* 388 nm due to 4-TBQ and 3,5-DTBC (ε(4-TBQ) = 1150 M<sup>-1</sup> cm<sup>-1</sup> and  
 19 ε(3,5-DTBQ) = 2200 M<sup>-1</sup> cm<sup>-1</sup>)[56,57] for 10 minutes at every one-minute interval.  
 20 Absorbance vs. time graphs were plotted, and initial reaction rates at every concentration  
 21 were determined directly utilizing the on-board Cary WinUV Kinetics software. Finally,  
 22 the initial reaction rates and the substrate concentrations were fitted, using Origin 2015  
 23 software, to the Michaelis–Menten equation (1) using non-linear regression analysis, in  
 24 order to obtain the maximum reaction rate *V*<sub>max</sub>, Michaelis constant *K*<sub>M</sub> ([S] at ½*V*<sub>max</sub>) and  
 25 turnover frequency *k*<sub>cat</sub> values. The turnover frequency was calculated using the formula  
 26 (2), where [*E*<sub>T</sub>] is the catalyst concentration.

$$27 \quad v = \frac{V_{\max} \times [S]}{K_M + [S]} \quad (1)$$

$$28 \quad k_{\text{cat}} = \frac{V_{\max}}{[E_T]} \quad (2)$$

#### 29 **Iodometric assay**

30 Hydrogen peroxide was determined from the catechol oxidation reaction mixture based  
 31 on I<sub>3</sub><sup>-</sup>, which has a characteristic UV-Vis absorbance at *ca.* 353 nm in water. A 100 mL  
 32 Erlenmeyer flask was charged with *ca.* 1 g 4-*tert*-butylcatechol and dissolved in 30 mL  
 33 CHCl<sub>3</sub>. 1 mol-%, or *ca.* 48 mg **3** was added to the reaction mixture, which was  
 34 subsequently stirred for several days at RT. Afterwards, the reaction mixture was  
 35 extracted with 25 mL distilled water. The aqueous phase was adjusted to a pH ~ 2 with  
 36 dilute aqueous H<sub>2</sub>SO<sub>4</sub>, and a portion of it was treated with *ca.* 0.32 M KI (aq) solution and  
 37 was allowed to react overnight. I<sub>3</sub><sup>-</sup> was observed by UV-Vis spectroscopy at *ca.* 353 nm,  
 38 whereas no such signal was observed for the bare KI solution, which was allowed to react



1 with atmospheric oxygen overnight. Additionally, to further confirm the presence of  
2 hydrogen peroxide, a portion of the KI solution was treated with excess amounts of 30 %  
3 aqueous H<sub>2</sub>O<sub>2</sub>, and the formation of a 353 nm band was again observed (ESI Figure S57)

#### 4 **Mechanistic catechol oxidation investigations**

5 The UV-Vis spectroscopic investigations were performed in 1 cm quartz cuvettes by  
6 combining 1.500 mL 0.100 M 4-TBC chloroform solution containing 0.010 M BHT, DMSO  
7 or H<sub>2</sub>O<sub>2</sub>, and 1.500 mL chloroform solutions consisting of  $1 \times 10^{-4}$  M **3**. For H<sub>2</sub>O<sub>2</sub> tests,  
8 0.020 and 0.030 M solutions were additionally used. For control reactions, 0.100 M 4-TBC  
9 reacts with 0.030 M H<sub>2</sub>O<sub>2</sub> in the absence of **3** (control 1), and 0.100 M 4-TBC reacts with  
10  $1 \times 10^{-4}$  M **3** in the absence of H<sub>2</sub>O<sub>2</sub> (control 2). The initial reaction rates were determined  
11 using the on-board Cary WinUV Kinetics software as described above. All measurements  
12 were repeated once, and the results given in figure 7 represent average values obtained  
13 from the two measurements.

14 For <sup>51</sup>V NMR and ESI-HRMS studies, 6 mg of **3** was suspended in 650 μL CDCl<sub>3</sub> and treated  
15 with 100 equivalents of 3,5-DTBC (166 mg) or 4-TBC (124 mg) to reflect the  
16 stoichiometry of the kinetic catechol oxidation experiments. Prior to measurements the  
17 solutions were filtered through a small cotton plug to remove any undissolved solids. <sup>51</sup>V  
18 NMR measurements were performed immediately after filtration. The same CDCl<sub>3</sub>  
19 solutions were diluted in MS grade acetonitrile, and subsequently analyzed by ESI-HRMS  
20 immediately after the <sup>51</sup>V NMR measurements.

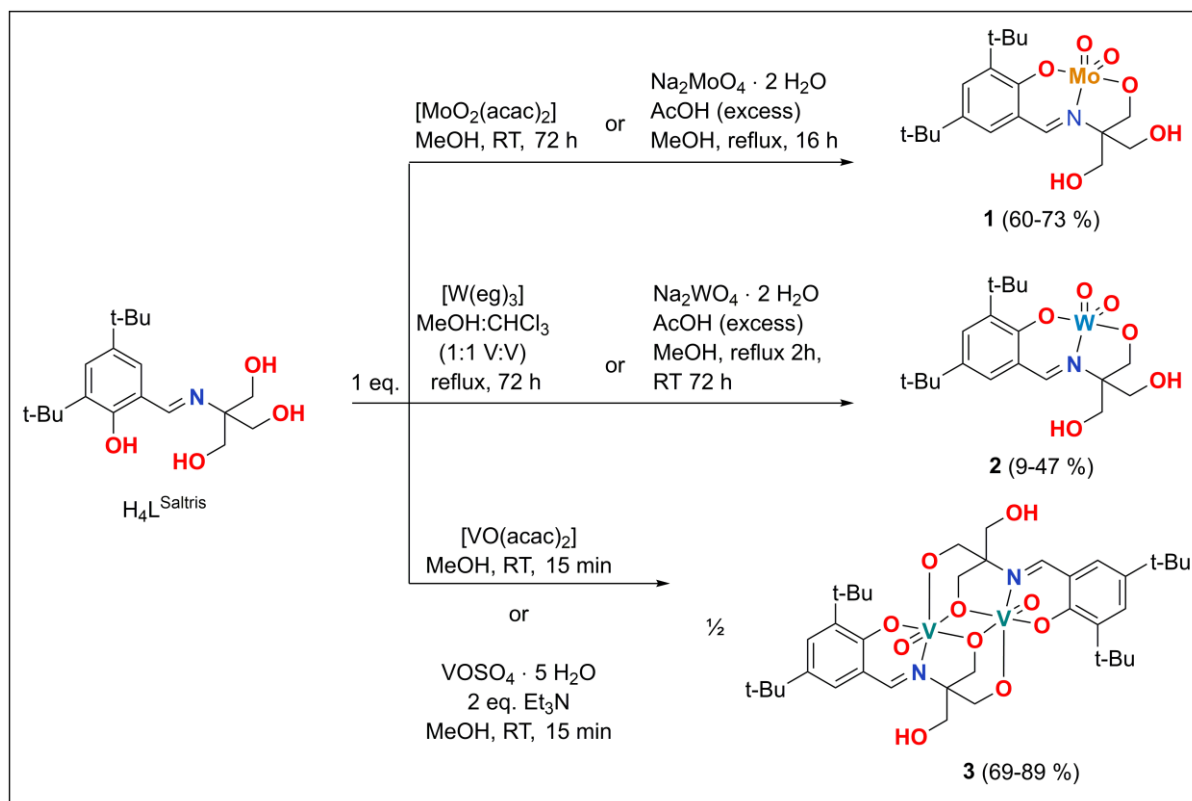
21 To determine the product distribution in the oxidation of 3,5-DTBC, 500 mg of 3,5-DTBC  
22 and 18 mg of **3** (1 mol-%) were allowed to react in 30 mL boiling chloroform under  
23 ambient atmosphere overnight for *ca.* 16 hours in a 50 mL round-bottomed flask  
24 equipped with a magnetic stir-bar and a reflux condenser. Afterwards, the reaction  
25 mixture was analyzed by TLC on silica plates with technical DCM. Three components with  
26 R<sub>f</sub>-values *ca.* 0.85, 0.58 and 0.33 were detected. The compounds were isolated by column  
27 chromatography on silica gel (technical DCM), and identified with <sup>1</sup>H, <sup>13</sup>C NMR and  
28 positive mode ESI-HRMS according to published data (ESI).[35]

## 29 **Results and discussion**

### 30 **Syntheses and characterization of complexes**

31 The ligand precursor H<sub>4</sub>L<sup>Saltris</sup> was synthesized according to a slightly modified  
32 procedure.[28] We were also interested in the corresponding H<sub>5</sub>L<sup>Saltris</sup> amine analogue,  
33 but our attempts to obtain it through reductive amination from H<sub>4</sub>L<sup>Saltris</sup> proved  
34 unsuccessful. However, Mannich reaction did provide the target compound, although  
35 unselectively, and with trace yields only. Accordingly, complexations were not tested  
36 with H<sub>5</sub>L<sup>Saltris</sup>. The Mannich reaction including the product distribution is presented the  
37 electronic supplementary information (Scheme S1). The spectral data of all products are  
38 also listed in the ESI.

1  $H_4L^{\text{Saltris}}$  reacts with equimolar amounts of molybdenum precursors  $[MoO_2(acac)_2]$  or  
 2  $Na_2MoO_4 \cdot 2 H_2O$  in methanol to afford *cis*- $[MoO_2(H_2L^{\text{Saltris}})]$  (**1**) as a yellow crystalline  
 3 solid, whereas the use of  $[MoO_2(acac)_2]$  provides slightly better yields. Likewise, the  
 4 tungsten precursor  $[W(eg)_3][42]$  gave *cis*- $[WO_2(H_2L^{\text{Saltris}})]$  (**2**) in methanol as pale  
 5 crystals, although the (isolated) yield was very low. However,  $Na_2WO_4 \cdot 2 H_2O$  can be used  
 6 to obtain **2** in a moderate yield. Vanadium precursor  $[VO(acac)_2]$  rapidly reacts with  
 7  $H_4L^{\text{Saltris}}$  in methanol at RT to form an umber-colored precipitate in a high yield. The solid  
 8 can be crystallized from hot acetonitrile to yield analytically pure **3** with a formula  
 9  $[VO(HL^{\text{Saltris}})]_2$ . The schematic representation of the complex syntheses is given in scheme  
 10 1.



12 All complexes were fully characterized by means of high-resolution electrospray  
 13 ionization time-of-flight mass spectrometry in positive and negative ionization modes,  
 14 also utilizing MSMS. Characteristic to all complexes in the positive ionization mode is the  
 15 presence of multiple singly charged sodium adducts of the type  $[M - nH + (n+1)Na]^+$ ,  
 16 where  $n = 1, 2, 3$ . In these species the protons in the trisbase alcohol arms of the  
 17 complexes are sequentially substituted by sodium ions. However, singly protonated  
 18 adducts of the type  $[M + H]^+$  are generally not observed. Other common adducts in the  
 19 positive ionization are singly charged oligomerized adducts of general type  $[nM+Na]^+$ ,  
 20 where  $n = 1, 2, 3$ . These are most likely formed by electrostatic interactions, and not  
 21 representative of genuine species observable outside ionizing conditions. In the negative  
 22 ionization mode, the most characteristic ionization adduct is the  $[M-H]^-$  from the loss of  
 23 a proton. Other common adducts are chlorido adducts of the type  $[M+Cl]^-$ , and  
 24 dealkylation adducts  $[M-R]^-$ , where R represent a generic alkyl or alcohol group.

1 Similarly to what is observed in the positive ionization mode, electrostatic  
2 oligomerization is evident in the negative mode also.

3 The UV-Vis spectra of  $H_4L^{Saltris}$ , **1**, **2** and to a certain extent **3**, are rather similar, and  
4 contain three clear electronic absorptions. All UV-Vis spectra are presented in ESI figure  
5 14, and the spectral data has been tabulated in Table 1. By comparing the UV-Vis  
6 spectrum of  $H_4L^{Saltris}$  to those of **1–3** it may be concluded that all signals below *ca.* 400 nm  
7 in the case of **1** and **2**, or below *ca.* 460 nm in the case of **3** are primarily of intraligand  
8 origin. While all electronic absorptions in the proligand are visible in the UV-Vis spectra  
9 of the complexes, a noticeable redshift is observed in all signals, consistent with  
10 coordination. All high-intensity ( $\epsilon \geq 2000\text{--}20000\text{ M}^{-1}\text{ cm}^{-1}$ ) electronic absorptions below  
11 *ca.* 240 nm, and the second, lower energy ( $\epsilon \sim 1150\text{--}17700\text{ M}^{-1}\text{ cm}^{-1}$ ) absorptions at *ca.*  
12 260–287 nm in all compounds are attributed to  $\pi \rightarrow \pi^*$  transitions in the aromatic ring  
13 and due to extended conjugation in the salicylaldimine chromophore. The third band at  
14 *ca.* 328–363 for  $H_4L^{Saltris}$ , **1** and **2** is assigned to the  $n \rightarrow \pi^*$  transitions ( $\epsilon \sim 40\text{--}3200\text{ M}^{-1}$   
15  $\text{cm}^{-1}$ ) in the aldimine functionality.[58] **3** also contains this transition, however, it is  
16 eclipsed by what are clearly multiple overlapping absorptions in the general area  
17 between *ca.* 320–460 nm. These multiple signals have been assigned as LMCT transitions,  
18 most likely attributable to alkoxido to vanadium(V) transitions.[59] A very low-intensity  
19 LMCT at *ca.* 520 nm is assigned to phenolato  $\rightarrow$  V(V) ( $p_\pi \rightarrow d$ ), with an  $\epsilon \sim 550\text{ M}^{-1}\text{ cm}^{-1}$ .  
20 [59] The  $p_\pi \rightarrow d$  transitions in **1** and **2**, although they must exist, are too weak to be  
21 appropriately assigned. Likewise, the alkoxido to metal LMCT bands are much weaker in  
22 **1** and **2** compared to **3**.

