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Author(s): Beyeh, Ngong Kodiah; Jo, Hyun Hwa; Kolesnichenko, Igor; Pan, FangFang; Kalenius,

Elina; Anslyn, Eric Van; Ras, Robin H. A.; Rissanen, Kari

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Resorcinarene Chlorides

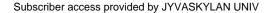
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Article

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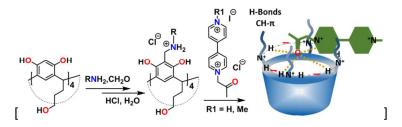
# The Recognition of Viologen Derivatives in Water by *N*-Alkyl Ammonium Resorcinarene Chlorides

Ngong Kodiah Beyeh,<sup>a\*</sup> Hyun Hwa Jo,<sup>b</sup> Igor Kolesnichenko,<sup>b</sup> Fangfang Pan,<sup>c</sup> Elina Kalenius,<sup>d</sup> Eric V. Anslyn,<sup>b\*</sup> Robin H. A. Ras<sup>a</sup> and Kari Rissanen<sup>d\*</sup>

<sup>a</sup> Aalto University, School of Science, Department of Applied Physics, Puumiehenkuja 2, FI-02150 Espoo, Finland

<sup>b</sup> The University of Texas at Austin, Department of Chemistry, Austin, Texas 78712, USA

Supporting Information Placeholder



**ABSTRACT:** Three water-soluble *N*-alkyl ammonium resorcinarene chlorides decorated with terminal hydroxyl groups at the lower rims were synthesized and characterized. The receptors were decorated at the upper rim with either terminal hydroxyl, rigid cyclohexyl or flexible benzyl groups. The binding affinities of these receptors towards three viologen derivatives, two of which possess an acetylmethyl group attached to one of the pyridine nitrogens, in water were investigated via <sup>1</sup>H NMR spectroscopy, fluorescence spectroscopy, and isothermal titration calorimetry (ITC). ITC quantification of the binding process gave association constants of up to 10<sup>3</sup> M<sup>-1</sup>. Analyses reveal a spontaneous binding process which are all exothermic, and are both enthalpy and entropy driven.

### INTRODCUTION

Molecular receptors with pre-organized cavities suitable for guest recognition is a continuously developing area in supramolecular chemistry, material science, and biology. Receptors capable of guest recognition in aqueous media and biological fluids have a growing importance, relative to receptors that primarily function in organic media. However, receptors that can operate in aqueous media have proven to be difficult to design. However, they have potential biocompatibility if one can exploit the cohesive force of water.

Viologens, with their extended  $\pi$ -conjugation, can exhibit excellent electrochromic and photochromic properties. <sup>14-16</sup> These species are commonly used in supramolecular chemistry to construct capsular assemblies or threaded structures with several host compounds. <sup>17-21</sup> Their cationic nature makes them suitable guests for  $\pi$ -rich receptors. <sup>17-21</sup> The complexation of viologen derivatives, by receptors such as curcubiturils, can substantially alter the kinetics and thermodynamics of their electron transfer reactions. <sup>17,18</sup> Modified viologens have potential as sensors for bisulfite in water which can be utilized in the beverage industry. <sup>22-25</sup>

*N*-Alkyl ammonium resorcinarene halides (NARXs), resulting from the ring opening of tetrabenzoxazines in the presence of mineral acids under refluxing conditions, are stabilized by a seam of hydrogen-bonded cation-anion interactions. <sup>26,27</sup> The NARX receptors possess four spatially fixed halide anions with deep cavities for guest binding. Neutral and anionic guests have been shown to reside in the cavity of NARXs, interacting with the host mainly through

<sup>&</sup>lt;sup>c</sup> Central China Normal University, College of Chemistry, Key Laboratory for Pesticides and Chemical Biology, 152 Luoyu Road, Wuhan, Hubei 430079, China

<sup>&</sup>lt;sup>d</sup> University of Jyvaskyla, Department of Chemistry, Nanoscience Centre, P. O. Box 35, FI-40014 Jyvaskyla, Finland

