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Determination of mercury at picogram level in natural waters with inductively coupled plasma mass spectrometry by using 3D printed metal scavengers

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1 Determination of mercury at picogram level in natural waters with inductively coupled

2 plasma mass spectrometry by using 3D printed metal scavengers

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6 Abstract

7 The determination of ultra-trace concentrations of Hg in natural water samples via 8 preconcentration using 3D printed metal scavenger technique followed by inductively 9 coupled plasma mass spectrometry (ICP-MS) was developed. The determination of Hg in 10 certified reference material ERM-CA615 (groundwater) was performed with high accuracy and precision resulting in recovery of $100 \pm 3\%$ and RSD <2.5%, respectively. Selective laser 11 12 sintering (SLS) 3D printing was used to fabricate the scavengers using a mixture of 13 polyamide-12 powder with thiol-functionalized silica. The preconcentration procedure is 14 based on the adsorption of the Hg on the scavenger and followed by elution of the 15 preconcentrated Hg from the filter with 0.3% thiourea in 8% HCl prior to its determination by 16 ICP-MS. A preconcentration factor of 92.8 can be achieved by filtering 495 mL of water 17 followed with the elution step. Very low instrumental detection limit and method detection limits were obtained resulting in 0.013 and 0.037 ng L⁻¹, respectively. The method was 18 19 applied successfully for the determination of Hg in different lake and river water samples. 20 The developed method is the first preconcentration method enabling simple and accurate determination of Hg in pg L^{-1} concentrations in natural waters with ICP-MS. 21 22 Keywords: Mercury; Inductively coupled plasma mass spectrometry; Ultra-trace concentration; Preconcentration; 3D printing; Natural water. 23

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1 1. Introduction

2 Mercury is one of the most toxic elements due to its accumulative and persistent nature in the 3 environment and living organisms. It occurs naturally in the environment in a variety of 4 chemical and physical forms at very low concentrations [1]. In pristine natural waters, Hg concentrations are typically between pg and ng L^{-1} range. Recent reports estimate a total 5 mercury concentration in natural waters ranging from 0.03 to 90 ng L⁻¹, while contaminated 6 waters may contain up to several μ g L⁻¹ of Hg [1,2]. Although mercury is not an abundant 7 8 element in nature, its presence in polluted waters and the uptake of Hg by aquatic media has 9 become an area of great environmental concern and a potential risk to human health [3]. All forms of mercury are poisonous and its toxicity is generally increased by transformation into 10 11 organic forms via biomethylation [1]. Monitoring of mercury traces in the hydrosphere has 12 been regulated in many countries, e.g., in Europe by the European Water Framework Directive [2,4]. Hg is included in Annex I of the Directive 2013/39/EU as priority hazardous 13 substance and a maximum allowable concentration of Hg and its compounds of 0.07 μ g L⁻¹ in 14 15 inland and other surface waters has been established [5,6]. In Finland, heavy metal 16 concentrations in natural waters are usually at a very low level. For this reason, the method 17 detection limits for Ni, Cd, Pb, and Hg are stricter than those presented in the Water Framework Directive. For Hg, the recommendation for method detection limit is 5 ng L^{-1} [7]. 18 19 Accurate quantification of Hg traces in natural waters is challenging due to very low Hg 20 concentrations. Highly sensitive detection techniques are available for mercury e.g. cold 21 vapor atomic fluorescence spectrometry (CV-AFS) and inductively coupled plasma mass 22 spectrometry (ICP-MS) but most modern analytical techniques do not succeed in direct determination of low Hg concentrations [1,2,8]. Thus, a preconcentration step is needed to 23 24 reach the detection limits required for the environmental monitoring of Hg [9]. The analytical 25 performance of non-chromatographic methods using different types of preconcentration

methods has been reviewed previously [1,10,11]. Most of these methods are using a tedious
cold vapor generation coupled to atomic spectroscopic techniques with detection limits
between 0.05 and 500 ng L⁻¹ for Hg²⁺. Still, many of these approaches have inadequate
detection limit for ultra-trace Hg analysis in environmental samples and the development of
simple and sensitive methods are highly desirable.
During the preconcentration step, the analyte is usually also separated from most of the

7 matrix elements, in order to eliminate possible interferences and lowering the detection limit 8 [12]. Various approaches have been applied for separation and preconcentration of Hg based 9 mainly on liquid-liquid extraction [13–15], co-precipitation [16,17] and solid-phase extraction (SPE) [3,6,18–20]. SPE is the most commonly used preconcentration method for 10 11 trace metals due to its simplicity, flexibility, and ability to achieve high enrichment factors. 12 Extraction efficiency and selectivity are greatly dependent on the sorbent material used and therefore the choice of the appropriate adsorbent with high affinity and selectivity for Hg is 13 14 the most critical step in SPE method development [12]. Numerous materials, such as 15 chelating resins [21], ion exchangers [22–24], modified carbon nanotubes [25,26] and 16 nanoparticles [2,3], modified silica gel [27] and polyurethane foam [28,29] have been 17 suggested as SPE sorbents. There are also some commercially available metal scavengers for Hg but their disadvantage is that they are usually sold as powdery materials and hence 18 19 centrifugation or filtration is needed to recover the used adsorbent or to purify the sample 20 solutions from remaining particles. These difficulties make the use of the adsorbent laborious 21 and challenging. Moreover, the use of solid samplers instead of small particle material would 22 enable in situ sampling which would be almost impossible with a material consisting of small 23 particles. These challenges can be avoided by anchoring the chemically active component to a sorbent matrix by using 3D printing. 24