23 In the IR spectra of all compounds, a broad signal at 3100–3500 corresponds to the  
24 phenolic and/or alcoholic OH groups while a strong stretching vibration of the imine  
25 group occurs at around  $1622\text{ cm}^{-1}$ . The complexation of Mo, W and V to  $H_4L^{Saltris}$  can be  
26 substantiated from corresponding metal oxido stretching vibrations.[29] Specifically, the  
27 characteristic symmetric and asymmetric  $\nu(MO_2)$  ( $M = Mo, W$ ) stretches are found at 930  
28 and  $889\text{ cm}^{-1}$  for **1**, and at 946 and  $893\text{ cm}^{-1}$  for **2**, respectively.[47] These are in line  
29 with what has been reported for similar dioxidomolybdenum complexes.[60] For **3**,  
30 featuring only a single oxido ligand, a characteristic  $\nu(V=O)$  stretching vibration is found  
31 at  $952\text{ cm}^{-1}$ , which is consistent with experimental values obtained for other  
32 oxidovanadium(V) complexes bearing similar ligands.[49,59] Furthermore, a low energy  
33 signal at  $553\text{ cm}^{-1}$  can be seen, which is assigned as  $\nu(V-N)$  stretch.[61] This signal is  
34 also found in **1** at  $557\text{ cm}^{-1}$  and is tentatively assigned to  $\nu(Mo-N)$ , since similar  
35 stretching vibrations are known in dioxidomolybdenum(VI) complexes bearing variable  
36 bidentate cysteamine-type ligands.[62] The signal seen in the IR spectrum of **2** at  $562\text{ cm}^{-1}$   
37 is cautiously assigned to  $\nu(W-N)$  stretches by virtue of similarity to **1**. It should be  
38 emphasized, that the strong signal in question is missing in the IR spectrum of the  
39 proligand. IR spectral data for the most characteristic signals in all compounds in  
40 presented in table 1.

41

1

**Table 1.** IR and UV-Vis data for H<sub>4</sub>L<sup>Saltris</sup> and **1–3**. See text for further information

Compound	IR (cm <sup>-1</sup> )				
	$\nu(\text{O—H})$	$\nu(\text{C=N})$	$\nu(\text{M=O})$	$\nu_{\text{asymm.}}(\text{M=O})$	$\nu(\text{M—N})$
H <sub>4</sub> L <sup>Saltris</sup>	3100–3500	1622	–	–	–
<b>1</b>	3100–3500	1620	930	889	557
<b>2</b>	3100–3500	1621	946	893	562
<b>3</b>	3570	1623	952	–	553
Compound	UV-Vis				
	$\lambda_{\text{max}}$ (nm)	$\epsilon$ (M <sup>-1</sup> cm <sup>-1</sup> )	transition		
H <sub>4</sub> L <sup>Saltris</sup>	225	$\geq 2000$	$\pi \rightarrow \pi^*$		
	262	1150	$\pi \rightarrow \pi^*$		
	328	400	$n \rightarrow \pi^*$		
<b>1</b>	230	$\geq 20000$	$\pi \rightarrow \pi^*$		
	260	16700	$\pi \rightarrow \pi^*$		
	355	3200	$n \rightarrow \pi^*$		
<b>2</b>	230	$\geq 20000$	$\pi \rightarrow \pi^*$		
	274	12500	$\pi \rightarrow \pi^*$		
	363	2500	$n \rightarrow \pi^*$		
<b>3</b>	240	$\geq 20000$	$\pi \rightarrow \pi^*$		
	287	17700	$\pi \rightarrow \pi^*$		
	320–460	$\sim 7600\text{--}700$	$n \rightarrow \pi^*$		
	520	550	$p\pi \rightarrow d$		

2

3 For H<sub>4</sub>L<sup>Saltris</sup> the reported <sup>1</sup>H NMR data[28] significantly differs from that obtained by us,  
4 possibly due to the some difficulties in the measurements in the previous report. In any  
5 case, the <sup>1</sup>H and <sup>13</sup>C NMR data obtained by us are in good agreement with the expected  
6 structure of the ligand precursor and as such, we find re-publication of good NMR data  
7 warranted. Striking features in the <sup>1</sup>H NMR spectrum of H<sub>4</sub>L<sup>Saltris</sup>, very useful for structure  
8 determination, are the proton signals corresponding to the phenol, aldimine carbon, and  
9 tris-alcohol sidearms. The <sup>1</sup>H NMR data for H<sub>4</sub>L<sup>Saltris</sup> and **1–3** is given in table 2.

10 The signal for the phenolic proton is found at 14.90 ppm, which is significantly downfield  
11 for what would generally be expected for phenols or salicylaldehydes, at around 10 ppm  
12 or below. Schiff bases derived from salicylaldehydes are known to exist as tautomers,  
13 shifting between their enolimine (OH) and ketoenamine (NH) forms, through what is  
14 known as RAHB (resonance assisted intramolecular hydrogen bonding).[63,64] The  
15 significant downfield resonance for the phenol proton in H<sub>4</sub>L<sup>Saltris</sup> can be attributed to  
16 moderate de-shielding effect caused by the intramolecular hydrogen bonding to the  
17 aldimine nitrogen (ArO—H---N=R<sup>1</sup>R<sup>2</sup>).[64,65] Furthermore, with similar molecules the  
18 <sup>13</sup>C chemical shift of the *ipso* carbon atom connected to phenol OH is known to be sensitive  
19 to the relative tautomeric population between OH and NH forms, with lower average  
20 values of *ca.* 155 ppm, and higher average values of *ca.* 180 ppm generally being expected  
21 for OH, and NH forms, respectively.[64,65] In H<sub>4</sub>L<sup>Saltris</sup> the corresponding <sup>13</sup>C NMR signal  
22 is found at 160.21 ppm in DMSO-*d*<sub>6</sub>, indicative of tautomeric equilibrium favoring the OH

1 form, a feature that is also common for similar compounds.[64] A small J-coupling (2.2  
2 Hz) between the phenol-apparent and aldimine carbon proton can be seen in the  $^1\text{H}$  NMR  
3 spectrum in DMSO-*d*<sub>6</sub>. In light of tautomeric equilibrium, this coupling has been shown  
4 to arise from  $^3\text{J}_{\text{NH},\alpha\text{H}}$  vicinal coupling between NH and CH of the ketoenamine form.[64]  
5 Small J-couplings of this type have been encountered before for similar compounds.[64]  
6 Other information, particularly useful for determining coordination mode of  $\text{H}_4\text{L}^{\text{Saltris}}$  to  
7 various metals, are the proton signals of the tris-alcohol sidearms. In  $\text{H}_4\text{L}^{\text{Saltris}}$  the three  
8 alcohol protons are clearly visible at  $\delta$  4.70, as a triplet, with a  $^3\text{J}_{\text{HH}} = 5.4$  Hz coupled to the  
9 six vicinal methylene protons, appearing as a duplet, at  $\delta$  3.62 ( $^3\text{J}_{\text{HH}} = 5.4$  Hz) when  
10 relatively dry DMSO-*d*<sub>6</sub> is used.

11 The coordination mode of  $\text{H}_4\text{L}^{\text{Saltris}}$  to Mo, W and V was unequivocally determined by NMR  
12 in addition to single-crystal XRD. Quite unsurprisingly, the corresponding *cis*-  
13 dioxidomolybdenum(VI) and -tungsten(VI) complexes *cis*- $[\text{MO}_2(\text{H}_2\text{L}^{\text{Saltris}})]$ , where M = Mo  
14 or W, share analogous structures in DMSO solution as well.  $\text{H}_4\text{L}^{\text{Saltris}}$ , as a potentially  
15 pentadentate ligand, coordinates to both metal centers in tridentate, dianionic manner  
16 *via* the phenolato oxygen, aldimino nitrogen and one alcoholato sidearm.[66,67] The  
17 coordination mode is clearly evident from  $^1\text{H}$  NMR in dry DMSO-*d*<sub>6</sub>. For example, the  
18 phenol proton at  $\delta$  14.90 is missing, and the aromatic protons have been shifted  
19 downfield, relative to  $\text{H}_4\text{L}^{\text{Saltris}}$ , for *ca.* 0.21 ppm and 0.26 ppm for **1** and **2**, respectively.  
20 Curiously, the signal corresponding to aldimine CH proton is shifted upfield very slightly  
21 ( $< 0.10$  ppm) in both complexes relative to  $\text{H}_4\text{L}^{\text{Saltris}}$ . These effects might be explained by  
22 inductive effects: Upon coordination of the aldimine nitrogen, its electron density is  
23 withdrawn towards the electron-deficient Mo(VI) and W(VI) centers, manifesting as a  
24 slight shielding effect for the aldimine CH proton.

25 As was disclosed earlier, the three alcohol protons in  $\text{H}_4\text{L}^{\text{Saltris}}$  are clearly visible as a  
26 triplet, however, in the complexes these same triplets only correspond to two protons. In  
27 **1**, the triplet corresponding to two free sidearm alcohol protons is found slightly  
28 downfield relative to  $\text{H}_4\text{L}^{\text{Saltris}}$  at  $\delta$  4.95 ppm, ( $^3\text{J}_{\text{HH}} = 5.5$  Hz), whereas in **2** it is located at  $\delta$   
29 4.99 ppm. In both complexes, the four methylene protons corresponding to the two  
30 uncoordinated alcohol sidearms appear as duplets of duplets, at  $\delta$  3.73 and 3.64 for **1**, and  
31  $\delta$  3.72 and 3.63 ppm for **2**, respectively. Accordingly, the methylene CH<sub>2</sub> for the  
32 coordinated alcoholato sidearm is found, as a singlet, at  $\delta$  4.43 for **1** and  $\delta$  3.58 for **2**.

33 The most distinguishable feature of the  $^{13}\text{C}$  NMR spectra for **1** and **2** relative to that of  
34  $\text{H}_4\text{L}^{\text{Saltris}}$  are the methylene CH<sub>2</sub> carbons in the alcohol sidearms. In **1** and **2** these become  
35 inequivalent due to expected slightly different electronic environments and steric  
36 perturbations caused by coordination to the metals. Except for the quaternary carbon  
37 connecting to the alcoholato arms and aldimino nitrogen, which is upshifted in both  
38 complexes for *ca.* 18 ppm, the alcohol and alcoholato methylene protons have been  
39 downshifted for *ca.* 15 ppm in both **1** and **2**, consistent with coordination and changes in  
40 electronic environment.

**Table 2.**  $^1\text{H}$  NMR spectral data of  $\text{H}_4\text{L}^{\text{Saltris}}$ , **1–3** ( $\delta$  in ppm). Multiplicity: s = singlet, d = duplet, t = triplet, m = multiplet (unresolved), b = broad.

Compound	ArOH	CH=N	Arom. H	NC(CH <sub>2</sub> OH) <sub>3</sub>	NC(CH <sub>2</sub> OH) <sub>3</sub>	tBu
$\text{H}_4\text{L}^{\text{Saltris}}$	14.90 (d, 1H)	8.55 (d, 1H)	7.27, 7.22 (d, 1H)	4.70 (t, 3H)	3.61 (d, 6H)	1.37, 1.27 (s, 9H)
<b>1</b>	-	8.49 (s, 1H)	7.47, 7.44 (d, 1H)	4.95 (t, 2H)	3.73; 3.64 (dd, 2H); (dd, 2H)	1.37, 1.29 (s, 9H)
<b>2</b>	-	8.46 (s, 1H)	7.53, 7.47 (d, 1H)	4.99 (bt, 2H)	3.72; 3.64 (dd, 2H); (dd, 2H)	1.38, 1.30 (s, 9H)
<b>3</b>	-	9.08 (s, 1H)	7.66, 7.53 (d, 1H)	4.58 (t, 1H)	5.25 (d, 1H) 5.13 (m, 2H) 4.70 (d, 1H) 4.04 (dd, 2H)	1.51, 1.35 (s, 9H)

1

2 In the oxidovanadium(v) complex **3**,  $\text{H}_4\text{L}^{\text{Saltris}}$  coordinates in a tetradentate, trianionic  
3 fashion *via* phenolato, aldimino and two alcoholato groups, in stark contrast to **1** and **2**.  
4 In principle,  $^1\text{H}$  NMR spectrum of **3** agrees well with a trigonal bipyramidal coordination  
5 mode, which is common for vanadium(v) complexes. In the absence of single crystal XRD  
6 and  $^{51}\text{V}$  NMR measurements, comparable 5-coordinate solution-state structure has also  
7 been proposed for several very similar vanadium(v) complexes.[59] However, in our  
8 case, single crystal XRD reveals that **3** is dinuclear in the solid state (see discussion  
9 below), with both vanadium centers adopting a distorted octahedral coordination mode.  
10 Furthermore,  $^{51}\text{V}$  NMR shows only one signal, which supports the dinuclear configuration  
11 in solution state as well, since both vanadium centers are chemically equivalent.  
12 Generally speaking, 5-coordinate vanadium complexes may isomerize to 6-coordinate  
13 species *via*, for example, solvent ligand exchange, and vice versa. Furthermore, 6-  
14 coordinate vanadium complexes may have multiple isomers if a solvent molecule is  
15 coordinated. These dynamics are frequently observed as multiple signals in the  $^{51}\text{V}$  NMR,  
16 although the choice of NMR solvent may or may not greatly influence this. The single  $^{51}\text{V}$   
17 NMR signal supports the notion that **3** is 6-coordinate, and is rigid *i.e.* does not contain  
18 any coordinated solvent molecules in the solution state.

19 In **3**, the aldimine carbon proton and aromatic protons have been moderately shifted  
20 downfield for *ca.* 0.34 and 0.27 ppm, respectively, relative to  $\text{H}_4\text{L}^{\text{Saltris}}$ . Similarly to **1** and  
21 **2**, where the tris-alcohol sidearms become inequivalent relative to the free ligand, in **3**  
22 these changes are much more pronounced. For example, the methylene CH<sub>2</sub> protons for  
23 the two alcoholato sidearms, have been shifted downfield by 0.79 – 1.35 ppm. They also  
24 appear separately as diastereotopic duplets, due to formation of rigid 5- and 6-membered  
25 ring structures, respectively. The single free alcohol sidearm, on the other hand, appears  
26 0.59 ppm downshifted for OH, and 0.13 ppm for the CH<sub>2</sub> protons, respectively.