CH- $\pi$  interactions and hydrogen bonds.<sup>28,29</sup> We recently reported the binding of small neutral molecules such as amides and diamides in organic media with cooperativity by NARX receptors.<sup>30</sup> The four spatially fixed anions act as halogen bond acceptors leading to a variety of complex architectures, such as deep-cavity cavitands, pseudo-capsular, and capsular assemblies.<sup>31,32</sup> The NARXs are generally soluble in alcoholic and non-polar solvents. Recently we synthesized the first water soluble NARX receptors by attaching terminal hydroxyl groups at the upper rim.<sup>33</sup> Therein, we showed that the water soluble NARXs exist in  $C_{4\nu}$  crown conformation and bind a variety of aliphatic, aromatic and halogenated alkanes and arenes in aqueous media.<sup>33</sup>

In this study, three new water-soluble NARCl receptors decorated with four terminal hydroxyl groups at the lower rims were synthesized and characterized (Figure 1). The first NARCl receptor (5), is also functionalized at the upper rim with four terminal hydroxyl groups, making it extremely water soluble (35 mg/mL). The receptor (7) is functionalized at the upper rim with four rigid cyclohexyl groups and the third receptor (9) possesses four flexible benzyl groups at the upper rim. Mono-methyl 4,4'-bipyridine (11),<sup>34</sup> mono-acetylmethyl 4,4'-bipyridine (12), and hetero methyl-acetylmethyl 4,4'-bipyridine (13) molecules were also synthesized as potential guests. The recognition of these guests (11-13) by the NARX receptors (5, 7 and 9) were investigated in water via <sup>1</sup>H NMR spectroscopy, fluorescence spectroscopy, and isothermal titration calorimetry (ITC) analyses.

#### **RESULTS AND DISCUSSION**

**Synthesis of the receptors and the viologen guests.** The synthesis of **5**, **7** and **9** starts with resorcinarene **3**, which was synthesized through reported procedures.<sup>35</sup> Ethanolamine, cyclohexyl amine and benzyl amine in the presence of excess formaldehyde participate in a Mannich condensation with **3** to form tetrabenzoxazines **4**, **6** and **8**, respectively.<sup>36,37</sup> The reaction with ethanolamine leads to a mixture of the five and six membered azoxazine rings **4a** and **4b**, respectively. The un-isolated crude product containing the five- and six-membered ring compounds in the presence of concentrated HCl under refluxing conditions, lead to the same final product **5** (Figure 1). Cleavage of the pure tetrabenzoxazines **6** and **8** under similar conditions give NARCls **7** and **9**. The detailed synthetic procedures of the NARCl receptors are reported in the Supporting Information (Schemes S1-S3; Figures S1-S5).

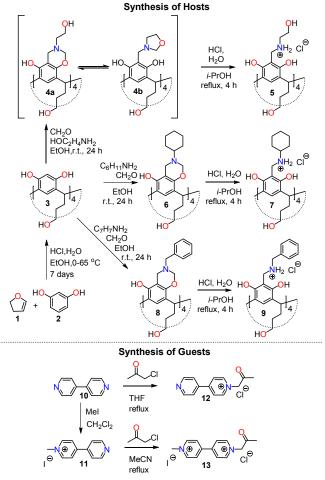
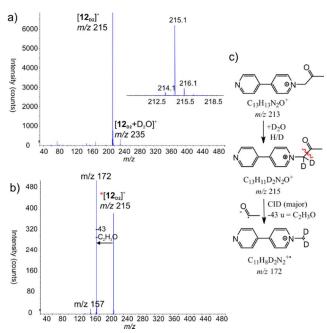


Figure 1. Synthesis of water soluble NARCls 5, 7 and 9 as receptors and derivatives of 4,4'-bipyridine 11-13 as guests.

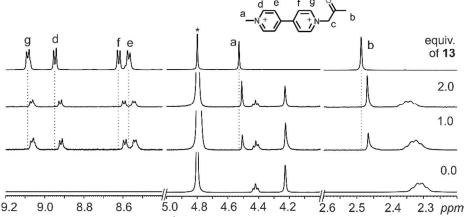
The 4,4'-bipyridine guest 11 was synthesized according to a reported procedure.<sup>34</sup> The other bipyridine guests, 12 and 13, were synthesized by reacting chloroacetone with 4,4'-bipyridine or guest 11, respectively (Schemes S4, S5; Figures S6-S8). Suitable single crystals of mono-methyl 4,4'-bipyridine (11) and hetero- methyl-acetylmethyl 4,4-bipyridine (13) were obtained and analyzed (Figures S9-S11). Structural analysis verified the bipyridine was successfully substituted. In 11, the *N*-methyl 4,4'-bipyridyl cation is paired with the iodide anion with the anion close to the cationic nitrogen of the viologen molecule. While in 13, although the dicationic viologen is as expected, the original counter anions are replaced by 1.5  $\Gamma$  and 0.5  $\Gamma$  during the crystallization. Electrostatic forces contribute to the arrangement of the ion pairs. A mechanism for formation of the triiodide from iodide was proposed in a recent crystallographic study.<sup>38</sup>