1	Over the past few years, 3D printing has received increasing attention as a way to fabricate
2	macroscopic objects with actual functionality instead of just mechanical or aesthetical
3	properties [30–35]. 3D printing has found many potential applications in the fields of
4	separation sciences [36] and analytical chemistry [37]. However, many of these reports have
5	been focusing on either extrusion-based methods or stereolithography whereas methods such
6	as Selective Laser Sintering (SLS) have received less attention. This is even though the SLS
7	3D printing possesses some obvious advantages if targeting, for example, any kind of flow-
8	related processes. The challenge with the aforementioned extrusion-based and
9	stereolithography methods is that they produce objects, where the surfaces of the printed
10	objects are completely solid and the reactive surface area inside the object is not accessible.
11	The SLS 3D printing doesn't suffer from similar challenges as it uses a laser to partially
12	sinter small polymer beads together, leaving accessible voids between the polymer particles.
13	These voids are crucial for analytical chemistry purposes, as they allow the fluid to flow
14	through the pores of the objects and therefore allow for controlled chemical interaction
15	between the fluid and the object. This level of sintering, and therefore the porosity, can be
16	controlled by adjusting the printing parameters such as laser power, laser speed and
17	temperature [38–41]. Even different areas of the same object can be made to possess different
18	porosities. For example, the outer walls of the object can be made impermeable for fluids
19	while leaving the inside of the object highly macroporous [42]. The chemical functionality
20	can be embedded into the printed objects either by simply mixing the additive into the
21	starting material or by post-processing the printed objects. Almost anything can be used as
22	the additive since as long as the additive can withstand the temperature of the printing
23	process, it can be printed to form chemically functionalized macroscopic objects without
24	losing the activity of the additive during the process [42,43]. Turning powdery materials into

1 easily handleable objects with customizable shape, size and porosity often make their use in 2 actual applications much more feasible if compared to using them in their powdery form. 3 This paper presents a 3D printed Hg scavenger (3D-Thiol) method for the preconcentration 4 and determination of ultra-trace mercury concentrations in natural waters. A series of 5 sorption experiments were conducted using flow-through procedures to study the effect of 6 sorption, elution, and selectivity for mercury and some other metals. In addition, adsorption 7 isotherms for thiol functionalized silica (Thiol) and 3D-Thiol were determined. Finally, an 8 analytical method for Hg determination in ultra-trace concentrations was developed, based on 9 sorption of Hg on 3D-Thiol and elution using HCl and thiourea solution followed by ICP-MS detection. To our knowledge, this is the very first study enabling Hg detection with ICP-MS 10 for pg L^{-1} concentrations without the utilization of cold vapor technique. In addition, this is 11 12 the first instance where thiol functionalized material, use-case and functionality have been combined with SLS 3D printing. 13

14 **2. Experimental Section**

15 **2.1. Reagents**

16 All reagents were of analytical grade and all aqueous solutions were prepared in high purity

- 17 water (18.2 M Ω cm) produced by Elga Purelab Ultra water purification system
- 18 (Buckinghamshire, U.K.). Single-element standard of Hg as Hg^{2+} (10 mg L⁻¹, PurePlus), Cd,
- 19 Co, Cr, Fe, Ir, K, Mg, Mn, Na, Ni, and Pb (1000 mg L^{-1} , Pure) were supplied by PerkinElmer.
- 20 1000 mg L⁻¹ stock solution of Ca was prepared from CaCO₃ (Merck, \geq 99.0%). The ICP-MS
- 21 performance was checked daily with a NexION Setup Solution (1 μ g L⁻¹ Be, Ce, Fe, In, Li,
- 22 Mg, Pb, and U in 1% HNO₃, PerkinElmer). Thiourea (99.8%) was obtained from VWR
- 23 Chemicals and SiliaMetS Thiol was purchased from SiliCycle (Quebec City, Canada).
- 24 Polyamide-12 (PA12), thermoplastic polyurethane and polypropylene printing powders were

1	purchased from BASF (Ludwigshafen am Rhein, Germany). Certified reference material
2	(ERM-CA615, groundwater) was produced and certified by the Joint Research Centre,
3	Institute for Reference Materials and Measurements (JRC-IRMM). Ultra-pure hydrochloric
4	acid (34–37%) was purchased from ANALYTIKA, spol. s r.o. (Prague, Czech Republic).
5	Puriss. p.a. hydrochloric acid (\geq 37%) and nitric acid (\geq 65%) were purchased from Merck and
6	were used for wash solutions with ESI prepFAST. All glass containers that could come into
7	contact with samples and standards were filled with 10% (v/v) hydrochloric acid (\geq 37%) for
8	at least 24 h and tested for Hg followed by ICP-MS analysis. Sample bottles were first rinsed
9	thoroughly with ultrapure water and then completely filled with it. A blank sample was
10	prepared by using this water and was then analyzed with ICP-MS. Sample bottles were used
11	if the measured Hg concentration was below the instrument detection limit (IDL). After
12	testing, glass containers were filled with dilute HCl (~0.5%) and were thoroughly rinsed with
13	high purity water before usage. Calibration standards and smaller volume samples were
14	prepared into 50 mL polypropylene centrifuge tubes (VWR International).