27 The  $^{13}\text{C}$  NMR spectrum of **3** is very similar to those of **1** and **2**. Most striking features are  
28 the collapse of the equivalent (in  $\text{H}_4\text{L}^{\text{Saltris}}$ ) tris-alcohol carbons to inequivalent carbons in  
29 **3**, with approx. 15 ppm shift downfield observed for the two coordinating alcoholato  
30 methylene protons. Strikingly, the  $^{51}\text{V}$  NMR spectrum of **3** shows only one signal at *ca.* –  
31 562 ppm. The simple nature of the  $^{51}\text{V}$  NMR spectrum points towards that **3** is present, in  
32 solution, as a one stable species, and that there is probably little if any solution dynamics

1 commonly encountered with vanadium compounds. The  $^{51}\text{V}$  NMR observation is well in  
 2 line with single-crystal XRD results, in that the complex unit is symmetric, and both  
 3 oxidovanadium(v) centers adopt rigid six-coordinate distorted octahedral  
 4 configurations, making the vanadium centers chemically equivalent. The obtained  $^{51}\text{V}$   
 5 NMR spectrum is closely similar to those of other related oxidovanadium(v)  
 6 complexes.[49] Furthermore, the  $^{51}\text{V}$  NMR signal is present in the expected general area<sup>†</sup>  
 7 attributable to oxidovanadium systems bearing redox-innocent  $\text{O}_4\text{N}$  type ligands.[33,68]

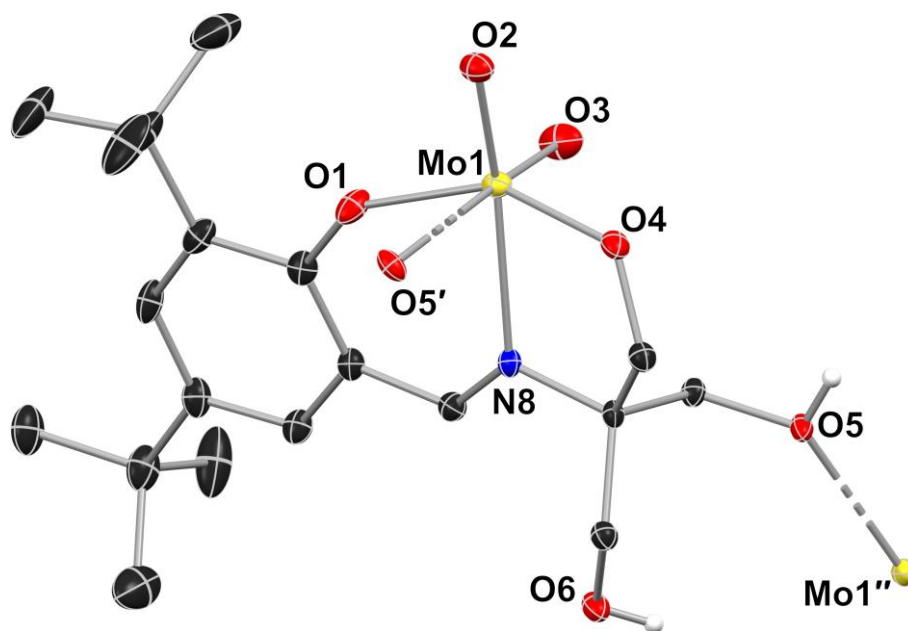
## 8 Description of crystal and molecular structures

9 Good quality single crystals of **1–3** could be grown thus enabling their structural  
 10 characterization by single crystal X-ray diffraction methods. **1** and **2** crystallized directly  
 11 from the reaction mixtures, *i.e.* from methanol as brightly yellow needle-like crystals (**1**)  
 12 and pale yellow needle-like crystals (**2**). **3**, on the other hand, deposited from the  
 13 methanol reaction mixture in a non-crystalline form. Umber-colored block-like crystals  
 14 could be obtained from boiling acetonitrile upon slow cooling. All three complexes  
 15 crystallize in the monoclinic crystal system and have the same space group ( $P2_1/c$ ).

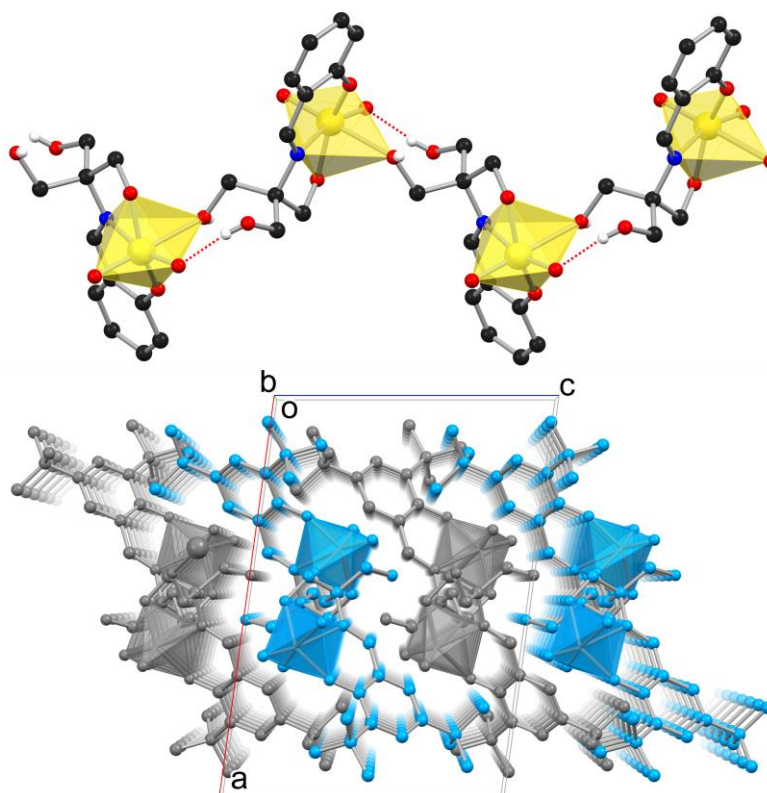
16 The X-ray structure of **1** (Figures 1 and 2) reveals that the reaction between molybdenum  
 17 precursor and  $\text{H}_4\text{L}^{\text{Saltris}}$  has involved the deprotonation of two proligand OH groups – the  
 18 phenolic and one of the three alcoholic OH moieties – and coordination of the respective  
 19 O atoms to the dioxidomolybdenum(vi) center. The rather distorted octahedral  
 20 coordination sphere of the Mo(vi) ion is fulfilled by the aldimino N donor and one of the  
 21 alcoholic OH groups of an adjacent complex unit (for selected bond parameters, see  
 22 Figure 1). A CSD survey[69] of dioxidomolybdenum(vi) complexes with similar trisbase  
 23 salicylaldimine-type ligands reveals that in such complexes only one of the three available  
 24 alcoholic groups tends to coordinate to the central metal ion, similarly to **1**. Furthermore,  
 25 the remaining sixth coordination site in the previously reported complexes is occupied  
 26 by a solvent molecule ( $\text{H}_2\text{O}$ , MeOH, DMF or DMSO), which leads to discrete molecular  
 27 complexes. In the crystal structure of **1**, however, the distinguishable complex units  
 28 couple together via  $\text{Mo}—\text{O}_{\text{alcohol}}$  bonds thus creating a polymeric 1D chain along the  
 29 crystallographic *b*-axis. The polymeric system is further stabilized by intrapolymer  
 30  $\text{Mo}=\text{O}\cdots\text{H}—\text{O}$  [ $d(\text{O}2\cdots\text{O}6') = 2.65 \text{ \AA}$ ] hydrogen bonds (HB) as well as interpolymer HBs  
 31 occurring *via*  $\text{Mo}—\text{OH}\cdots\text{OHCH}_2\text{CN}$  [ $d(\text{O}5'\cdots\text{O}6'') = 2.63 \text{ \AA}$ ] contacts. It is noteworthy that,  
 32 albeit being polymeric in the solid state, **1** dissolves readily in DMSO which can be  
 33 attributed to the degradation of the polymeric chains into the respective monomeric  
 34 complex units (see NMR analysis above). This confirms the lability of the  $\text{Mo}—\text{O}_{\text{alcohol}}$   
 35 bonds and the discrete nature of the complex upon dissolution.

---

<sup>†</sup> V(v) complexes bearing a redox-innocent  $\text{O}_5\text{N}_1$  donor set generally appear between *ca.* –600 to –400 ppm according to ref. [33] and at lowest *ca.* –650 ppm according to ref. [68] ( $\delta$  ppm vs.  $\text{VOCl}_3$ ).



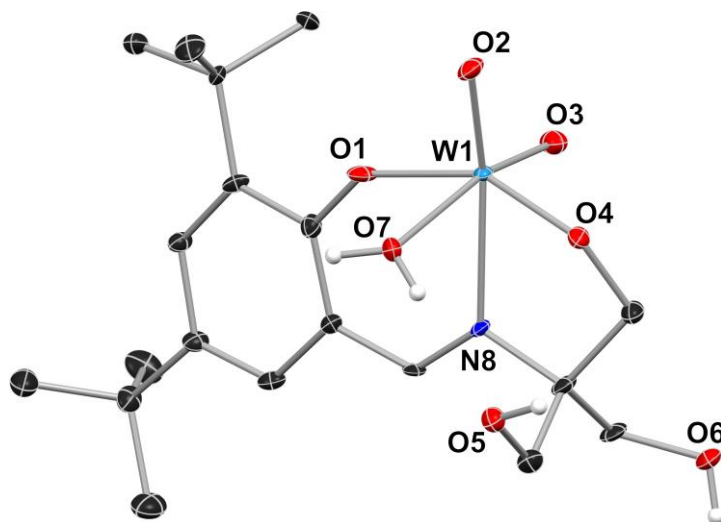
1 **Figure 1.** Presentation of the asymmetric unit of **1** with bonds to the neighboring complex  
 2 units shown in dashed lines. All C—H hydrogen atoms are omitted for clarity. Principal  
 3 ellipsoids are presented at the 50 % probability level. Relevant bond lengths (Å): Mo1—  
 4 O1 = 1.9300(14), Mo1—O2 = 1.7237(12), Mo1—O3 = 1.6942(13), Mo1—O4 =  
 5 1.9060(13), Mo1—O5' = 2.4123(13) and Mo1—N8 = 2.2912(14). Symmetry operations:  
 6 (') = 1-x, ½+y, ½-z; (')' = 1-x, -½+y, ½-z.



7 **Figure 2.** Partial view of the 1D polymeric structure of **1** (top) and illustration of packing  
 8 of the discrete polymers (alternating blue and grey colors, below) viewed along the  
 9 crystallographic *b*-axis. Top: C—H hydrogen atoms and *tert*-butyl groups omitted. Below:  
 10 All hydrogen atoms omitted.



1 In contrast to **1**, the respective tungsten analogue **2** crystallizes as monomeric units in  
 2 which the  $\text{WO}_2$  unit is bound to a dianionic tridentate  $\text{H}_2\text{L}^{\text{Saltris}}$  ligand in agreement with  
 3 the NMR analyses. The remaining coordination site, which is *trans* to O3, has been taken  
 4 up by a water molecule (Figure 3). Although there is one example of tungsten complex  
 5 bearing a very similar ligand, the  $[\text{WO}_2(\text{sapd})]$  ( $\text{sapd}$  = salicylidene propanediol)[70], to  
 6 the best of our knowledge, **2** is the first report of a tungsten complex characterized by  
 7 XRD that bears a Saltris-type Schiff base ligand. It should be noted, that the solid state  
 8 structure of **2** closely resembles some known Mo-complexes<sup>‡</sup> such bearing Saltris-like  
 9 ligands.[58,71] In **2**, two of the three ligand alcoholic OH groups remain uncoordinated  
 10 and are thus available for hydrogen bonding. In case of **2**, this leads to an intricate  
 11 hydrogen bonding network in the crystal lattice in which the monomeric units build into  
 12 2D sheets, terminated by the aliphatic regions of the molecules (ESI, Figure S59).

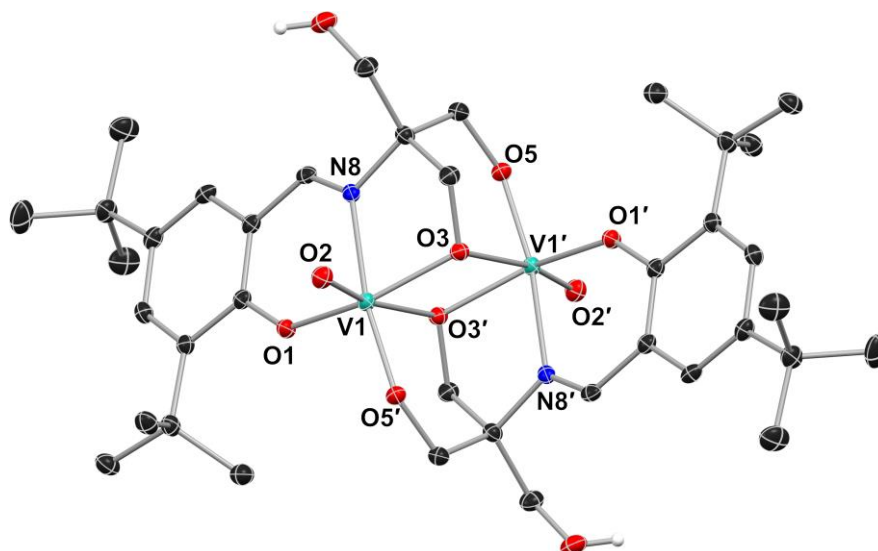


13 **Figure 3.** Illustration of the asymmetric unit of structure of **2**. All C—H hydrogen atoms  
 14 are omitted for clarity. Principal ellipsoids are presented at the 50 % probability level.  
 15 Relevant bond lengths (Å): W1—O1 = 1.928(4), W1—O2 = 1.764(4), W1—O3 = 1.715(4),  
 16 W1—O4 = 1.902(4), W1—O7 = 2.309(4) and W1—N8 = 2.283(4).