**Hydrogen/Deuterium (H/D) Exchange Studies of the Viologen Guests.** In D<sub>2</sub>O, the labile -NCH<sub>2</sub>CO- hydrogens undergo H/D exchange. These protons are therefore not observed in the <sup>1</sup>H NMR spectra of all samples containing guests **12** and **13** in protic deuterated solvents. The lability of these hydrogens was verified by electrospray ionization mass spectrometry (ESI-MS) in a combined experiment of solution H/D-exchange and collision induced dissociation (CID). In D<sub>2</sub>O (Figure 2, Figures. S12, S13), two H/D-exchanges of -NCH<sub>2</sub>CO- hydrogens were observed. The location of exchanged hydrogens was verified by CID experiment, which showed the fragmentation to be initiated by elimination of undeuterated acetyl radical (C<sub>2</sub>H<sub>3</sub>O•, 43 u), leaving only one plausible location for H/D-exchange at -NCH<sub>2</sub>CO-. The CID experiments also showed the increased stability of keto tautomer as compared to possible enol form. This is likely due to existence of two resonance structures of keto tautomer (see SI, Figure S12).



**Figure 2**. ESI-MS of **12** in D<sub>2</sub>O. (a) Profile spectrum showing two H/D-exchanges, inset showing zoomed view for m/z 215. (b) CID for isolated  $[\mathbf{12}_{D2}]^+$  ion (CE=29). (c) Main fragmentation pathway for ion  $[\mathbf{12}_{D2}]^+$ .

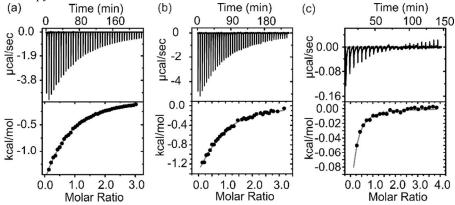
Complexation Studies via NMR Spectroscopy. Complexation studies between the NARCl receptors (5, 7 and 9) with the modified 4,4-bipyridine guests (11-13) were investigated by  $^{1}H$  NMR experiments in  $D_{2}O$ . The receptors possess  $C_{4\nu}$  symmetry in solution as observed from their relatively simple  $^{1}H$  NMR spectra (Figures S1, S3 and S5). Varying complexation-induced shift changes of the different guest signals were observed from either the shielding effects of the aromatic rings of the bowl-shaped host cavity or interaction with the cation-anion seam. The complexation process is fast on the NMR time scale at 298 K. The guests are soluble in water and can interact with the hosts mainly through CH- $\pi$  and hydrogen bond interactions with the cation-anion seam of the receptors. The small shift changes of the guests can be attributed to the highly competitive nature of the bulk water. Taking the complex 13@7 as an example, upfield shifts of the guest  $-COCH_{3}$ - protons are observed (Figure 3). The carbonyl oxygens are known to interact with the cation-anion seam in organic media as reported in the binding of amides by NARXs. Upfield shift of methyl protons of ammonium cations are usually observed when located deep in the cavity of resorcinarene-type receptors. The relatively small upfield shifts of the methyl protons of guest 13 therefore suggest the binding interaction to occur mainly at the upper rim of the receptors involving the hydrogen-bonded cation-anion seam with minimal interaction with the electron rich interior cavity of the resorcinarene cavity (Figure 3).



**Figure 3**. Selected region of the <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O, 298 K) spectra observed upon the addition of the guest **13** to the host **7**. Dotted lines gives an indication of the complexation induced shift changes.