15

2.2. Sampling and sample pretreatment

A total of seven different water samples were collected in 2L glass bottles from central 16 17 Finland. Tap water was collected from the research laboratory of the Department of 18 Chemistry (Jyväskylä), groundwater from Heinäahonlähde (Laukaa), Lake water 1 from lake 19 Päijänne (Korpilahti), Lake water 2 from lake Jyväsjärvi (Jyväskylä), River water from river 20 Tourujoki (Jyväskylä), Pond water 1 from Myllylampi (Tikkakoski) and pond water 2 from 21 Mustalampi (Jyväskylä). Samples were filtered (Whatman No. 41) only if they contained 22 solid material. Then they were acidified with ultra-pure HCl to obtain 3% (v/v) acid 23 concentration and stored at 4 °C in a refrigerator overnight if not analyzed immediately after 24 the sample collection.

1 **2.3. Instrumentation**

2 All mercury analyses were performed on a PerkinElmer (Massachusetts, U.S.A.) NexION 3 350D inductively coupled plasma mass spectrometer (ICP-MS) in standard mode. The ICP-4 MS was interfaced with an Elemental Scientific (Nebraska, U.S.A.) prepFAST 4DX 5 sample/standard autodilution system. A NexION Setup Solution was used for the daily 6 optimization procedures for the ICP-MS according to manufacturer's instructions. Data were 7 acquired in counts per second (cps) for Hg (the sum of the counts at m/z 200, 201 and 202), and Ir (m/z 193) was used as the internal standard. The most abundant W isotope ¹⁸⁴W was 8 9 monitored during all measurements, and it was concluded that WO did not interfere with Hg in the studied water samples when the 3D-Thiol method was used. Calibration standards were 10 prepared in the range of 0–500 ng L^{-1} (blank, 50, 250 and 500 ng L^{-1}) Hg and the standards 11 12 and samples were adjusted to contain 3% (v/v) HCl. For analysis of elution solutions containing thiourea and HCl, the standards were matrix-matched with 0.1125% (m/v) 13 thiourea and 3% (v/v) HCl. The system was first rinsed with a mixture of 0.75% (v/v) HNO₃ 14 and 2.25% (v/v) HCl and then with 3% (v/v) hydrochloric acid between samples. The 15 16 optimum operating conditions and measurement parameters for ICP-MS are listed in Table 1. 17 Elements present in the selectivity study were analyzed on Avio 500 ICP-OES (PerkinElmer, 18 Massachusetts, U.S.A.) or with ICP-MS. More detailed information about the analytical 19 procedure is presented in Tables S1 and S2. Black/white 3.18 mm i.d. PVC peristaltic pump 20 tubing (PerkinElmer, Massachusetts, U.S.A.) was used for the preconcentration process and 21 the flow rate of the samples was adjusted using an ISCO (Nebraska, U.S.A.) WIZ peristaltic 22 pump. Helium imaging was performed with Carl Zeiss (Oberkochen, Germany) ORION NanoFab helium ion microscope (HIM). The beam energy was set to around 30 kV while the 23 24 beam current varied between 0.211 and 0.234 pA. Scan dwell time of 0.2 µs with a viewing 25 distance of 5.101 to 7.833 millimeters was utilized for the imaging. Flood gun was used

throughout the imaging to counteract the charging event present. Samples did not receive any
 additional pretreatment before the imaging process.

3 **2.4. Designing and preparation of the 3D printed scavengers**

4 The objects were designed using FreeCad v. 0.16 and then sliced with Slic3r v. 1.2.9 into 100 5 µm layers to obtain filters with desired shape and size (5 mm tall disc with 16.5 mm 6 diameter). The starting material for the SLS 3D printing was made by mixing polyamide-12 7 powder (~50 µm diameter) with either 5 or 10 wt% of thiol-functionalized silica (40–63 µm). The material was then mechanically mixed and placed into the 3D printer. The actual printing 8 9 process was done using Sharebot SnowWhite SLS 3D printer with a laser power of 30–40% (of a maximum of 14 W), a laser speed of 1800–2400 mm s⁻¹ and printing temperature of 170 10 11 °C. No post-processing, aside for cleaning the objects of any unsintered powder, was done for the 3D printed objects. 12

13 **2.5. Adsorption Experiments**

14 The developed 3D-Thiol method was tested with model solutions prior to the separation and 15 preconcentration of Hg from real samples. For this purpose, 3D printed filters were placed 16 into a 10 mL syringe (Fig. 1) and washed from any unsintered powder with ultrapure water. 17 Extraction of Hg from aqueous solutions was investigated in a flow-through process. The sorption was performed in centrifuge tubes containing 100 ng L^{-1} of Hg in 40 mL of 3% (v/v) 18 19 HCl. Initial tests were carried out using a peristaltic pump with a maximum flow rate (~8.3 mL min⁻¹). The adsorbed concentration of Hg was determined using ICP-MS. Adsorption 20 21 percentage was calculated using the following equation:

Adsorption% =
$$\frac{C_{A-} C_B}{C_A} \cdot 100\%$$
 (1)

1 where C_A and C_B are initial and final concentration (ng L⁻¹) in the solution, respectively [44].