17 X-ray structure of **3** shows a formation of a dinuclear vanadium complex consisting of  
 18 two oxidovanadium(v) centers and two ligands wherein each ligand is coordinated in  
 19 tetradentate trianionic fashion (Figure 4). The crystal structure is solved with half of the  
 20  $[\text{VO}(\text{HL}^{\text{Saltris}})]_2$  dimer constituting the asymmetric unit while the complete  
 21 centrosymmetric complex is generated via an inversion center between the V atoms. Also,  
 22 an acetonitrile solvent molecule is located in the asymmetric unit. The crystallo-  
 23 graphically unique complex moiety comprises of oxidovanadium(v) ion chelated in ONO  
 24 fashion by phenolato (O1), aldimino (N8) and alcoholato (O3) groups of  $\text{HL}^{\text{Saltris}}$ . The  
 25 dimer is upheld by alcoholato atoms O3 and O3', which act as bridging donors between  
 26 the V(v) centers, and alcoholato atoms O5 and O5' which coordinate to neighboring V1'  
 27 and V1, respectively. This bonding scheme yields an approximately planar (maximum  
 28 deviation from planarity is ca. 17°)  $\text{V}_2\text{O}_6$  system to which two alcoholato and two  
 29 aldimino groups coordinate perpendicularly. The V—O distances in the V1—O3/O3'—

<sup>‡</sup> Solid state structure of **2** is very similar to  $\text{MoO}_2$  complexes **1–5** from ref. [58] and **1–3** from ref. [71].

1 V1' bridges show high degree of asymmetry (ca. 0.4 Å). Structurally, **3** is very similar to  
 2 some other reported dimeric vanadium complexes bearing analogous ligands.[49,66,72]



3 **Figure 4.** Presentation of the  $[VO(HL^{\text{Saltris}})]_2$  complex found in the crystal structure of **3**.  
 4 All C—H hydrogen atoms are omitted for clarity. Principal ellipsoids are presented at  
 5 the 50 % probability level. Relevant bond lengths (Å): V1—O1 = 1.8646(12), V1—O2 =  
 6 1.6009(13), V1—O3 = 1.9028(12), V1—O3' = 2.3066(12), V1—O5' = 1.7981(12) and  
 7 V1—N8 = 2.1323(15). Symmetry operations: (') = -x, 1-y, 1-z.

## 8 X-ray powder diffraction analysis

9 Bulk samples of **1–3** were analyzed by powder X-ray diffraction (PXRD). The unit cell  
 10 parameters of complexes **2** and **3** were determined by Pawley analysis[73] within the  
 11 HighScore Plus 4.7 program using the cell parameters of corresponding single crystal  
 12 structures as the starting point of the least-squares refinements. The variable parameters  
 13 in the refinements were zero-offset, unit cell and peak profile parameters. The fitted  
 14 diffraction graphs for **2** and **3** are found in ESI figures S61–62 whereas the refined unit  
 15 cells and the resulting R-factors and goodness-of-fit values are presented in ESI table S2.  
 16 For **2**, due to instrument limitations, we could not record one of the major low angle peaks  
 17 which resides at ca. 4.5° according to the simulated single crystal pattern.

18 In case of complex **1**, the experimental PXRD pattern shows some similarities with the  
 19 pattern simulated from the respective single crystal structure but it is in general  
 20 distinctively different (ESI figure S60). We suspect that this may arise from a degradation  
 21 of the polymeric structure of **1** while preparing the sample of PXRD analysis by grinding  
 22 of the crystalline bulk material. In contrast, the unit cell parameters of bulk materials of  
 23 **2** and **3**, derived from the Pawley analysis, are in good agreement with the corresponding  
 24 single crystal structures and thus demonstrate their structural similarity. The refined  
 25 unit cells (PXRD) show ca. 2 % increase in unit cell volume which is reasonable  
 26 considering the thermal expansion due to the differences in data recording temperature  
 27 between the single crystal and powder data (120 vs. 298 K)

## 1 Cyclic voltammetry

2 To gain insight into the reactivity of the complexes, and specifically to study the ability of  
 3 **1–3** to undergo reversible redox-reactions relevant regarding catechol oxidation,  
 4 electrochemical investigations were undertaken. The cyclic voltammograms of **1–3** are  
 5 presented in the electronic supplementary material, while the electrochemical data has  
 6 been tabulated in table 3. **3** has a reversible electrochemical response with  $E_{1/2} = -415$  mV  
 7 with Pt working electrode, and  $E_{1/2} = -422$  mV using GC working electrode, respectively.  
 8 This response has been assigned as the V(v)/V(iv) redox-couple, since it is also observed  
 9 in the same potential area with some other structurally similar<sup>§</sup> Schiff base  
 10 oxidovanadium(v) complexes.[49,59]. With higher scan rates, slight quasi-reversibility is  
 11 observed, although  $E_{1/2}$  does not change significantly. In the positive potential range three  
 12 irreversible oxidation responses occur at  $E_{1/2} = ca.$  +900 mV, +1.496 and +1.683 V (Pt WE),  
 13 that are most likely ligand-centered, since they are also varyingly present in **1** and **2**. For  
 14 example, in some unrelated complexes phenoxyl radical formation has been suggested to  
 15 occur at approx. +650–750 mV.[74] Accordingly, we tentatively assign the  $\sim +900$  mV  
 16 redox-response to  $PhO \rightarrow PhO^{\bullet+}$ , and subsequent oxidation events at higher potentials to  
 17 alkoxy-centered oxidations *i.e.*  $RO \rightarrow RO^{\bullet+}$ . The cyclic voltammograms of **1** and **2** are  
 18 remarkably featureless in comparison to those of **3**. All redox-processes present in **1** and  
 19 **2** are irreversible, with the one-electron reductions *e.g.* Mo(VI)  $\rightarrow$  Mo(V) and W(VI)  $\rightarrow$   
 20 W(V) tentatively assigned as  $-0.896$  (Pt) and  $-1.296$  V (GC), respectively.[66] Similar  
 21 ligand-centered irreversible oxidation events are present in **1** and **2** than what is found  
 22 in **3**.

**Table 3.** Electrochemistry data obtained for **1–3**.

Complex	ligand-centered <i>e.g.</i> $RO \rightarrow RO^{\bullet+}$		ligand-centered <i>e.g.</i> $RO \rightarrow RO^{\bullet+}$		ligand-centered <i>e.g.</i> $PhO \rightarrow PhO^{\bullet+}$		metal-centered $M(ox)/M(ox+1)$	
	Pt	GC	Pt	GC	Pt	GC	Pt	GC
<b>1</b>	–	+2.159	+1.691	+1.717	–	+0.942	–	–0.986 irrev.
<b>2</b>	–	–	+1.653	+1.684	+0.934	+0.959	–	–1.296 irrev.
<b>3</b>	+1.683	1.682	+1.496	+1.484	+0.896	+0.917	–0.415 ps. rev.	–0.422 ps. rev.

The values are given in V vs. Ag/AgCl calibrated against Fc/Fc<sup>+</sup> redox-couple.[54] GC and Pt represent glassy carbon and platinum working-electrodes (WE), respectively.

23

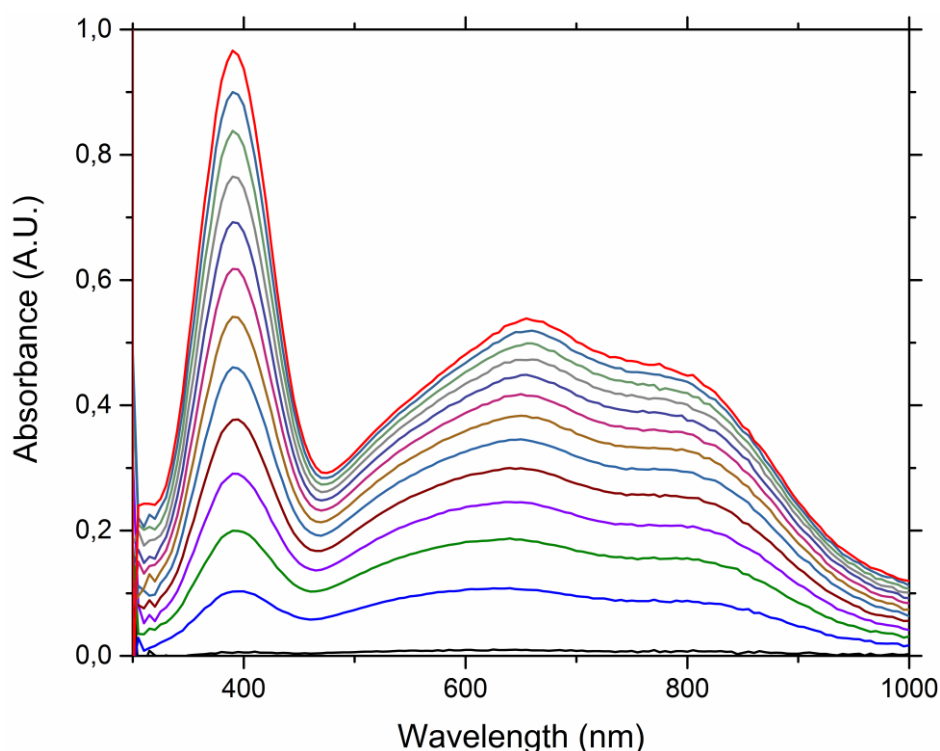
## 24 Catechol oxidase mimetic activity

25 In various catechol oxidase studies 3,5-DTBC is the substrate of choice[19,20,28], owing  
 26 to its relatively easy oxidation due to its low redox potential.[75] As such, the catechol  
 27 oxidase mimetic activity of all complexes was studied in the aerial oxidation of 3,5-DTBC  
 28 as well as 4-*tert*-butylcatechol (4-TBC) and pyrocatechol in chloroform, acetonitrile and

<sup>§</sup> Structurally related V(v) complexes such as **3** and **6–8** from ref [49] and **1, 2, 4** and **5** from ref. [59] display a V(iv)/V(v) redox couple at  $E_{1/2}$  values  $-470$  to  $-400$  mV vs. Ag/AgCl in DMF.

1 methanol. 4-TBC and pyrocatechol were chosen because they are more resistant to  
2 autoxidation compared to 3,5-DTBC.

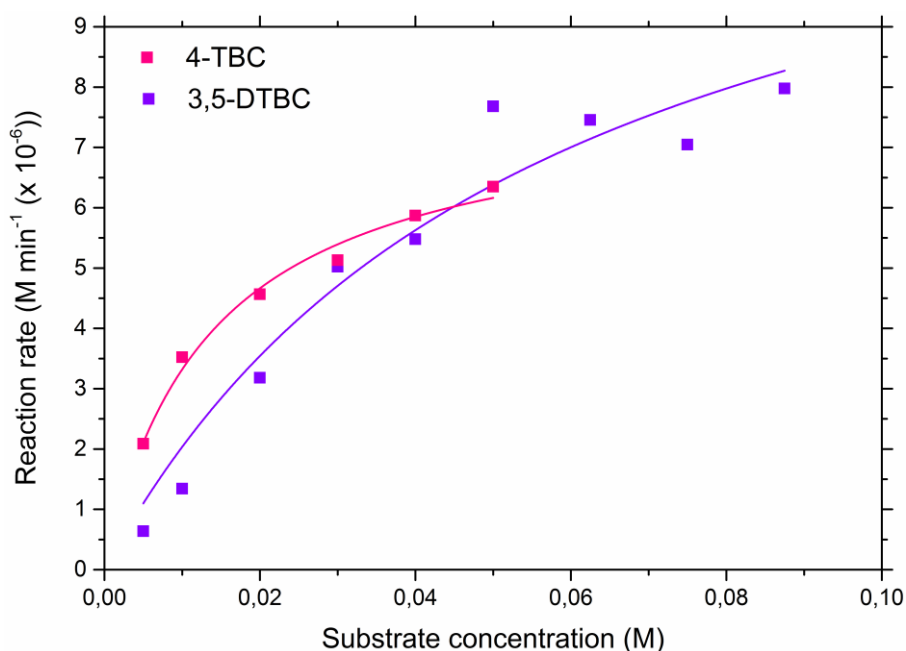
3 Preliminary investigations indicated that 3,5-DTBC and 4-TBC readily reacted to form the  
4 corresponding *o*-benzoquinones, namely 3,5-di-*tert*-butyl-1,2-benzoquinone (3,5-DTBQ)  
5 and 4-*tert*-butyl-1,2-benzoquinone (4-TBQ), respectively, when **3** was used as the  
6 catalyst in CHCl<sub>3</sub>, MeOH and MeCN. However, **1** and **2** showed no noticeable catechol  
7 oxidase-like activity in the same time frame, although UV-Vis and ESI-HRMS  
8 measurements confirmed that the catechols coordinate to the metal centers (ESI Figures  
9 S35–S46). Furthermore, pyrocatechol was observed not to react in any solvent with any  
10 of the catalysts. With **3**, catechol coordination was also evident from the fact that all of  
11 the reaction solutions turned blue after the addition of the catechols, a characteristic  
12 color for vanadium-catecholato species[31], with also typical vanadium-catecholato  
13 LMCT bands observable in the UV-Vis spectra at *ca.* 650 and 830 nm (figure 5).[31,33,76]  
14 Furthermore, the solutions eventually turned green and finally yellow-brown due to the  
15 formation of *o*-benzoquinones. UV-Vis spectroscopic measurements confirmed the UV  
16 bands of 3,5-DTBQ and 4-TBQ, that appear at approx. 400 nm.[56,57]



17 **Figure 5.** The formation characteristic vanadium-catecholato LMCT bands at approx. 650  
18 and 830 nm during 3,5-DTBC oxidation by **3**. The reaction was monitored for 1 hour and  
19 spectra were recorded at every 5-minute interval.