The <sup>1</sup>H NMR spectra of the 1:1 mixture between the guests **11** and **12** reveal mainly downfield shifts of the guests signals (Figures S15-S20). Limited and/or no shift changes were observed for the guests methyl signals. This again suggest the interaction between the guests **11** and **12** with the receptors to be mainly at the upper rim of the receptors involving the hydrogen-bonded cation-anion seam of the receptors and the free pyridine nitrogens of the guests (Figures S15-S22). Such hydrogen bond interactions between the pyridine nitrogen of the guests (11 and 12) leads to de-shielding contrary to the guest 13 with no free pyridine nitrogen.

Quantification of the Binding Process via Isothermal Titration Calorimetry (ITC) Studies. The interaction between the hosts and guests was quantified through a series of ITC experiments in  $H_2O$  (Figure 4, Figures S23, S24). The thermodynamic parameters of host-guest binding  $(K, \Delta H, \Delta S, \text{ and } \Delta G)$  were extracted from fitting to a single binding site model (Table 1). When comparing the ITC titrations of the three NARCl receptors (5, 7 and 9) with the guests (11-13), several considerations can be made. The  $\Delta H$  and  $\Delta G$  values indicate the binding process to be exothermic and spontaneous at 298 K.  $\Delta H$  and  $T\Delta S$  results also indicate that complexation of guests 11-13 by the receptors to be both enthalpy and entropy driven in most cases. In three cases (11@5, 13@5, 13@9), negative  $\Delta S$  values indicate these process to be enthalpy driven.



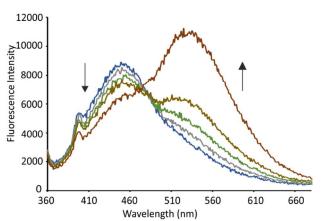
**Figure 4**. ITC traces of the titration of guests (10 mM) into host 7 (1 mM) in H<sub>2</sub>O at 298 K. (a) guest 11, (b) guest 12, (c) guest 13. All data were fitted into a one site-model.

Table 1. Thermodynamic binding parameters of formed complexes between the hosts 5,7 (10 mM) and the guests 9-11 (1 mM) by ITC<sup>[a]</sup>

Complex	$K(\times 10^3)$	$\Delta H$	$T\Delta S$	$\Delta G$
	$[M^{-1}]$	[kcal/mol]	[kcal/mol]	[kcal/mol]
11@5	$0.91\pm0.09$	-8.698	-4.657	-4.041
11@7	$1.17 \pm 0.04$	-4.085	0.103	-4.188
11@9	$0.95\pm0.02$	-3.422	0.644	-4.066
12@5	$2.29\pm0.30$	-1.686	2.896	-4.582
12@7	$1.49\pm0.14$	-2.602	1.728	-4.330
12@9	$1.05\pm0.09$	-3.595	0.527	-4.122
13@5	$1.70\pm0.50$	-25.01	-20.58	-4.422
13@7	$2.78\pm0.83$	-0.567	4.130	-4.697
13@9	1.37±0.14	-45.970	-41.660	-4.310

 $<sup>^{[</sup>a]}$  ITC in  $H_2O$  at 298 K.

Complexation Studies via Fluorescence Spectroscopic Analysis. We also used optical titrations to analyze the binding processes. Fluorescence enhancement and/or quenching were observed from titration experiments conducted at 298 K with solutions of the hosts 5, 7 and the guests 11-13. A solution of the guests (0.1 M) at 298 K was titrated into a solution of the hosts 5 and 7 (2 mL, 125  $\mu$ M). In all cases, fluorescence spectra were collected after thermal equilibration at 298 K (Figure 5, Figure S25-S29).



**Figure 5**. Fluorescence changes of guest **13** (0.1 M) when titrated to host **7** (125 μM, 2 mL) in H<sub>2</sub>O at 298 K. Total of 4 additions of guest **13** (0.4, 1.0, 2.0, 8.0 equiv.) were added to host **7**.

The red shift in fluorescence emission spectra of the guest 13, registering two maximum (458 nm and 545 nm), was observed. The lack of a clear isosbestic point indicates that there are more than two species in the system, which is hypothesized to be the formation of aggregates due to the high concentration of guests used in the system. Fluorescence enhancement was observed for the guests during the titration with the hosts 5 and 7. The excited-state vibrational dynamics of the viologen guests 11-13 appears to play the key role in the observed enhancement, as well as the red shift of the emission when the viologens concentration is increased. Studies by Galoppini and coworkers, and Pal and coworkers, show similar behavior of a viologen and the dye Brilliant Green, a triphenylmethane derivative, where restriction of intramolecular bond rotations between the aryl rings induced by complexation/encapsulation with cucurbiturils resulted in enhanced fluorescence.