2 In the elution step, 2–10 mL of 0.3–2% (w/v) thiourea and 8–40% (v/v) hydrochloric acid

3 solution was passed through 3D-Thiol at a flow rate of $\sim 1-8.3$ mL min⁻¹.

4 **2.6. Determination of adsorption capacity**

5 Equilibrium isotherm studies were carried out with different initial concentrations of Hg in 6 flow-through experiments. The adsorption capacity was determined for both Thiol and 3D-7 Thiol. The adsorption efficiency of Thiol was performed by adding 10 mg of beads into 40 mL of different solutions containing different concentrations of Hg (1, 25, 50, 75 and 100 mg 8 9 L^{-1}) in 3% (v/v) HCl. The tube was sealed and then agitated using Stuart SF1 at 500 osc min⁻¹ 10 for 2 h at room temperature. The thiol-functionalized silica was removed from the Hg 11 solution by filtration (Whatman No. 42) and samples were analyzed by using ICP-MS after 12 appropriate dilution.

13 The adsorption capacity for 3D-Thiol was performed with the same sample volume and Hg 14 concentrations as above. To obtain approximately 10 mg of Thiol in the printed scavenger, 15 the weight fraction of the additive was dropped to 5 wt% in the printed scavenger and 3D-Thiol filters were placed into a 5 mL syringe. 40 mL solutions were passed through one filter 16 five times with a peristaltic pump at a maximum flow rate ($\sim 8.3 \text{ mL min}^{-1}$). After adsorption, 17 18 the residual concentration of Hg was analyzed by ICP-MS after appropriate dilution. The 19 concentration of Hg adsorbed per unit mass of the adsorbents was calculated from the 20 following equation:

$$q_e = \frac{(C_0 - C_e)V}{m}$$
(2)

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- 1 where q_e is the adsorption capacity (mg g⁻¹) of the adsorbent at equilibrium; C_0 and C_e are the
- 2 initial and final concentration of Hg (mg L^{-1}), respectively; V is the volume of the
- 3 solution (L) and *m* is the mass of the adsorbent (g).

4 **2.7. Selectivity experiments**

- 5 A variety of 40 mL samples containing spiked lake water or synthetic sample were passed
- 6 through a 10 ml syringe containing one 3D-Thiol filter with a \sim 8.3 mL min⁻¹ flow rate.
- 7 Spiking levels were 5 ng L^{-1} of Hg, 10 mg L^{-1} of Ca, 6 mg L^{-1} of Na, 2 mg L^{-1} of Mg, 1 mg L^{-1}
- 6 of K, 1 mg L⁻¹ of Fe and 50 µg L⁻¹ of Cd, Co, Cr, Mn, Ni, and Pb. Ca, Na, Mg, K, Fe, Cr, and
- 9 Mn were measured with ICP-OES and Co, Ni, Cd, Pb, and Hg were analyzed with ICP-MS.
- 10 More detailed information about the analytical procedure is presented in Table S3.

11 **2.8.** Spiking studies and the treatment of the real sample

Samples were spiked with 20 ng L^{-1} of Hg and prepared in 45 mL volume of which 5 ml was 12 set aside for direct ICP-MS analysis. The real samples were prepared into 500 mL volume 13 14 and 5 mL of the sample was saved for direct measurement with ICP-MS. The sample solution was pushed through the 3D-Thiol with ~ 5 mL min⁻¹ flow rate using a peristaltic pump 15 16 resulting in the time needed for preconcentration to be about 1 h 40 min. The elution step was 17 done without a peristaltic pump in backflush mode. The retained Hg was eluted with 2 mL of 0.3% (w/v) thiourea in 8% (v/v) HCl solution. The syringe plunger was used to gently push 18 19 the first drop of the eluent through 3D-Thiol. Then the rest of the eluent was allowed to drain 20 through the filter. Finally, the 3D-Thiol was washed with 3.335 mL of ultrapure water to 21 obtain 3% (v/v) HCl concentration in the final solution, which was analyzed by ICP-MS. The 22 elution and washing take approximately 5 minutes.

- 23
- 24

1 **3. Results and discussion**

2 **3.1. Analytical performance of ICP-MS analysis**

The method showed good linearity over the calibration range of $0-500 \text{ ng L}^{-1}$ (Fig. S1) and 3 the correlation coefficient of linear regression (r^2) was higher than 0.9999. IDL achieved for 4 Hg determination was 1.2 ng L^{-1} (calculated as three times the standard deviation of a series 5 6 of 10 replicate measurement of the calibration blank). The method detection limit (MDL) 7 was determined according to U.S Environmental Protection Agency method detection limit procedure [45] and it was calculated to be 3.5 ng L^{-1} . The short and long term precision were 8 estimated by measuring a quality control sample containing 20 ng L^{-1} of Hg, 20 mg L^{-1} of C, 9 5 mg L^{-1} of Ca, 1 mg L^{-1} of Fe, 2 mg L^{-1} of K, 2 mg L^{-1} of Mg and 5 mg L^{-1} of Na and 2 mg 10 L^{-1} of S. The precision (relative standard deviation, RSD) within one day was 2.9% (n=7) and 11 12 during four weeks 4.0% (n=23). In addition, the precision and accuracy of the ICP-MS method were estimated using a certified groundwater reference material (ERM-CA615, 13 certified value $37 \pm 4 \text{ ng L}^{-1}$; expanded uncertainty at 95% confidence level). Replicate 14 analyses on two different days gave a mean value of 37.0 ng L^{-1} (n = 5) with a precision of 15 2.5%. 16