20 From the kinetics data it can be concluded that the oxidation of 4-TBC and 3,5-DTBC  
21 catalyzed by **3** follows typical Michaelis–Menten like kinetics (figure 6). At low substrate  
22 concentrations, the measured absorbance increases linearly as a function of time,  
23 indicative of a first order reaction with respect to the substrate. However, upon  
24 significant excess (> 10 000-fold) of the substrate relative to catalyst the reaction rate

1 becomes pseudo zero-order. At this regime subsequent substrate additions will not speed  
 2 up the reaction, as the catalyst becomes saturated. The kinetic parameters  $V_{\max}$   
 3 (maximum reaction rate), Michaelis-constant  $K_M$  ( $[S]$  at  $\frac{1}{2}V_{\max}$ ) and  $k_{\text{cat}}$  (turnover  
 4 frequency) are given in table 4. From the kinetics data it can be seen that **3** compares well  
 5 with other oxidovanadium systems, with its catalytic activity falling well within the range  
 6 of that of others (table 4).[18–21] However, the most remarkable aspect is the fact that **3**  
 7 offers a comparable catechol oxidase-like activity to the structurally very similar dicobalt  
 8 complex  $[\text{Co}(\text{H}_2\text{L}^{\text{Saltris}})(\text{OAc})]_2$ . [28]



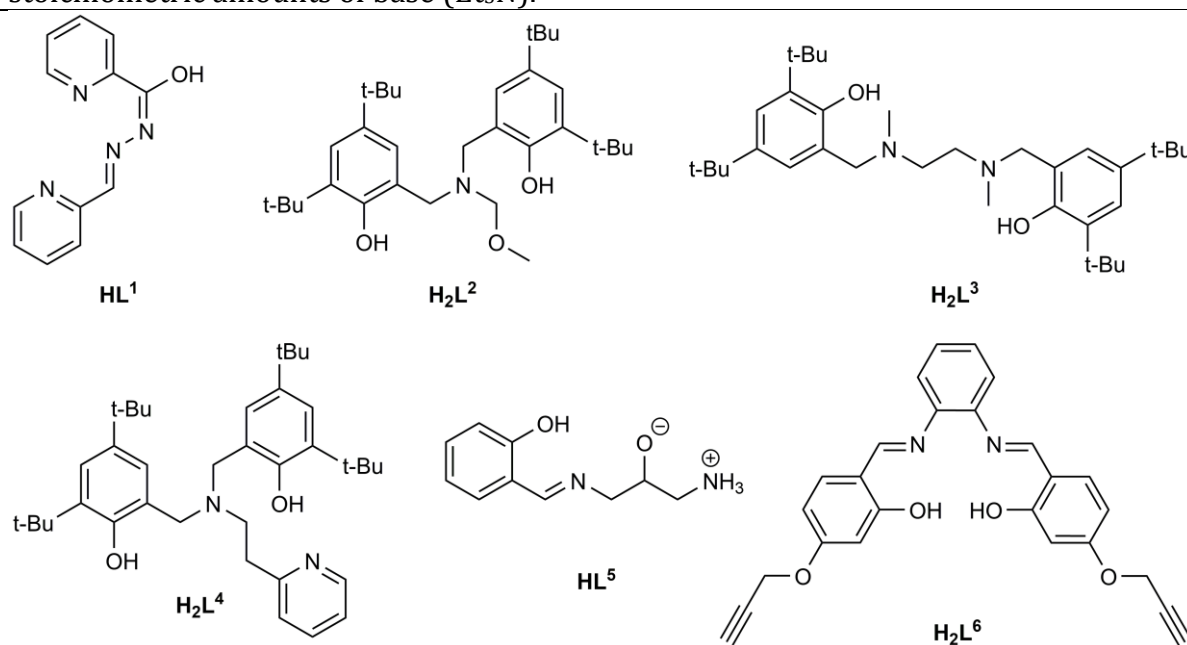
9 **Figure 6.** Michaelis–Menten plots of the oxidations of 4-TBC and 3,5-DTBC by **3** in  $\text{CHCl}_3$ .

10 While the natural catechol oxidase catalyzes the oxidation of catechols to corresponding  
 11 *o*-benzoquinones, producing water[15] as the sole by-product, a number of model  
 12 transition metal complexes produce hydrogen peroxide as well.[28,77–81] Recently, a  
 13 dicobalt complex  $[\text{Co}(\text{H}_2\text{L}^{\text{Saltris}})(\text{OAc})]_2$  structurally strikingly similar (lacking oxido  
 14 ligands) to **3** was assessed in the oxidation of 3,5-DTBC.[28] Mechanistic investigations  
 15 involving MS measurements strongly hint towards a single and double 3,5-DTBSQ  
 16 adducts  $[(\text{Co}(\text{III})(\text{HL}^{\text{Saltris}})\text{Co}(\text{II})(\text{H}_2\text{L}^{\text{Saltris}})(3,5\text{-DTBSQ}))]$  and  $[\text{Co}(\text{II})(\text{H}_2\text{L}^{\text{Saltris}})(3,5\text{-}$   
 17  $\text{DTBSQ})]_2$ . Some studies highlight the importance of metal–metal distance, which may  
 18 give insights into the overall catalytic mechanisms of any given catechol oxidase model  
 19 system.[78,79] In light of the obvious structural similarities between the dinuclear  
 20 cobalt(III) complex and our dinuclear oxidovanadium(v) complex, as well as the fact that  
 21 there are only a limited[20,23] mechanistic investigations on catechol oxidation involving  
 22 modern oxidovanadium systems, we conducted mechanistic investigations of our own  
 23 using UV-Vis,  $^{51}\text{V}$  NMR and ESI-HRMS spectrometry. The detailed descriptions of the  
 24 experiments are given in supplementary material.

**Table 4.** Catechol oxidation results for the present  $[\text{VO}(\text{HL}^{\text{Saltris}})]_2$  as well as some other selected vanadium-based catechol oxidase mimetic systems.

Compound	$k_{\text{cat}}$ ( $\text{h}^{-1}$ )	$K_{\text{M}}$ (M)	$V_{\text{max}}$ ( $\text{M min}^{-1}$ )	Ref.
<b>3</b>	164 $\pm$ 32 <sup>a</sup>	0.05727	$1.37 \times 10^{-5}$	This work
<b>3</b>	94 $\pm$ 4 <sup>b</sup>	0.01363	$7.84 \times 10^{-6}$	This work
$[\text{VO}(\text{L}^1)]_2 \text{SO}_4$	1439 <sup>bc</sup>	–	–	[18]
$[\text{VO}(\text{OMe})(\text{L}^2)]$	12 <sup>a</sup>	0.00115	$1.01 \times 10^{-5}$	[21]
$[\text{VO}(\text{OMe})(\text{L}^3)]$	13 <sup>a</sup>	0.00107	$1.04 \times 10^{-5}$	[21]
$[\text{VO}(\text{OMe})(\text{MeOH})(\text{L}^4)]$	3 <sup>a</sup>	0.00557	$7.66 \times 10^{-6}$	[19]
$[\text{Co}(\text{H}_2\text{L}^{\text{Saltris}})(\text{OAc})_2$	80 <sup>a</sup>	0.0087	$1.33 \times 10^{-5}$	[28]
$[\text{VO}_2(\text{L}^5)]$	2063 <sup>a</sup>	0.00079	$5.73 \times 10^{-5}$	[20]
$[\text{VO}(\text{L}^6)]$	395 <sup>a</sup>	0.00103	$6.58 \times 10^{-3}$	[23]

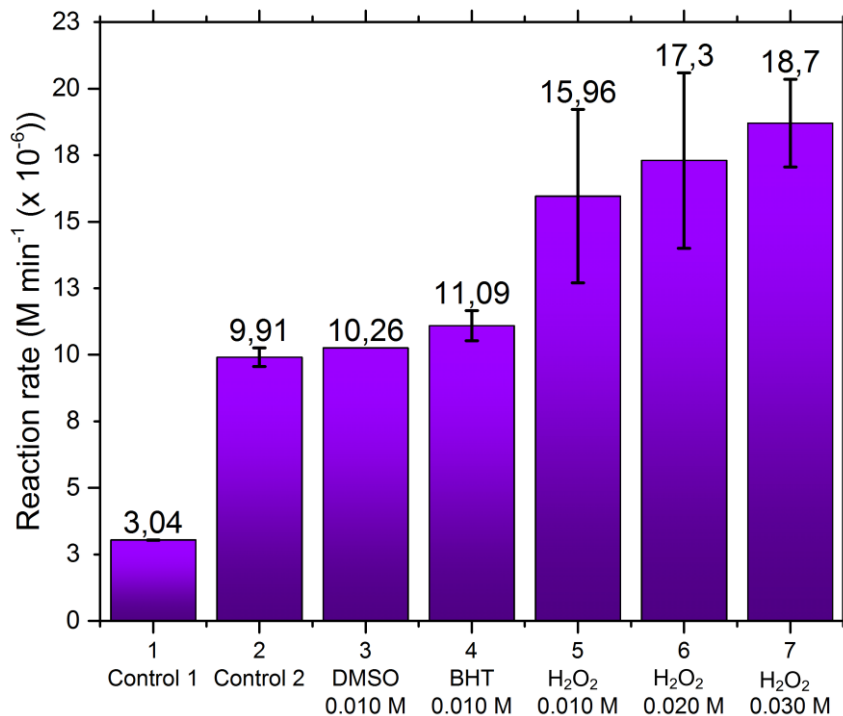
<sup>a</sup> = For oxidation of 3,5-DTBC. <sup>b</sup> = For oxidation of 4-TBC. <sup>c</sup> = In the presence of stoichiometric amounts of base ( $\text{Et}_3\text{N}$ ).



1

2 The initial reaction rate of the oxidation of 4-TBC by 0.1 mol-% of **3** was measured by UV-  
 3 Vis spectroscopy in the presence of DMSO (hydroxyl radical quencher), BHT (butyrated  
 4 hydroxytoluene, radical inhibitor), and varying concentrations of  $\text{H}_2\text{O}_2$  in chloroform  
 5 (figure 7). The addition of DMSO or BHT (entries 3 and 4) have little effect on the initial  
 6 reaction rate when compared to the effects of **3** alone (entry 2), so it may be concluded  
 7 that hydroxyl radicals or any other radicals most likely do not significantly contribute to  
 8 the overall oxidation mechanism. However, hydrogen peroxide seems to accelerate the  
 9 oxidation to some extent in the presence of **3**, with reaction rate increasing approx.  
 10 linearly with increasing  $\text{H}_2\text{O}_2$  concentration (entries 5–7). However, and importantly,  
 11 even in relatively high concentrations  $\text{H}_2\text{O}_2$  does not significantly oxidize 4-TBC in the  
 12 absence of **3** (entry 1), so the reaction rate enhancement most probably does not come  
 13 from the direct substrate oxidation by  $\text{H}_2\text{O}_2$ . Indeed, it has been previously shown that

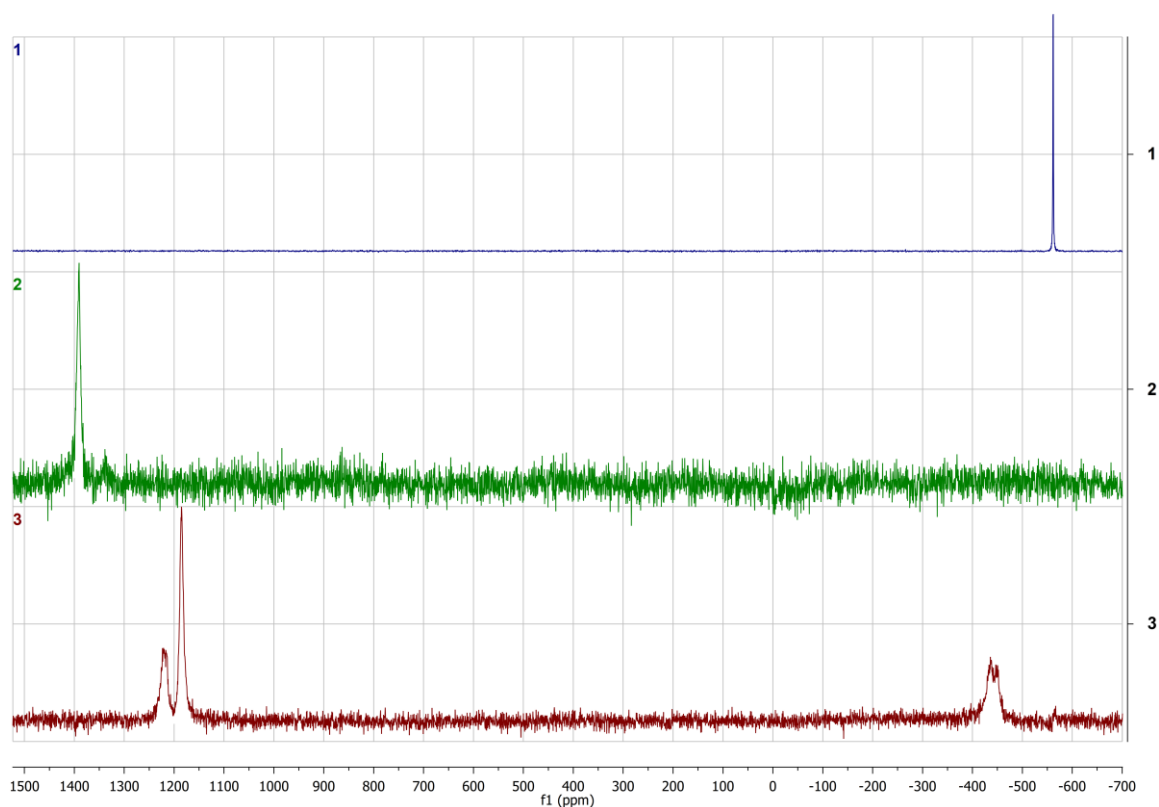
1 H<sub>2</sub>O<sub>2</sub> may assist the dissociation, or leaching, of vanadium from vanadium-containing  
 2 species, which then rapidly convert to catalytically active catecholato-bearing vanadium  
 3 species *e.g.* [VO(3,5-DTBC)(3,5-DTBSQ)] and [V(3,5-DTBC)<sub>3</sub>] in the presence of very large  
 4 excess of catechol, 3,5-DTBQ and H<sub>2</sub>O<sub>2</sub> relative to vanadium.[36,38] In a separate  
 5 experiment, an iodometric assay was done to confirm the formation of hydrogen peroxide  
 6 in our system.



7 **Figure 7.** Initial reaction rate of 4-TBC (0.1 M) oxidation over the period of 20 minutes in  
 8 the presence of **3** ( $1 \times 10^{-4}$  M) and/or several additives. See text for further information.  
 9 Control 1: 0.1 M 4-TBC + 0.030 M H<sub>2</sub>O<sub>2</sub> in the absence of **3**. Control 2: 0.1 M 4-TBC + **3**  
 10 in the absence of H<sub>2</sub>O<sub>2</sub>. All measurements run in duplicate.