### CONCLUSION

In conclusion, we synthesized three water-soluble NARCI receptors (5, 7 and 9) with varying hydrophilicity of the upper rim substituents. These receptors exists in the  $C_{4v}$  bowl-shaped conformation as observed from their relatively simple <sup>1</sup>H NMR spectra. The binding properties of the NARCI receptors and three water-soluble viologen derivatives (11-13) were investigated in water via NMR, ITC, and fluorescence studies. Binding constants of  $10^3$  M<sup>-1</sup> were observed via ITC analyses. The hosts show higher affinity towards the acetylmethyl-derived viologen guests 12 and 13 over the methyl viologen derivative 11. The higher affinity can be attributed to hydrogen bond interactions between the host cation-anion seam and the guest carbonyl groups. This study illustrates the versatility of the NARXs, which in water possesses hydrophobic cavities and hydrophilic cation-anion seam. The ease of functionalization of resorcinarene type receptors into water soluble NARCI receptors make resorcinarenes a very interesting class of receptor compounds. This versatility renders the NARXs as suitable receptors for a variety of guests in water.

#### **EXPERIMENTAL SECTION**

General Methods. The *N*-alkyl ammonium resorcinarene chlorides 5, 7 and 9 were synthesized accordingly to modified procedures. <sup>26,27,36</sup> <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker Avance DRX 500 (500 MHz for <sup>1</sup>H and 126 MHz for <sup>13</sup>C) and DRX 400 (400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C) spectrometers. All signals are given as δ values in ppm using residual solvent signals as the internal standard. Coupling constants are given in Hz. Melting points were determined with a Mettler Toledo FP62 capillary melting point apparatus and a Stuart SMP30 melting point apparatus. Experimental details for the synthesis and characterization data of receptors 5, 6, 7, 8 and 9, and guests 12 and 13 are below. Compounds 3, and 11 are known compounds and have been reported in previous references. <sup>34,35,41</sup> Mass spectrometry experiments were performed with ABSciex QSTAR Elite ESI-Q-TOF mass spectrometer, equipped with an API 200 TurbolonSpray ESI source from AB Sciex. Nitrogen was used as drying and nebulization gas. VP-ITC instrument made by MicroCal were used to determine the molar enthalpy (ΔH) of complexation. Subsequent fitting of the data to a 1:1 binding model using Origin software provides binding constant (K) and the entropy (ΔS). Fluorescence spectra were recorded on a PTI QuantaMasterTM 40 intensity based spectrofluorometer equipped with 814 photomultiplier detection system (V=1000 volts). A 75 W Xenon arc lamp was used as the excitation source. The data for crystals of 11, 11b, and 13 were collected at 123 K for 11b with an Agilent Super-Nova diffractometer using mirror-monochromatized Cu-Kα (λ=1.54184Å) radiation, and at 100 K for 11 and 13 with the same diffractometer using mirror-monochromatized Mo-Kα (λ=0.71073Å) radiation.

General procedure for the synthesis of tetrabenzoxazines from the resorcinarene 3. To a solution of the resorcinarene 3 (5.5 mmols) and excess formaldehyde (6 mL) in ethanol (40 mL), the amine (23.3 mmols) in ethanol (15 mL) is added slowly and stirred at room temperature for 24 h. The precipitate that separated is filtered, recrystallized in a methanol/n-hexane mixture and dried.

General procedure for the synthesis of the *N*-alkyl ammonium resorcinarene chlorides from the tetrabenzoxazines. Into a solution of the tetrabenzoxazine (0.82 mmol), 3 mL concentrated HCl (37%) and 4 ml H<sub>2</sub>O in 50 ml isopropanol is heated under reflux. Water and formaldehyde are removed by azeotropic distillation with chloroform. The remaining isopropanol is evaporated and the crude product triturated with diethyl ether to give the *N*-alkyl ammonium resorcinarene chloride.