17 **3.2.** Selection of the chemically active component and supporting matrix

Several commercially available metal scavengers and ion exchange materials were examined for the preconcentration of Hg. Thiol-functionalized silica proved to be the most suitable scavenger for Hg so it was chosen for a chemically active component for 3D printed scavenger. Resins containing the thiol functional groups have been found to be able to adsorb Hg [46,47]. Unfortunately, many of these are no longer commercially available (Spheron-Thiol, Duolite GT73, Ambersep GT74). Polyamide-12, thermoplastic polyurethane, and polypropylene printing powders were tested for 3D printing supporting matrix. PA12 proved

1 to be a suitable material since it is a readily available, durable and easily printable polymer.

2 In addition, Hg was not retained by the PA12 after elution with acidic thiourea solution.

3 3.3. Macroscopic structure of the 3D printed filter

4 The HIM imaging was used to get information about the macroscopic structure of the printed 5 object as well as the distribution and the attachment of the additive. HIM imaging, instead of 6 the more traditional SEM imaging, was conducted for the printed filters to combat the 7 charging effect often when imaging nonconductive samples [48]. It is also evident, that the 8 silica particles have not been capsulated by the polymer and therefore they remain accessible 9 for chemical reactions (Fig. 2 left). The overall structure of the printed filter can be seen to be 10 highly macroporous, consisting of polymeric beads that have been partially sintered together 11 by the laser during the printing process. Additionally, the distribution of the silica particles 12 throughout the printed object seems to be rather even (Fig. 2 right). However, this was expected as the distribution and the attachment for a similar system has been previously 13 14 reported [42,43] and thus was not studied more in depth here.

15 **3.4. Adsorption and adsorption isotherm studies**

16 Initial adsorption experiments were conducted using a peristaltic pump. During the 17 experiment, up to 97% of the Hg was adsorbed, which shows the rather satisfactory retention kinetics. Complete adsorption could be achieved with a more moderate flow rate (~5 mL 18 min⁻¹). In this case, the Hg concentration in the remaining solution is very close or below the 19 instrument detection limit (1.2 ng L^{-1}) which indicates complete adsorption. Since the Hg 20 concentration in pristine natural waters can be only a few ng L^{-1} , a high preconcentration 21 22 factor is desired. The preconcentration factor can be maximized by increasing the sample 23 volume and decreasing the eluent volume. However, due to practical reasons such as ease of 24 sampling and duration of pretreatment, the volume of real natural water sample should be

1 kept reasonable. Taking these considerations into account, a sample volume of 500 mL was

- 2 chosen, of which 5 mL was set aside for direct ICP-MS analysis.
- 3 The equilibrium conditions of the sorption process can be described using sorption isotherms.
- 4 The most frequently used adsorption isotherm equations [23,49,50] of the Langmuir (3) and
- 5 Freundlich (4) isotherms were applied to utilize the linearized equations indicated below:

$$\frac{C_e}{q_e} = \frac{1}{K_L \cdot q_{max}} + \frac{C_e}{q_{max}}$$
(3)
$$\ln q_e = \ln K_F + \frac{1}{n} \ln C_e$$
(4)

where $q_e (\text{mg g}^{-1})$ is the equilibrium adsorption capacity, $C_e (\text{mg L}^{-1})$ is the Hg concentration 6 at equilibrium, q_{max} (mg g⁻¹) is the monolayer maximum adsorption capacity, K_L (L mg⁻¹) is 7 Langmuir equilibrium constant, K_F (L g⁻¹) is a coefficient related to the amount of adsorbed 8 9 metal ions and *n* is Freundlich adsorption constant related to surface heterogeneity. The 10 Langmuir equation assumes the same energy for all adsorption sites and that monolaver is formed when the solid surface reaches saturation [51]. The plot of Ce/qe against Ce gives a 11 straight line and the values of q_{max} and K_L are obtained from the slope and intercept of the 12 13 plot [23]. The Freundlich isotherm, in turn, is an empirical equation and assumes that 14 adsorption is multilayer and is based on heterogeneous surface adsorption suggesting that binding sites are not equivalent [23,51]. 15

Linear plots of $\ln q_e$ versus $\ln C_e$ revealed that the adsorption obeys well Freundlich isotherm (Fig. S2). The results showed that the Hg adsorption onto Thiol can be considered to be a multilayer adsorption system (Table 2). The loading capacity for Thiol is 264.8 mg g⁻¹ according to the manufacturer. The experimental value obtained from the Langmuir isotherm model for Thiol was 253.5 mg g⁻¹ and for 3D-Thiol 222.4 mg g⁻¹. These results indicate that 3D printing decreases the maximum capacity of the resin only slightly. Hg concentrations in

natural waters are so low that the maximum loading capacity is not easily reached. For
example, Hg concentration of 46 mg L⁻¹ with a sample volume of 500 mL is required to
saturate all absorption sites of 3D-Thiol.