11 <sup>51</sup>V NMR experiments were undertaken to determine qualitatively the nature of  
 12 vanadium-containing species formed during catechol oxidation, and to gain some insight  
 13 into the catalytically active vanadium species. In the experiments, 100 equivalents of 3,5-  
 14 DTBC and 4-TBC were reacted with **3** in CDCl<sub>3</sub> at RT, and the <sup>51</sup>V NMR spectra were  
 15 recorded immediately. The <sup>51</sup>V NMR spectra obtained from these experiments drastically  
 16 differed from the original <sup>51</sup>V NMR spectra of **3** (figure 8). In both cases, the original signal  
 17 corresponding to **3** at *ca.* -562 ppm vanished completely, which is a clear indication of  
 18 dynamic and very rapid changes to the first coordination sphere of **3**. New, very broad  
 19 signals were observed at *ca.* +1221 and +1185 with 4-TBC and at *ca.* +1391 with 3,5-  
 20 DTBC *viz.* significantly downfield relative to VOCl<sub>3</sub>.

21



1 **Figure 8.**  $^{51}\text{V}$  NMR spectra of 1) **3** in acetone- $d_6$ , 2) **3** + 100 eq. 3,5-DTBC in  $\text{CDCl}_3$ . and 3)  
2 **3** + 100 eq. 4-TBC in  $\text{CDCl}_3$ .

3 These prominent changes, and the absence of  $^{51}\text{V}$  NMR signals at approx.  $-400 - (-600)$   
4 ppm, the expected region for oxidovanadium(V) complexes bearing a redox-innocent  $\text{NO}_2$   
5 donor set, indicate that there are most likely no vanadium species bearing  $\text{HL}^{\text{Saltris}}$   
6 left.[33,68] In fact, the significantly downfield signals are almost certainly related to  
7 diamagnetic catecholato-bearing vanadium species with at least two or even three  
8 catechol ligands, since many reported non-innocent monocatecholato oxidovanadium(V)  
9 systems with mixed ON donors appear upfield, and at best only slightly downfield of  
10  $\text{VOCl}_3$ . [33] It should be noted, that the  $^{51}\text{V}$  NMR spectra did not drastically change over a  
11 period of two days. Furthermore, when 4-TBC was used a signal was observed at  $-438$   
12 ppm, only slightly downfield of **3**. This species has been tentatively assigned to  
13  $[(\text{VO}(\text{HL}^{\text{Saltris}})(\text{O}-4\text{-TBC}))]$ , where 4-TBC coordinates in a singly anionic monodentate  
14 fashion. These assignments are in line with observations from ESI-MS studies (see  
15 below), where bis- and tris(catecholato) oxidovanadium and non-oxidovanadium species  
16 are detected.

17 To supplement the rather cautious assignments done in the  $^{51}\text{V}$  NMR section above, the  
18 same NMR samples were analyzed by ESI-HRMS in acetonitrile. Some reported  
19 mechanistic studies investigating catechol oxidation by oxidovanadium complexes  
20 frequently involve ESI-MS only in the positive mode.[20,23] However, as per the findings  
21 by Finke and co-workers related to the nature of the true active catalyst, negative mode



1 MS studies might also reveal important information, and should always be run.[36]  
2 Accordingly, we report both positive and negative MS results.

3 In the case of **3** + 3,5-DTBC, and in the negative mode, the main ionization product was  
4  $[V(3,5\text{-DTBC})_3]^-$  at  $m/z = 711.3884$  (calcd.  $m/z = 711.3835$ ). Other species in the negative  
5 mode are  $[VO(3,5\text{-DTBC})_2]^-$  at  $m/z = 507.2336$  (calcd.  $m/z = 507.2321$ ). A very low-  
6 intensity signal can be observed for  $[VO(\text{HL}^{\text{Saltris}})(3,5\text{-DTBC})]^-$  at  $m/z = 622.2976$  (calcd.  
7  $m/z = 622.2954$ ). In the positive mode the only clearly detected species is  $[3,5\text{-DTBQ} +$   
8  $\text{Na}]^+$  at  $m/z = 243.1400$  (calcd.  $m/z = 243.1356$ ). However, there exists a very low-  
9 intensity (*ca.* 2 %) signal at  $m/z = 646.2889$ . We have tentatively assigned this to the  
10 species  $[VO(\text{H}_2\text{L}^{\text{Saltris}})(O\text{-}3,5\text{-DTBC}) + \text{Na}]^+$ , which has a calculated  $m/z = 646.2919$ . There  
11 are no traces of the intact **3** in either positive or negative polarization. From this data, the  
12 detection of  $[VO(3,5\text{-DTBC})_2]^-$  or, perhaps  $[V(\text{IV})O(3,5\text{-DTBC})(3,5\text{-DTBSQ})]^-$ , is  
13 significant, as it has been linked to the Pierpont's complex  $[V(\text{V})O(3,5\text{-DTBC})(3,5\text{-}$   
14  $\text{DTBSQ})]_2$ , which is the dimerized resting state of the supposed active catalyst  $[VO(3,5\text{-}$   
15  $\text{DTBC})(3,5\text{-DTBSQ})]$ . [38] Pierpont's complex, having a calculated  $m/z = 1014.4631$ , is not  
16 detected, however. The MS spectra are presented in the ESI (figures S42–43).

17 With **3** + 4-TBC, a somewhat similar speciation is observed, however, the main species in  
18 the negative ionization corresponds to  $[VO(\text{HL}^{\text{Saltris}})(O\text{-}4\text{-TBC})]^-$  at  $m/z = 566.2312$   
19 (calcd.  $m/z = 566.2328$ ). The tris catecholato species  $[V(4\text{-TBC})_3]^-$  is also present at  $m/z$   
20  $= 543.1966$  (calcd.  $m/z = 543.1957$ ) with low intensity. These findings may suggest that  
21 in the presence of 4-TBC, which reacts slower than 3,5-DTBC, **3** is somewhat more  
22 inclined to form monocatecholato adducts. This would also give some credibility to the  
23  $^{51}\text{V}$  NMR assignment at  $-438$  ppm (see above). Similarly to the above case, the  
24  $[VO(\text{H}_2\text{L}^{\text{Saltris}})(O\text{-}4\text{-TBC}) + \text{Na}]^+$  low-intensity adduct is tentatively assigned to the  $m/z =$   
25  $590.2240$  observed in the positive mode (calcd.  $m/z = 590.2293$ ).  $[\text{3} + \text{Na}]^+$  is also  
26 detected in the positive ionization mode with  $m/z = 825.2625$  (calcd.  $m/z = 825.2706$ )  
27 unlike previously in the presence of 3,5-DTBC. The MS spectra are presented in the ESI  
28 (figures S44–S45).

29 After overnight reaction of 3,5-DTBC in the presence of 1 mol-% **3** three compounds were  
30 isolated (table 5 and ESI). The product distribution resembles that found in many 3,5-  
31 DTBC oxidation reactions catalyzed by a variety of different vanadium pre-  
32 catalysts.[35,36,38] For instance, the 7-membered 3,5-di-*tert*-butyl-1-oxacyclohepta-3,5-  
33 diene-2,7-dione (**4**) is generally obtained with the highest yield of 40–57 % of the  
34 reaction products. While our isolated yield falls slightly short of 40 %, it is still the main  
35 product at 31 % yield. The second to most abundant product is 3,5-di-*tert*-butyl-1,2-  
36 benzoquinone (**5**) with a mean yield of 9–25 % and agrees with our observations (yield  
37 = 23 %). Additionally, the dimeric 4',6,6',8-tetra-*tert*-butyl-3*H*-spiro[benzo[*b*][1,4]di-  
38 oxine-2,2'-pyran]-3-one (**6**) represents one of the low-yielding products (10–18 %), a fact  
39 which is also observed by us (18 % yield). However, we were unable to isolate, nor detect,  
40 the lactones 3,5-di-*tert*-butyl-2-pyrone (**7**) or 3,5-di-*tert*-butyl-5-(carboxymethyl)-2-  
41 furanone (**8**) which generally, and in combination, represent at most 20 % of the total

1 product yield. Accordingly, they might represent the “missing” 28 %, as the combined  
 2 *isolated* yield of our reaction amounts to only 72 %. TLC was however used to establish  
 3 that no 3,5-DTBC was present, and hence we estimate a minimum 95 % conversion. It is  
 4 important to emphasize that the formation of the products **4**, **6**, **7** and **8**, which represent  
 5 the dioxygenase intra- and extradiol cleavage products, have been linked[36,38] to the  
 6 vanadium species  $[V(3,5\text{-DTBC})_3]^-$  (detected with ESI-HRMS, see above), which is itself  
 7 intimately related to  $[VO(3,5\text{-DTBC})(3,5\text{-DTBSQ})]$  and the “common catalyst  
 8 hypothesis”.[36]

**Table 5.** The product distribution obtained by reacting 3,5-DTBC with air in boiling chloroform in the presence of 1 mol-% **3**.

Estimated conversion <sup>a</sup> (%)	Isolated product yield (%)				
	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>	<b>8</b>
≥95	31	23	18	nd <sup>b</sup>	nd <sup>b</sup>

<sup>a</sup> As determined by TLC. <sup>b</sup> Not detected. Reaction conditions: 500 mg 3,5-DTBC, 18 mg **3**, 30 mL CHCl<sub>3</sub>, reflux 16 h.

9 The mechanistic investigations described above provide compelling evidence in support  
 10 of the notion that neither **3**, nor a close structural derivative thereof, is the active catechol  
 11 oxidation catalyst. Consequently, a catalytic cycle resembling the one described for the  
 12 closely similar dicobalt system cannot be credibly proposed.[28] Instead, the results  
 13 obtained from UV-Vis, <sup>51</sup>V NMR, and from the ESI-HRMS and product distribution  
 14 experiments in particular, confer a rather different state of affairs. The combined results  
 15 clearly indicate that dioxygenase activity is presented during catalysis as well, and that  
 16 the system presented herein confers very similar behavior to many other vanadium  
 17 precatalysts. These results are decisively in favor of the “common catalyst  
 18 hypothesis”.[36]

### 19 Oxygen atom transfer activity

20 *cis*-Dioxidomolybdenum(VI) and -tungsten(VI) complexes are well known to catalyze  
 21 oxygen atom transfer (OAT) to suitable organic substrates such as, but not limited to,  
 22 benzoin and triphenylphosphine.[82–86] The propensity of Mo/W compounds to  
 23 catalyze this particular reaction is not at all surprising considering the role of these  
 24 metals in enzymes such as DMSO reductase that catalyzes similar reactions.[86] As a

1 result, oxidovanadium complexes have generally received much less attention in this  
2 regard. We chose tris(4-fluorophenyl)phosphine as the model compound, due to the ease  
3 of monitoring of the reaction by the very sensitive and quantitative  $^{19}\text{F}$  NMR.[55] Quite  
4 unexpectedly, however, the complexes were found to not have any significant OAT  
5 activity, even at moderate catalyst loadings of 10 mol-% (ESI figure S58).

## 6 Conclusions

7 Mononuclear complexes  $[\text{MO}_2(\text{H}_2\text{L}^{\text{Saltris}})]$  ( $\text{M} = \text{Mo}$  (**1**),  $\text{W}$  (**2**)), as well as a dinuclear  
8 complex  $[\text{VO}(\text{HL}^{\text{Saltris}})]_2$  (**3**) with a hydroxyl-rich Schiff base proligand *N*-(1,3-dihydroxy-  
9 2-(hydroxymethyl)propan-2-yl)-3,5-di-*tert*-butylsalicylaldehyde ( $\text{H}_4\text{L}^{\text{Saltris}}$ ), were pre-  
10 pared by the reaction with Mo, W and V metal precursors in methanol solutions. The  
11 dinuclear vanadium complex shows moderate activity in the oxidation of 4-TBC and 3,5-  
12 DTBC, mimicking the action of the dicopper enzyme catechol oxidase. Strikingly, the  
13 activity of **3** in catechol oxidation was observed to be similar to that of a structurally very  
14 similar dicobalt complex  $[\text{Co}(\text{H}_2\text{L}^{\text{Saltris}})(\text{OAc})]_2$ , whose mechanism of catalysis has been  
15 established. However, a set of experiments investigating the catechol oxidation  
16 mechanism by **3**, prompted by similarity to  $[\text{Co}(\text{H}_2\text{L}^{\text{Saltris}})(\text{OAc})]_2$ , strongly hint that **3** is  
17 not the catalytically active species in the solution. Rather, the presented mechanistic  
18 studies reinforce a strong argument in the literature which suggests that a catecholato-  
19 bearing vanadium species is formed *in-situ* during catalytic turnover conditions in the  
20 presence of 3,5-DTBC, is in fact the real active catalyst. It is further proposed, that such a  
21 complex is formed in virtually every scenario involving vanadium-based precatalysts and  
22 3,5-DTBC, a fact clearly demonstrated by the  $[\text{VO}(\text{HL}^{\text{Saltris}})]_2$  system presented herein.  
23 Furthermore, and quite unexpectedly, the complexes were revealed not to demonstrate  
24 any significant OAT activity in the oxidation of  $(\text{P}(p\text{-C}_6\text{H}_4\text{F})_3)_3$ , even when moderate  
25 catalyst loadings were employed.

## 26 Conflicts of interest

27 The authors declare no competing interests.

## 28 Electronic Supplementary Information

29 All spectra relevant to the characterization of the compounds are presented in the ESI,  
30 as well as detailed descriptions of the syntheses.

## 31 Acknowledgements

32 Dr. Milla Suominen is greatly acknowledged for assisting in electrochemistry  
33 measurements. Dr. Isabella Norrbo, Hannah Byron and Sami Vuori are accredited for  
34 performing the powder XRD measurements. University of Jyväskylä is kindly thanked for  
35 the access to their XRD facilities. AP gratefully acknowledges the Academy of Finland for  
36 funding (grant no. 315911). PS greatly acknowledges the University of Turku Graduate  
37 School (UTUGS) Doctoral Programme in Physical and Chemical Sciences (PCS) for  
38 funding.