*N-Ethanol ammonium resorcinarene chloride* (*5*):  $C_{propanol}$ -Resorcinarene **3** (4 g, 7.344 mmols), formaldehyde (8 mL), EtOH (60 mL), 2-aminoethanol (1.86 mL, 30.8 mmols), EtOH (15 ml). The product was a mixture of the six-membered and five-membered tetrabenzoxazines **4a/4b**. This mixture of tetrabenzoxazines was not separated and was used directly in the next step to obtain the *N*-Ethanol ammonium resorcinarene chloride **5**. The crude tetrabenzoxazines **4a/4b** (1.0 g, 1.129 mmol), 3 ml conc. HCl, 4 ml H<sub>2</sub>O, 40 ml isopropanol. *N*-Ethanol ammonium resorcinarene chloride **5** (0.68 g, 61 %). m.p. > 300 °C; HRMS (ESI-TOF) m/z calcd for  $C_{52}H_{77}N_4O_{16}$  [**5**-4Cl-3H]<sup>+</sup> 1013.5329, Found 1013.5331, (-0.2 ppm);. <sup>1</sup>H NMR (400 MHz, 298K in CD<sub>3</sub>OD) δ (ppm): 1.53 (m, 8H, CH<sub>2</sub>), 2.39 (m, 8H, CH<sub>2</sub>), 3.04 (t, J=5.06 Hz, 8H, OCH<sub>2</sub>), 3.66 (t, J=6.20 Hz, 8H, NCH<sub>2</sub>), 3.78 (t, J=5.12 Hz, 8H, OCH<sub>2</sub>), 4.32 (s, 8H, ArCH<sub>2</sub>N), 4.47 (t, J=7.80 Hz, 4H, CH), 7.48 (s, 4H, ArH); <sup>13</sup>C NMR: (100 MHz, 298K in D<sub>2</sub>O) δ (ppm) = 30.9, 32.0, 35.8, 42.6, 57.6, 62.7, 110.2, 126.7, 128.1, 151.9.

*N-Cyclohexyl Tetrabenzoxazine* (*6*):  $C_{propanol}$ -Resorcinarene (4 g, 5.5 mmols), formaldehyde (6 mL), ethanol (40 mL), cyclohexyl amine (2.31 mL, 23.3 mmols), ethanol (15 mL). Tetrabenzoxazine 6 (5.29 g, 79 %). m.p. 218-220 °C; HRMS (ESI-TOF) m/z calcd for  $C_{72}H_{101}N_4O_{12}$  [*6*+H]<sup>+</sup> 1213.7411, Found 1213.7422, (-0.9 ppm); <sup>1</sup>H NMR (400 MHz, 298 K in [D<sub>6</sub>]DMSO) δ (ppm): 0.98-1.92 (m, 48H, CH<sub>2</sub>), 2.24 (m, 8H, CH<sub>2</sub>), 2.43 (m, 8H, CH<sub>2</sub>), 3.44 (q, J=4.96 Hz, 8H, OCH<sub>2</sub>), 3.77 (dd, 8H, Ar-CH<sub>2</sub>-N), 4.08 (t, J=7.94 Hz, 4H, OCH<sub>2</sub>), 4.32 (t, J=4.90 Hz, 4H, OH), 5.06 (dd, J=9.64 Hz, 8H, Ar-CH<sub>2</sub>-O), 7.40 (s, 4H, Ar-H), 7.62 (s, 4H, Ar-OH); <sup>13</sup>C NMR: (100 MHz, 298 K in [D<sub>6</sub>]DMSO) δ (ppm) = 24.2, 24.8, 24.9, 25.4, 29.0, 30.3, 31.0, 31.8, 32.0, 42.9, 57.1, 60.4, 60.5, 80.3, 108.6, 122.1, 123.4, 123.8, 148.6.