4 **3.5. Selectivity**

5 In order to study the selectivity of 3D-Thiol towards Hg, competitive adsorption of Ca, Cd, 6 Co, Cr, Fe, K, Mg, Mn, Na, Ni, and Pb from spiked lake water and synthetic sample were investigated. In spite of the high concentrations of other metal ions $(0.05-10 \text{ mg L}^{-1})$. Hg was 7 retained to 3D-Thiol completely as the Hg concentration of the sample that passed through 8 9 the filter was below the limit of detection. Moreover, Ca, Na, Mg, K, Cr, and Mn were not 10 retained at all, Fe, Co, Ni, Cd, and Pb showed very low percentage retention values to be \leq 11 1.0% (Table S4). 3D-Thiol proved to be extremely selective for Hg in 3% (v/v) HCl. These results provide further evidence of the great selectivity of 3D-Thiol for Hg. 12

13 **3.6.** The effect of eluent composition and its volume

14 According to initial testing with different eluents and literature review [1,52,53], thiourea in 15 hydrochloric acid solution was found to be the most promising eluent. HCl has also been 16 reported [54] to have an enhanced performance due to its complexing abilities with mercury 17 and therefore it was of interest to investigate the influence of thiourea and HCl concentration on the elution efficiency for 3D-Thiol. Thiourea was found to affect the intensity level of the 18 19 internal standard in ICP-MS analysis, hence, the samples with different eluent compositions 20 were analyzed without internal standard correction (Table 3). To enable the use of internal 21 standard, matrix matching was employed in subsequent analysis.

22 Effect of thiourea and HCl concentration on elution efficiency was studied in duplicates with

- 23 10 mL eluent volumes and ~ 8.3 mL min⁻¹ flow rate without internal standard correction
- 24 (Table 3). Higher thiourea and acid concentration results in complete elution of Hg from the

3D-Thiol, however, it is desirable to keep the acid concentration and total dissolved solids
below 4% and 0.2%, respectively for ICP-MS [55]. Samples have to be diluted before
measurement if the acid concentration is above the tolerable limits and hence the higher
preconcentration factor would be lost. In addition, at least 4 mL sample volume is needed for
measurement. Because of these reasons, 0.3% thiourea in 8% HCl was chosen for further
studies.

7 The effect of eluent volume (2, 5 and 10 mL) and eluent flow rate (\sim 1, 3 and 5 mL min⁻¹)

8 were evaluated using a peristaltic pump with the selected eluent. It was observed, that the

9 recovery was satisfactory (98–110%) with all tested eluent volumes and flow rates. However,

10 these small eluent volumes were not completely recovered when using a peristaltic pump. To

11 obtain quantitative recoveries, the elution step was performed manually without a peristaltic

12 pump as described in section 2.8. Considering the sample volume of 495 mL, final elution

13 volume of 2 mL and a dilution factor of 2.67, the preconcentration factor becomes 92.8. The

14 method detection limit for 3D-Thiol method becomes 0.037 ng L^{-1} when the preconcentration

15 factor is taken into account.

16 **3.7. Reusability of the sorbent**

The reusability of 3D-Thiol as a sorbent was examined by assessing the change in the recoveries of the analyte through several sorption-elution cycles at a maximum flow rate of the peristaltic pump following the procedure described in section 2.5. The results revealed that one 3D-Thiol filter could be reused up to 10 times without a loss of adsorption efficiency (Fig. S3). The recovery and relative standard deviation throughout the reusability test for adsorption was 95.2% and 0.7%, respectively (n=2). The adsorption was satisfactory even when the highest flow rate was used. As stated previously, the extraction efficiency is easily

1 increased with a more moderate flow rate. Nonetheless, in order to avoid any contamination,

2 new 3D-Thiol filters were employed throughout the study.

3 **3.8. Spiking tests**

4 A total of five spiked real samples were analyzed with the 3D-Thiol method as well as 5 directly without any preconcentration. The results obtained using 3D-Thiol method were significantly closer to the spiked concentration of 20 ng L^{-1} compared with the direct 6 7 measurement as can be seen in Table 4. This can be seen more clearly in the case of yellow to 8 brown color waters (Lake water 1, River and Pond water) which contain more humic 9 substances than clearer waters (Tap water and Lake water 1). Some spiking tests were also performed at 10 ng L^{-1} level, for which direct measurement from tap water gave 10.2 ng L^{-1} 10 (RSD 7.2%) and 3D-Thiol method 10.2 ng L^{-1} (RSD 1.7%). It is likely that the challenging 11 12 matrix of the colored waters suppresses the signal of mercury in direct ICP-MS determination and might result in biased low concentrations (Table 4). With the 3D-Thiol method, Hg can 13 14 be separated from most of the matrix and interferences are reduced when compared to direct 15 determination. It was observed that RSD values could slightly be improved by using two 3D-16 filters in the same syringe (Tables 4 and 5). As the amount of the functional group is 17 increased it is more likely that Hg interacts with thiol group and different kind of samples 18 behave in the same manner. Although the RSD values were improved with the use of two 19 3D-Thiol filters, RSD values were considered to be sufficient even when only one filter was 20 used. Obtained results further confirm the usefulness of the proposed method.