## 1 **References**

- 2 [1] C.L. Hill, I.A. Weinstock, On the trail of dioxygen activation, *Nature*. 388 (1997)  
3 332–333. doi:10.1038/40986.
- 4 [2] J. Ibers, R. Holm, Modeling coordination sites in metallobiomolecules, *Science* (80-  
5 .). 209 (1980) 223–235. doi:10.1126/science.7384796.
- 6 [3] K. Karlin, Metalloenzymes, structural motifs, and inorganic models, *Science* (80-  
7 ). 261 (1993) 701–708. doi:10.1126/science.7688141.
- 8 [4] R. Hille, The molybdenum oxotransferases and related enzymes, *Dalt. Trans.* 42  
9 (2013) 3029–3042. doi:10.1039/c2dt32376a.
- 10 [5] R. Hille, T. Nishino, F. Bittner, Molybdenum enzymes in higher organisms, *Coord.*  
11 *Chem. Rev.* 255 (2011) 1179–1205. doi:10.1016/j.ccr.2010.11.034.
- 12 [6] R. Hille, The Mononuclear Molybdenum Enzymes, *Chem. Rev.* 96 (1996) 2757–  
13 2816. doi:10.1021/cr950061t.
- 14 [7] C. Schulzke, Molybdenum and Tungsten Oxidoreductase Models, in: *Bioinspired*  
15 *Catal.*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany, 2014: pp. 349–  
16 382. doi:10.1002/9783527664160.ch13.
- 17 [8] D.C. Crans, J.J. Smee, E. Gaidamauskas, L. Yang, The chemistry and biochemistry of  
18 vanadium and the biological activities exerted by vanadium compounds, *Chem.*  
19 *Rev.* 104 (2004) 849–902. doi:10.1021/cr020607t.
- 20 [9] M. Kirihaara, Aerobic oxidation of organic compounds catalyzed by vanadium  
21 compounds, *Coord. Chem. Rev.* 255 (2011) 2281–2302.  
22 doi:10.1016/j.ccr.2011.04.001.
- 23 [10] C. Bolm, Vanadium-catalyzed asymmetric oxidations, *Coord. Chem. Rev.* 237  
24 (2003) 245–256. doi:10.1016/S0010-8545(02)00249-7.
- 25 [11] R.R. Langeslay, D.M. Kaphan, C.L. Marshall, P.C. Stair, A.P. Sattelberger, M.  
26 Delferro, Catalytic Applications of Vanadium: A Mechanistic Perspective, *Chem.*  
27 *Rev.* 119 (2019) 2128–2191. doi:10.1021/acs.chemrev.8b00245.
- 28 [12] D. Rehder, J. Costa Pessoa, C.F.G.C. Geraldes, M.M. Castro, T. Kabanos, T. Kiss, B.  
29 Meier, G. Micera, L. Pettersson, M. Rangel, A. Salifoglou, I. Turel, D. Wang, In vitro  
30 study of the insulin-mimetic behaviour of vanadium(IV, V) coordination  
31 compounds, *JBIC J. Biol. Inorg. Chem.* 7 (2002) 384–396. doi:10.1007/s00775-  
32 001-0311-5.
- 33 [13] D. Rehder, The role of vanadium in biology, *Metallomics*. 7 (2015) 730–742.  
34 doi:10.1039/C4MT00304G.
- 35 [14] D. Rehder, The potentiality of vanadium in medicinal applications, *Future Med.*  
36 *Chem.* 4 (2012) 1823–1837. doi:10.4155/fmc.12.103.
- 37 [15] T. Klabunde, C. Eicken, J.C. Sacchettini, B. Krebs, Crystal structure of a plant  
38 catechol oxidase containing a dicopper center, *Nat. Struct. Biol.* 5 (1998) 1084–  
39 1090. doi:10.1038/4193.

- 1 [16] C. Gerdemann, C. Eicken, B. Krebs, The crystal structure of catechol oxidase: New  
2 insight into the function of type-3 copper proteins, *Acc. Chem. Res.* 35 (2002)  
3 183–191. doi:10.1021/ar990019a.
- 4 [17] E. Solem, F. Tuczek, H. Decker, Tyrosinase versus catechol oxidase: One  
5 asparagine makes the difference, *Angew. Chemie Int. Ed.* 55 (2016) 2884–2888.  
6 doi:10.1002/anie.201508534.
- 7 [18] A.A. El-Taras, I.M. EL-Mehasseb, A.E.-M.M. Ramadan, Synthesis, characterization,  
8 magnetic, thermal and electrochemical studies of oxidovanadium (IV) picolyl  
9 hydrazones as functional catechol oxidase models, *Comptes Rendus Chim.* 15  
10 (2012) 298–310. doi:10.1016/j.crci.2011.11.009.
- 11 [19] M.R. Maurya, B. Uprety, F. Avecilla, P. Adão, J. Costa Pessoa, Vanadium(V)  
12 complexes of a tripodal ligand, their characterisation and biological implications,  
13 *Dalt. Trans.* 44 (2015) 17736–17755. doi:10.1039/C5DT02716K.
- 14 [20] S.K. Mal, M. Mitra, H.R. Yadav, C.S. Purohit, A.R. Choudhury, R. Ghosh, Synthesis,  
15 crystal structure and catecholase activity of a vanadium(V) Schiff base complex,  
16 *Polyhedron.* 111 (2016) 118–122. doi:10.1016/j.poly.2016.03.033.
- 17 [21] P. Salonen, A. Peuronen, A. Lehtonen, Oxidovanadium(V) amine bisphenolates as  
18 epoxidation, sulfoxidation and catechol oxidation catalysts, *Inorg. Chem.*  
19 *Commun.* 86 (2017) 165–167. doi:10.1016/j.inoche.2017.10.017.
- 20 [22] M.R. Maurya, B. Uprety, F. Avecilla, P. Adão, M.L. Kuznetsov, J. Costa Pessoa,  
21 Solution behaviour and catalytic potential towards oxidation of dopamine by  
22 oxidovanadium(V) complexes of tripodal tetradentate ligands, *Eur. J. Inorg. Chem.*  
23 2017 (2017) 3087–3099. doi:10.1002/ejic.201700342.
- 24 [23] C. Balakrishnan, M.A. Neelakantan, Crystal structure and bio-catalytic potential of  
25 oxovanadium(IV) Schiff base complexes derived from 2-hydroxy-4-(prop-2-yn-1-  
26 yloxy)benzaldehyde and alicyclic/aromatic diamines, *Inorganica Chim. Acta.* 469  
27 (2018) 503–514. doi:10.1016/j.ica.2017.09.060.
- 28 [24] M. Rolff, J. Schottenheim, H. Decker, F. Tuczek, Copper–O<sub>2</sub> reactivity of tyrosinase  
29 models towards external monophenolic substrates: molecular mechanism and  
30 comparison with the enzyme, *Chem. Soc. Rev.* 40 (2011) 4077.  
31 doi:10.1039/c0cs00202j.
- 32 [25] I.A. Koval, P. Gamez, C. Belle, K. Selmeczi, J. Reedijk, Synthetic models of the active  
33 site of catechol oxidase: mechanistic studies, *Chem. Soc. Rev.* 35 (2006) 814.  
34 doi:10.1039/b516250p.
- 35 [26] S.K. Dey, A. Mukherjee, Catechol oxidase and phenoxazinone synthase:  
36 Biomimetic functional models and mechanistic studies, *Coord. Chem. Rev.* 310  
37 (2016) 80–115. doi:10.1016/j.ccr.2015.11.002.
- 38 [27] M. Mitra, A.K. Maji, B.K. Ghosh, P. Raghavaiah, J. Ribas, R. Ghosh, Catecholase  
39 activity of a structurally characterized dinuclear iron(III) complex [FeIII<sub>2</sub>(L)<sub>2</sub>]  
40 [H<sub>3</sub>L=N,N'-bis(3-methoxysalicylalimine)-1,3-diaminopropan-2-ol], *Polyhedron.*  
41 67 (2014) 19–26. doi:10.1016/j.poly.2013.08.064.

- 1 [28] S.K. Dey, A. Mukherjee, The synthesis, characterization and catecholase activity of  
2 dinuclear cobalt(II/III) complexes of an O-donor rich Schiff base ligand, *New J.*  
3 *Chem.* 38 (2014) 4985–4995. doi:10.1039/C4NJ00715H.
- 4 [29] M.E. Cass, D.L. Green, R.M. Buchanan, C.G. Pierpont, Orthoquinone complexes of  
5 vanadium and their reactions with molecular oxygen, *J. Am. Chem. Soc.* 105  
6 (1983) 2680–2686. doi:10.1021/ja00347a027.
- 7 [30] C.L. Simpson, C.G. Pierpont, Complexes of vanadium(III) and vanadium(IV)  
8 containing bipyridine and tetrachlorocatecholate ligands. Insights into the  
9 tunicate vanadium(III) coordination environment, *Inorg. Chem.* 31 (1992) 4308–  
10 4313. doi:10.1021/ic00047a018.
- 11 [31] S.R. Cooper, Y.B. Koh, K.N. Raymond, Synthetic, structural, and physical studies of  
12 bis(triethylammonium) tris(catecholato)vanadate(IV), potassium  
13 bis(catecholato)oxovanadate(IV), and potassium tris(catecholato)vanadate (III), *J.*  
14 *Am. Chem. Soc.* 104 (1982) 5092–5102. doi:10.1021/ja00383a016.
- 15 [32] M.E. Cass, N.R. Gordon, C.G. Pierpont, Catecholate and semiquinone complexes of  
16 vanadium. Factors that direct charge distribution in metal-quinone complexes,  
17 *Inorg. Chem.* 25 (1986) 3962–3967. doi:10.1021/ic00242a027.
- 18 [33] C.R. Cornman, G.J. Colpas, J.D. Hoeschele, J. Kampf, V.L. Pecoraro, Implications for  
19 the spectroscopic assignment of vanadium biomolecules: Structural and  
20 spectroscopic characterization of monooxovanadium(V) complexes containing  
21 catecholate and hydroximate based noninnocent ligands, *J. Am. Chem. Soc.* 114  
22 (1992) 9925–9933. doi:10.1021/ja00051a026.
- 23 [34] J.P. Wilshire, L. Leon, P. Bosserman, D.T. Sawyer, Electrochemical and  
24 spectroscopic studies of molybdenum-catechol complexes: Models for  
25 molybdoenzymes and biological transport, in: *Molybdenum Chem. Biol.*  
26 *Significance*, Springer US, Boston, MA, 1980: pp. 327–344. doi:10.1007/978-1-  
27 4615-9149-8\_26.
- 28 [35] H. Weiner, R.G. Finke, An all-inorganic, polyoxometalate-based catechol  
29 dioxygenase that exhibits > 100 000 catalytic turnovers, *J. Am. Chem. Soc.* 121  
30 (1999) 9831–9842. doi:10.1021/ja991503b.
- 31 [36] C.-X. Yin, R.G. Finke, Vanadium-based, extended catalytic lifetime catechol  
32 dioxygenases: Evidence for a common catalyst, *J. Am. Chem. Soc.* 127 (2005)  
33 9003–9013. doi:10.1021/ja051594e.
- 34 [37] C.X. Yin, R.G. Finke, Kinetic and mechanistic studies of vanadium-based, extended  
35 catalytic lifetime catechol dioxygenases, *J. Am. Chem. Soc.* 127 (2005) 13988–  
36 13996. doi:10.1021/ja052998+.
- 37 [38] C.-X. Yin, Y. Sasaki, R.G. Finke, Autoxidation-product-initiated dioxygenases:  
38 Vanadium-based, record catalytic lifetime catechol dioxygenase catalysis, *Inorg.*  
39 *Chem.* 44 (2005) 8521–8530. doi:10.1021/ic050717t.
- 40 [39] N. Zwettler, J.A. Schachner, F. Belaj, N.C. Mösch-Zanetti, Hydrogen bond donor  
41 functionalized dioxido-molybdenum(VI) complexes as robust and highly efficient  
42 precatalysts for alkene epoxidation, *Mol. Catal.* 443 (2017) 209–219.

- 1 doi:10.1016/j.mcat.2017.09.036.
- 2 [40] M.K. Hossain, J.A. Schachner, M. Haukka, N.C. Mösch-Zanetti, E. Nordlander, A.  
3 Lehtonen, Catalytic epoxidation using dioxidomolybdenum(VI) complexes with  
4 tridentate aminoalcohol phenol ligands, *Inorganica Chim. Acta.* 486 (2019) 17–25.  
5 doi:10.1016/j.ica.2018.10.012.
- 6 [41] G.J.J. Chen, J.W. McDonald, W.E. Newton, Synthesis of molybdenum(IV) and  
7 molybdenum(V) complexes using oxo abstraction by phosphines. Mechanistic  
8 implications, *Inorg. Chem.* 15 (1976) 2612–2615. doi:10.1021/ic50165a008.
- 9 [42] F.A. Schröder, J. Scherle, Über die reaktionen von MoO<sub>3</sub> und WO<sub>3</sub> mit  
10 mehrwertigen alkoholen, *Zeitschrift Für Naturforsch. B.* 28 (1973) 46–55.  
11 doi:10.1515/znb-1973-1-213.
- 12 [43] G.R. Fulmer, A.J.M. Miller, N.H. Sherden, H.E. Gottlieb, A. Nudelman, B.M. Stoltz, J.E.  
13 Bercaw, K.I. Goldberg, NMR Chemical Shifts of Trace Impurities: Common  
14 Laboratory Solvents, Organics, and Gases in Deuterated Solvents Relevant to the  
15 Organometallic Chemist, *Organometallics.* 29 (2010) 2176–2179.  
16 doi:10.1021/om100106e.
- 17 [44] R.K. Harris, E.D. Becker, S.M. Cabral de Menezes, R. Goodfellow, P. Granger, NMR  
18 nomenclature: nuclear spin properties and conventions for chemical shifts. IUPAC  
19 Recommendations 2001. International Union of Pure and Applied Chemistry.  
20 Physical Chemistry Division. Commission on Molecular Structure and  
21 Spectroscopy, *Magn. Reson. Chem.* 40 (2002) 489–505. doi:10.1002/mrc.1042.
- 22 [45] A. Lehtonen, M. Wasberg, R. Sillanpää, Dioxomolybdenum(VI) and -tungsten(VI)  
23 complexes with tetradentate aminobis(phenol)s, *Polyhedron.* 25 (2006) 767–775.  
24 doi:10.1016/j.poly.2005.07.037.
- 25 [46] Y.-L. Wong, L.H. Tong, J.R. Dilworth, D.K.P. Ng, H.K. Lee, New dioxo-  
26 molybdenum(vi) and -tungsten(vi) complexes with N-capped tripodal N<sub>2</sub>O<sub>2</sub>  
27 tetradentate ligands: Synthesis, structures and catalytic activities towards olefin  
28 epoxidation, *Dalt. Trans.* 39 (2010) 4602–4611. doi:10.1039/b926864b.
- 29 [47] A. Peuronen, R. Sillanpää, A. Lehtonen, The syntheses and vibrational spectra of  
30 16O- and 18O-enriched cis-MO<sub>2</sub> (M=Mo, W) complexes, *ChemistrySelect.* 3  
31 (2018) 3814–3818. doi:10.1002/slct.201800671.
- 32 [48] W.P. Griffith, Transition metal oxo complexes, *Coord. Chem. Rev.* 5 (1970) 459–  
33 517. doi:10.1016/S0010-8545(00)80101-0.
- 34 [49] G. Asgedom, A. Sreedhara, J. Kivikoski, J. Valkonen, E. Kolehmainen, C.P. Rao,  
35 Alkoxo bound monooxo- and dioxovanadium(V) complexes: Synthesis,  
36 characterization, X-ray crystal structures, and solution reactivity studies, *Inorg.*  
37 *Chem.* 35 (1996) 5674–5683. doi:10.1021/ic960061r.
- 38 [50] M.K. Hossain, M. Haukka, G.C. Lisensky, A. Lehtonen, E. Nordlander,  
39 Oxovanadium(V) complexes with tripodal bisphenolate and monophenolate  
40 ligands: Syntheses, structures and catalytic activities, *Inorganica Chim. Acta.* 487  
41 (2019) 112–119. doi:10.1016/j.ica.2018.11.049.