*N-Cyclohexyl ammonium resorcinarene chloride (7):* Tetrabenzoxazine **6** (1.0 g, 0.824 mmol), 3 mL concentrated HCl (37%), 4 ml H<sub>2</sub>O, 50 ml isopropanol, *N*-Cyclohexyl ammonium resorcinarene chloride **7** (1.00 g, 92 %). m.p. > 300 °C; HRMS (ESI-TOF) m/z calcd for  $C_{68}H_{101}N_4O_{12}$  [7-4Cl-3H]<sup>+</sup> 1165.7411, Found 1165.7396, (1.3 ppm). <sup>1</sup>H NMR (500 MHz, 298K in D<sub>2</sub>O) δ (ppm): 0.88 (m, 12H, CH<sub>2</sub>), 1.11 (m, 12H, CH<sub>2</sub>), 1.45 (m, 12H, CH<sub>2</sub>), 1.56 (d, J=13.00 Hz, 8H, CH<sub>2</sub>), 1.79 (d, J=9.50 Hz, 8H, CH<sub>2</sub>), 2.26 (m, 8H, CH<sub>2</sub>), 2.68 (m, 4H, NCH), 3.61 (t, J=6.45 Hz, 8H, OCH<sub>2</sub>), 4.20 (s, 8H, Ar-CH<sub>2</sub>-N), 4.38 (t, J=7.77 Hz, 4H, CH), 7.38 (s, 4H, Ar-H); <sup>13</sup>C NMR: (126 MHz, 298K in D<sub>2</sub>O) δ (ppm) = 14.0, 23.8, 24.2, 28.7, 29.5, 29.7, 34.2, 38.6, 56.5, 61.5, 65.9, 109.0, 125.3, 126.9, 150.1.

*N-Benzyl Tetrabenzoxazine* (8):  $C_{propanol}$ -Resorcinarene (5 g, 6.9 mmols), formaldehyde (10 mL), ethanol (60 mL), benzyl amine (3.18 mL, 29.0 mmols), ethanol (15 mL). Tetrabenzoxazine 8 (4.74 g, 55 %). m.p. 179-181 °C; HRMS (ESITOF) m/z calcd for  $C_{76}H_{85}N_4O_{12}$  [8+H]<sup>+</sup> 1245.6159, Found 1245.6133, (2.1 ppm). <sup>1</sup>H NMR (400 MHz, 298 K in [D<sub>6</sub>]DMSO) δ (ppm): 1.36 (m, 8H, CH<sub>2</sub>), 2.24 (t, J=6.45 Hz, 8H, CH<sub>2</sub>), 3.42 (m, 8H, CH<sub>2</sub>), 3.70-3.85 (m, 16H, Ar-CH<sub>2</sub>-N, OCH<sub>2</sub>), 4.20 (t, J=7.87 Hz, 4H, OH), 4.33 (t, J=5.42 Hz, 4H, CH), 4.89 (dd, J=9.35 Hz, 8H, Ar-CH<sub>2</sub>-O), 7.19-7.26 (m, 20H, Ph-H), 7.41 (s, 4H, Ar-H), 7.64 (s, 4H, Ar-OH); <sup>13</sup>C NMR: (100 MHz, 298 K in [D6]DMSO) δ (ppm) = 29.3, 31.0, 31.1, 45.3, 54.8, 60.5, 107.4, 122.6-128.5, 138.1, 147.9, 149.6.

*N-Benzyl ammonium resorcinarene chloride* (*9*): Tetrabenzoxazine **8** (1.0 g, 0.803 mmol), 3 mL concentrated HCl (37%), 4 ml H<sub>2</sub>O, 50 ml isopropanol. *N*-Benzyl ammonium resorcinarene chloride **9** (0.95 g, 88 %). m.p. > 300 °C; HRMS (ESI-TOF) m/z calcd for  $C_{72}H_{85}N_4O_{12}$  [9-4Cl-3H]<sup>+</sup> 1197.6159, Found 1197.6178, (-1.6 ppm). <sup>1</sup>H NMR (500 MHz, 298K in [D<sub>6</sub>]DMSO) δ (ppm): 1.37 (m, 8H, CH<sub>2</sub>), 2.32 (m, 8H, CH<sub>2</sub>), 3.46 (t, J=6.58 Hz, 8H, CH<sub>2</sub>), 3.98 (br, 8H, NCH<sub>2</sub>Ph), 4.14 (br, 8H, ArCH<sub>2</sub>N), 4.31 (t, J=7.57 Hz, 4H, CH), 7.38 (m, 12H, Ph-H), 7.56 (m, 8H, Ph-H), 7.64 (m, 4H, Ar-H), 9.05 (br, 8H, NH<sub>2</sub>), 9.44 (br, 8H, Ar-OH); <sup>13</sup>C NMR: (126 MHz, 298K in [D<sub>6</sub>]DMSO) δ (ppm) = 25.8, 29.2, 31.3, 34.5, 41.0, 50.6, 60.7, 109.3, 126.4, 126.8, 128.9, 129.2, 130.4, 131.9, 150.5.