21 **3.9.** Analysis of mercury concentration in real samples

The proposed method was applied for the determination of Hg in different water samples.
The samples were analyzed both with the direct determination and with the preconcentration
method. With direct determination, Hg concentration of the water samples was below the

MDL except for one sample (Table 5). This represents the need for a 3D-Thiol method to
enable a reliable analysis of extremely low concentrations of Hg in natural waters. The
reproducibility of the method is very good considering the low ng L⁻¹ concentrations. In
Central Finland, Hg concentrations of natural waters are at a very low level and with the 3DThiol method, the recommended 5 ng L⁻¹ method detection limit was easily reached.

6 **4. Conclusions**

7 A SLS 3D printed 3D-Thiol scavenger for Hg preconcentration was prepared and applied for the determination of Hg from water samples with ICP-MS. The proposed method offers a 8 9 simple and highly selective method for the extraction and determination of ultra-trace Hg. 10 3D-Thiol has an excellent selectivity towards Hg over Ca, Cd, Co, Cr, Fe, K, Mg, Mn, Na, 11 Ni, and Pb. The interfering effects of the matrix are reduced when 3D-Thiol method is applied. In addition, the loading capacity of 222.4 mg g^{-1} for 3D-Thiol is significantly higher 12 than adequate for this application. The captured Hg can be quantitatively eluted from the 3D-13 14 Thiol filter with a mixture of 0.3% (w/v) thiourea and 8% (v/v) HCl solution. If desired, the 15 3D-Thiol scavenger can be reused at least 10 times with no decrease in its extraction efficiency. The results indicate that the new 3D-Thiol is an exceptional candidate for Hg 16 17 sorption from natural waters. Due to the high preconcentration factor of 92.8 and extremely low detection limit of 0.037 ng L^{-1} , ultra-trace amounts of Hg at pg L^{-1} levels in water 18 19 samples can be quantified by the method developed.

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3 Appendix A. Supplementary data

4 Supplementary data to this article can be found online at https://doi.org.

5 Author Contributions

- 6 The manuscript was written through contributions of all authors. All authors have given
- 7 approval to the final version of the manuscript. S.K. did the Hg adsorption and elution
- 8 experiments and carried out the ICP-MS and ICP-OES analysis. E.L. prepared the 3D model
- 9 of the scavenger and performed 3D printing. S.P. assisted in the designing of the experiments
- 10 and A.V. supervised the project.

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1 **Captions to figures**

- 2 Fig. 1. SLS 3D printed 5 mm (diam. 16.5 mm) 3D-Thiol filter (left) and filter placed in a 3 syringe (right).
- 4 Fig. 2. HIM image showing the attachment of the thiol-functionalized silica particle onto the
- 5 3D printed PA12 framework (left). HIM image of the overall macroporous structure of the
- 6 3D printed filter containing 10 wt% of functionalized silica on the PA12 framework. Thiol-
- 7 functionalized silica particles are highlighted for clarity.
- 8

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1 Fig. 1



- 3
- 4 Fig. 2



Table 1. ICP-MS instrumental operating parameters and conditions

System/ Operating parameter	Type/Value/Mode
Sample introduction	ESI prepFAST
Nebulizer	PFA-ST nebulizer for prepFAST
Spray chamber	Baffled cyclonic, glass, cooled to 2 °C
RF power	1600 W
Plasma gas flow rate	18 L min ⁻¹
Auxiliary gas flow rate	1.2 L min ⁻¹
Nebulizer gas flow rate	0.89–0.93 L min ⁻¹
Isotope	The sum of ²⁰⁰ Hg, ²⁰¹ Hg, and ²⁰² Hg
Internal standard	193 Ir at 100 µg L ⁻¹ with 10 ms dwell time
Detector mode	Dual
Sweeps/reading	50
Replicates	5
Dwell time	100 ms
Scan mode	Peak hopping
Curve type	Linear through zero

Table 2. Constants of examined isotherms for Hg adsorption

Isotherm model	Constants	Value for 3D-Thiol (n=1)	The average value for Thiol
			(n=3)
Langmuir	$q_{max} (\mathrm{mg g}^{-1})$	222.4	253.5
	K_L (L mg ⁻¹)	0.160	0.337
	R^2	0.975	0.940
Freundlich	$K_F(\mathrm{L} \mathrm{g}^{-1})$	23.5	54.2
	n	1.53	2.07
	R^2	0.982	0.980
	Jour		

Table 3. Recovery $(\%) \pm$ std. deviation for Hg from 3D-Thiol using different eluent

2 compositions

Eluent	Recovery (%)	Required dilution factor for analysis
0.3% thiourea in 8% HCl	90.1 ± 3.8	2.67
1% thiourea in 8% HCl	99.7 ± 4.9	5
2% thiourea in 10% HCl	98.8 ± 2.9	10
2% thiourea in 20% HCl	104.0 + 3.7	13 33
	10 110 - 017	10100
0.3% thiourea in 40% HCl	105.1 ± 2.7	13 33
	103.1 ± 2.7	15.55
20% thiourso in 40% HCl	101.2 ± 4.2	12 22
270 unourea in 40% HCI	101.3 ± 4.2	15.55

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- 1 **Table 4.** Average Hg concentration (ng L^{-1}) ± std. deviation for 20 ng L^{-1} spike to different
- 2 matrices (n=3)