- 1 [51] CrysAlisPRO, Rigaku Oxford Diffraction, (2015).
- 2 [52] G.M. Sheldrick, A short history of SHELX, *Acta Crystallogr. Sect. A Found.*  
3 *Crystallogr.* 64 (2008) 112–122. doi:10.1107/S0108767307043930.
- 4 [53] G.M. Sheldrick, Crystal structure refinement with SHELXL, *Acta Crystallogr. Sect. C*  
5 *Struct. Chem.* 71 (2015) 3–8. doi:10.1107/S2053229614024218.
- 6 [54] R.R. Gagne, C.A. Koval, G.C. Lisensky, Ferrocene as an Internal Standard for  
7 Electrochemical Measurements, *Inorg. Chem.* 19 (1980) 2854–2855.  
8 doi:10.1021/ic50211a080.
- 9 [55] K. Most, J. Hoßbach, D. Vidović, J. Magull, N.C. Mösch-Zanetti, Oxygen-Transfer  
10 Reactions of Molybdenum- and Tungstendioxo Complexes Containing  $\eta^2$  -  
11 Pyrazolate Ligands, *Adv. Synth. Catal.* 347 (2005) 463–472.  
12 doi:10.1002/adsc.200404265.
- 13 [56] L.M. Rzepecki, J.H. Waite, A chromogenic assay for catecholoxidases based on the  
14 addition of L-proline to quinones, *Anal. Biochem.* 179 (1989) 375–381.  
15 doi:10.1016/0003-2697(89)90148-6.
- 16 [57] W. Flaig, T. Ploetz, A. Küllmer, Über ultravioletspektren einiger benzochinone,  
17 *Zeitschrift Für Naturforsch. B.* 10 (1955) 668–676. doi:10.1515/znb-1955-1202.
- 18 [58] Y. Sui, X. Zeng, X. Fang, X. Fu, Y. Xiao, L. Chen, M. Li, S. Cheng, Syntheses, structure,  
19 redox and catalytic epoxidation properties of dioxomolybdenum(VI) complexes  
20 with Schiff base ligands derived from tris(hydroxymethyl)amino methane, *J. Mol.*  
21 *Catal. A Chem.* 270 (2007) 61–67. doi:10.1016/j.molcata.2007.01.032.
- 22 [59] G. Asgedom, A. Sreedhara, C.P. Rao, Oxovanadium(V) schiff base complexes of  
23 trishydroxymethylaminomethane with salicylaldehyde and its derivatives:  
24 Synthesis, characterization and redox reactivity, *Polyhedron.* 14 (1995) 1873–  
25 1879. doi:10.1016/0277-5387(94)00461-M.
- 26 [60] V.S. Sergienko, V.L. Abramenko, A. V. Churakov, Y.N. Mikhailov, M.D. Surazhskaya,  
27 Synthesis and crystal and molecular structure of the  $[\text{MoO}_2(\text{L})(\text{H}_2\text{O})] \cdot \text{H}_2\text{O}$   
28 complex (L 2- is the anion of 2-[N-(hydroxynaphtylidene)amino]propan-1,2,3-  
29 triol), *Crystallogr. Reports.* 59 (2014) 523–526.  
30 doi:10.1134/S106377451404018X.
- 31 [61] G.A. Kolawole, K.S. Patel, The stereochemistry of oxovanadium(IV) complexes  
32 derived from salicylaldehyde and polymethylenediamines, *J. Chem. Soc. Dalt.*  
33 *Trans.* (1981) 1241–1245. doi:10.1039/dt9810001241.
- 34 [62] L.J. Willis, T.M. Loehr, K.F. Miller, A.E. Bruce, E.I. Stiefel, Raman and infrared  
35 spectroscopic studies of dioxomolybdenum(VI) complexes with cysteamine  
36 chelates, *Inorg. Chem.* 25 (1986) 4289–4293. doi:10.1021/ic00243a045.
- 37 [63] V.I. Minkin, A. V. Tsukanov, A.D. Dubonosov, V.A. Bren, Tautomeric Schiff bases:  
38 Iono-, solvato-, thermo- and photochromism, *J. Mol. Struct.* 998 (2011) 179–191.  
39 doi:10.1016/j.molstruc.2011.05.029.
- 40 [64] Z. Rozwadowski, E. Majewski, T. Dziembowska, P. Erik Hansen, Deuterium isotope



- 1 effects on  $^{13}\text{C}$  chemical shifts of intramolecularly hydrogen-bonded Schiff bases,  
2 *J. Chem. Soc. Perkin Trans. 2.* (1999) 2809–2817. doi:10.1039/a903200b.
- 3 [65] R.M. Claramunt, C. López, M.D. Santa María, D. Sanz, J. Elguero, The use of NMR  
4 spectroscopy to study tautomerism, *Prog. Nucl. Magn. Reson. Spectrosc.* 49  
5 (2006) 169–206. doi:10.1016/j.pnmrs.2006.07.001.
- 6 [66] C.P. Rao, A. Sreedhara, P.V. Rao, M.B. Verghese, K. Rissanen, E. Kolehmainen, N.K.  
7 Lokanath, M.A. Sridhar, J.S. Prasad, Syntheses, structure, reactivity and species  
8 recognition studies of oxo-vanadium(V) and -molybdenum(VI) complexes, *J.*  
9 *Chem. Soc. Dalt. Trans.* (1998) 2383–2394. doi:10.1039/a801226a.
- 10 [67] V.L. Abramenko, A. V Churakov, V.S. Sergienko, Y.N. Mikhailov, M.D. Surazhskaya,  
11 Inner complex compounds of dioxomolybdenum(VI) with o-oxyazomethines,  
12 derivatives of substituted salicylaldehydes and  
13 tris(hydroxymethyl)aminomethane. crystal structures of two complexes  
14  $[\text{MoO}(\text{L})] \cdot \text{CH}_3\text{OH}$ ; L = Z-substituted salicylalimines, Z = 3-NO<sub>2</sub> and 3, *Russ. J.*  
15 *Coord. Chem.* 39 (2013) 553–559. doi:10.1134/S1070328413080010.
- 16 [68] D. Rehder, C. Weidemann, A. Duch, W. Priebisch, Vanadium-51 shielding in  
17 vanadium(V) complexes: A reference scale for vanadium binding sites in  
18 biomolecules, *Inorg. Chem.* 27 (1988) 584–587. doi:10.1021/ic00276a029.
- 19 [69] C.R. Groom, I.J. Bruno, M.P. Lightfoot, S.C. Ward, The Cambridge Structural  
20 Database, *Acta Crystallogr. Sect. B Struct. Sci. Cryst. Eng. Mater.* 72 (2016) 171–  
21 179. doi:10.1107/S2052520616003954.
- 22 [70] Y.-J. Jang, S.-J. Mo, B.-K. Koo, Molybdenum(VI)- and tungsten(VI)- dioxo complexes  
23 with Schiff base ligands, *Bull. Korean. Chem Soc.* 19 (1998) 587–590.
- 24 [71] U. Sandbhor, S. Padhye, E. Sinn, Cis di-oxomolybdenum(VI) complexes with a  
25 tridentate ONO donor ligand; synthesis, X-ray crystal structure, spectroscopic  
26 properties and oxotransfer chemistry, *Transit. Met. Chem.* 27 (2002) 681–685.  
27 doi:10.1023/A:1019839308548.
- 28 [72] D.F. Back, C.R. Kopp, G. Manzoni de Oliveira, P.C. Piquini, New oxidovanadium(V)  
29 complexes of the cation  $[\text{VO}]^{3+}$ : Synthesis, structural characterization and DFT  
30 studies, *Polyhedron.* 36 (2012) 21–29. doi:10.1016/j.poly.2012.01.015.
- 31 [73] G.S. Pawley, Unit-cell refinement from powder diffraction scans, *J. Appl.*  
32 *Crystallogr.* 14 (1981) 357–361. doi:10.1107/S0021889881009618.
- 33 [74] T. Weyhermüller, T.K. Paine, E. Bothe, E. Bill, P. Chaudhuri, Complexes of an  
34 aminebis(phenolate) [O,N,O] donor ligand and EPR studies of isoelectronic,  
35 isostructural Cr(III) and Mn(IV) complexes, *Inorganica Chim. Acta.* 337 (2002)  
36 344–356. doi:10.1016/S0020-1693(02)01000-9.
- 37 [75] X.-Q. Zhu, C.-H. Wang, H. Liang, Scales of Oxidation Potentials, pK<sub>a</sub>, and BDE of  
38 Various Hydroquinones and Catechols in DMSO, *J. Org. Chem.* 75 (2010) 7240–  
39 7257. doi:10.1021/jo101455m.
- 40 [76] M.J. Sever, J.J. Wilker, Visible absorption spectra of metal–catecholate and metal–  
41 tironate complexes, *Dalt. Trans.* (2004) 1061–1072. doi:10.1039/B315811J.

- 1 [77] S. Mukherjee, T. Weyhermüller, E. Bothe, K. Wiegardt, P. Chaudhuri, Dinuclear  
2 and mononuclear manganese(IV)-radical complexes and their catalytic  
3 catecholase activity, *Dalt. Trans.* (2004) 3842–3853. doi:10.1039/B410842F.
- 4 [78] I.A. Koval, K. Selmeczi, C. Belle, C. Philouze, E. Saint-Aman, I. Gautier-Luneau, A.M.  
5 Schuitema, M. van Vliet, P. Gamez, O. Roubeau, M. Lüken, B. Krebs, M. Lutz, A.L.  
6 Spek, J.-L. Pierre, J. Reedijk, Catecholase activity of a copper(II) complex with a  
7 macrocyclic ligand: Unraveling catalytic mechanisms, *Chem. - A Eur. J.* 12 (2006)  
8 6138–6150. doi:10.1002/chem.200501600.
- 9 [79] J. Ackermann, F. Meyer, E. Kaifer, H. Pritzkow, Tuning the activity of catechol  
10 oxidase model complexes by geometric changes of the dicopper core, *Chem. - A*  
11 *Eur. J.* 8 (2002) 247–258. doi:10.1002/1521-3765(20020104)8:1<247::AID-  
12 CHEM247>3.0.CO;2-P.
- 13 [80] J. Reim, B. Krebs, Synthesis, structure and catecholase activity study of dinuclear  
14 copper(II) complexes, *J. Chem. Soc. Dalt. Trans.* 0 (1997) 3793–3804.  
15 doi:10.1039/a704245k.
- 16 [81] I.A. Koval, C. Belle, K. Selmeczi, C. Philouze, E. Saint-Aman, A.M. Schuitema, P.  
17 Gamez, J.-L. Pierre, J. Reedijk, Catecholase activity of a  $\mu$ -hydroxodicopper(II)  
18 macrocyclic complex: structures, intermediates and reaction mechanism, *JBIC J.*  
19 *Biol. Inorg. Chem.* 10 (2005) 739–750. doi:10.1007/s00775-005-0016-2.
- 20 [82] H. Arzoumanian, Molybdenum-oxo chemistry in various aspects of oxygen atom  
21 transfer processes, *Coord. Chem. Rev.* 178–180 (1998) 191–202.  
22 doi:10.1016/S0010-8545(98)00056-3.
- 23 [83] M.R. Maurya, B. Mengesha, B. Uprety, N. Jangra, R. Tomar, F. Avecilla, Oxygen atom  
24 transfer between DMSO and benzoin catalyzed by cis -dioxidomolybdenum(VI)  
25 complexes of tetradentate Mannich bases, *New J. Chem.* 42 (2018) 6225–6235.  
26 doi:10.1039/C7NJ03551A.
- 27 [84] C.J. Whiteoak, G.J.P. Britovsek, V.C. Gibson, A.J.P. White, Electronic effects in oxo  
28 transfer reactions catalysed by salan molybdenum(VI) cis-dioxo complexes, *Dalt.*  
29 *Trans.* 2 (2009) 2337. doi:10.1039/b820754b.
- 30 [85] M.K. Hossain, J.A. Schachner, M. Haukka, A. Lehtonen, N.C. Mösch-Zanetti, E.  
31 Nordlander, Dioxidomolybdenum(VI) and -tungsten(VI) complexes with tripodal  
32 amino bisphenolate ligands as epoxidation and oxo-transfer catalysts,  
33 *Polyhedron.* 134 (2017) 275–281. doi:10.1016/j.poly.2017.04.036.
- 34 [86] A. Majumdar, S. Sarkar, Bioinorganic chemistry of molybdenum and tungsten  
35 enzymes: A structural–functional modeling approach, *Coord. Chem. Rev.* 255  
36 (2011) 1039–1054. doi:10.1016/j.ccr.2010.11.027.