Synthesis of N-methylcarbonylmethyl-4-,4'-bipyridinium chloride (12). A flask was flame dried and back filled with nitrogen. 1.72 g (11 mmol) of 4,4'-bipyridyl were added to this and dissolved in 22 mL of dry THF. 3.6 mL (44 mmol) of chloroacetone were added and the solution was stirred at room temperature overnight. The solvent was removed under reduced pressure, yielding 2.08 g (76 %) of 12. HRMS (ESI-TOF) m/z calcd for C<sub>13</sub>H<sub>13</sub>N<sub>2</sub>O [12+H]<sup>+</sup> 213.1022, Found 213.1020, (1.3 ppm). <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were obtained in D<sub>2</sub>O and [D<sub>6</sub>]-DMSO. <sup>1</sup>H NMR (400 MHz, 298 K in

[D<sub>6</sub>]-DMSO)  $\delta$  (ppm): 2.34 (s, 3H, CH<sub>3</sub>), 5.95 (s, 2H, CH<sub>2</sub>), 8.06 (d, J=6.16, 2H, CH<sub>2</sub>), 8.71 (d, J=6.88, 2H, ArH), 8.86 (d, J=6.12 Hz, 2H, ArH), 9.11 (d, J=6.92 Hz, 2H, ArH); <sup>13</sup>C NMR: (100 MHz, 298K in [D6]-DMSO)  $\delta$  (ppm) = 27.6, 68.2, 122.4, 125.4, 146.8, 151.4.

Synthesis of N-methyl-N'-methylcarbonylmethyl-4,4'-bipyridinium dihalide (13). 18.10 g (61 mmol) of 11 were dissolved in 400 mL of acetonitrile. 4.45 mL (61 mmol) of chloroacetone were added and the solution was refluxed overnight, producing a light brown precipitate. The solid was filtered out and dried under reduced pressure to yield 15.47 g (73 %) of 11. HRMS (ESI-TOF) m/z calcd for  $C_{14}H_{16}N_2O$  [13-Cl-I]<sup>2+</sup> 114.0626 (228.1252 u), Found 114.0629, (3.01 ppm).  $^{1}$ H- and  $^{13}$ C-NMR spectra were obtained in  $D_2O$  and  $[D_6]$ -DMSO.  $^{1}$ H NMR (400 MHz, 298 K in  $[D_6]$ -DMSO)  $\delta$  (ppm): 2.36 (s, 3H, CH<sub>3</sub>), 4.47 (s, 3H, CH<sub>3</sub>), 5.98 (s, 2H, CH<sub>2</sub>), 8.82 (d, J=6.80, 2H, ArH), 8.89 (d, J=6.84, 2H, ArH), 9.23 (d, J=6.76 Hz, 2H, ArH), 9.35 (d, J=6.72 Hz, 2H, ArH);  $^{13}$ C NMR: (100 MHz, 298K in  $[D_6]$ -DMSO)  $\delta$  (ppm) = 27.6, 48.4, 68.7, 126.6, 147.1, 147.3.

#### **ASSOCIATED CONTENT**

### **Supporting Information**

The Supporting Information is available free of charge on the ACS Publications website at http://pubs.acs.org.

Experimental details, copies of the <sup>1</sup>H and <sup>13</sup>C NMR, NMR and fluorescence spectroscopy, ITC details, and Mass Spectrometry (PDF).

X-ray crystallographic data. CCDC 1525501-1525503 for 11, 11b and 13, respectively contains the supplementary crystallographic data for this paper and can be obtained free of charge from the Cambridge Crystallographic Data Centre via <a href="https://www.ccdc.cam.ac.uk/data">www.ccdc.cam.ac.uk/data</a> request/cif.

#### **AUTHOR INFORMATION**

# **Corresponding Authors**

E-mail: kodiah.beyeh@aalto.fi
E-mail: anslyn@austin.utexas.edu
E-mail: kari.t.rissanen@jyu.fi

#### ORCID

Ngong Kodiah Beyeh: 0000-0003-3935-1812 Eric Anslyn: 0000-0002-5137-8797 Kari Rissanen: 0000-0002-7282-8419

#### **Author Contributions**

The manuscript was written through contributions of all authors.

#### Notes:

The authors declare no competing financial interest.

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