Sample	3D-Thiol method	Direct determination
Tap water ^b	23.27 ± 0.90	19.36 ± 0.45
Lake water 1 ^c	20.75 ± 1.46	15.23 ± 0.40
Lake water 2	20.81 ± 1.88	13.08 ± 0.09
River water	21.24 ± 3.96	12.02 ± 1.01
Pond water 1 ^{a,b}	18.96 ± 1.43	12.67 ± 0.44

3 ^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters, cn=2

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1 **Table 5.** Average Hg concentration (ng L^{-1}) ± std. deviation for different water samples with

Sample	3D-Thiol method	Direct determination
Tap water	0.61 ± 0.06	<mdl< td=""></mdl<>
Groundwater	0.34 ± 0.02	<mdl< td=""></mdl<>
Lake water 1	0.94 ± 0.09	<mdl< td=""></mdl<>
Lake water 2	1.79 ± 0.13	<mdl< td=""></mdl<>
River water ^{a,b}	1.39 ± 0.08	<mdl< td=""></mdl<>
Pond water 1 ^a	0.61 ± 0.12	<mdl< td=""></mdl<>
Pond water 1 ^{a,b}	0.65 ± 0.04	<mdl< td=""></mdl<>
Pond water 2	2.94 ± 0.12	4.44 ± 0.12

2 the 3D-Thiol method and direct determination (n=3)

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System/ Operating parameter	Type/Value/Mode
Sample introduction	ESI prepFAST
Nebulizer	PFA-ST nebulizer for prepFAST
Spray chamber	Baffled cyclonic, glass, cooled to 2 °C
RF power	1600 W
Plasma gas flow rate	18 L min ⁻¹
Auxiliary gas flow rate	1.2 L min ⁻¹
Nebulizer gas flow rate	0.89–0.93 L min ⁻¹
Isotope	The sum of ²⁰⁰ Hg, ²⁰¹ Hg, and ²⁰² Hg
Internal standard	¹⁹³ Ir at 100 μ g L ⁻¹ with 10 ms dwell time
Detector mode	Dual
Sweeps/reading	50
Replicates	5
Dwell time	100 ms
Scan mode	Peak hopping
Curve type	Linear through zero

Table 1. ICP-MS instrumental operating parameters and conditions

Isotherm model	Constants	Value for 3D-Thiol (n=1)	The average value for Thiol
			(n=3)
Langmuir	$q_{max} (\mathrm{mg g}^{-1})$	222.4	253.5
	K_L (L mg ⁻¹)	0.160	0.337
	R^2	0.975	0.940
Freundlich	$K_F(\mathrm{Lg}^{-1})$	23.5	54.2
	n	1.53	2.07
	R^2	0.982	0.980

Table 2. Constants of examined isotherms for Hg adsorption

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Table 3. Recovery (%) \pm std. deviation for Hg from 3D-Thiol using different eluent

compositions

Eluent	Recovery (%)	Required dilution factor for analysis
0.3% thiourea in 8% HCl	90.1 ± 3.8	2.67
1% thiourea in 8% HCl	99.7 ± 4.9	5
2% thiourea in 10% HCl	98.8 ± 2.9	10
2% thiourea in 20% HCl	104.0 ± 3.7	13.33
0.3% thiourea in 40% HCl	105.1 ± 2.7	13.33
2% thiourea in 40% HCl	101.3 ± 4.2	13.33

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Sample	3D-Thiol method	Direct determination
Tap water ^b	23.27 ± 0.90	19.36 ± 0.45
Lake water 1 ^c	20.75 ± 1.46	15.23 ± 0.40
Lake water 2	20.81 ± 1.88	13.08 ± 0.09
River water	21.24 ± 3.96	12.02 ± 1.01
Pond water 1 ^{a,b}	18.96 ± 1.43	12.67 ± 0.44

Table 4. Average Hg concentration (ng L^{-1}) ± std. deviation for 20 ng L^{-1} spike to different matrices (n=3)

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Table 5. Average Hg concentration (ng L^{-1}) ± std. deviation for different water samples with the 3D-Thiol method and direct determination (n=3)

Sample	3D-Thiol method	Direct determination
Tap water	0.61 ± 0.06	<mdl< td=""></mdl<>
Groundwater	0.34 ± 0.02	<mdl< td=""></mdl<>
Lake water 1	0.94 ± 0.09	<mdl< td=""></mdl<>
Lake water 2	1.79 ± 0.13	<mdl< td=""></mdl<>
River water ^{a,b}	1.39 ± 0.08	<mdl< td=""></mdl<>
Pond water 1 ^a	0.61 ± 0.12	<mdl< td=""></mdl<>
Pond water 1 ^{a,b}	0.65 ± 0.04	<mdl< td=""></mdl<>
Pond water 2	2.94 ± 0.12	4.44 ± 0.12

^afiltered with Whatman No. 41, ^btwo 3D-Thiol filters





Highlights

- Highly selective and accurate preconcentration method for Hg
- Hg adsorption onto 3D printed metal scavenger and elution with acidic thiourea solution
- Accurate Hg determination at picogram level in natural waters with a preconcentration factor of 92.8
- Determination of background concentrations in natural water with ICP-MS

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Declaration of interests

 \boxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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