Promoter targeted histone acetylation of chromatinized parvoviral genome is essential for infection progress

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ABSTRACT

The association of host histones with parvoviral DNA is poorly understood. We analyzed the chromatinization and histone acetylation of canine parvovirus DNA during infection by confocal imaging and *in situ* proximity ligation assay combined with chromatin immunoprecipitation and high-throughput sequencing. We found that at late infection parvovirus replication bodies were rich in histones bearing modifications characteristic of transcriptionally active chromatin, i.e. histone H3 lysine 27 acetylation (H3K27ac). The H3K27ac, in particular, was located in close proximity to the viral DNA-binding protein NS1. Importantly, our results show for the first time that in the chromatinized parvoviral genome, particularly the two viral promoters were rich in H3K27ac. Histone acetyltransferase (HAT) inhibitor efficiently interfered with expression of viral proteins and infection progress. Altogether, our data suggest that acetylation of histones on parvoviral DNA is essential for viral gene expression and completion of viral life cycle.

IMPORTANCE

Viral DNA introduced into cell nuclei is exposed to cellular responses to foreign DNA including chromatinization and epigenetic silencing, both of which determine the outcome of infection. How the incoming parvovirus resists cellular epigenetic down-regulation of its genes is not understood. Here, the critical role of epigenetic modifications in regulation of parvovirus infection was demonstrated. We showed for the first time that a successful parvovirus infection is characterized by deposition of nucleosomes with active histone acetylation on the viral promoter areas. The results provide new insights to regulation of parvoviral gene expression which is an important aspect in the development of parvovirus-based virotherapy.

INTRODUCTION

Nuclear chromatin is composed of DNA and histone proteins (1). The histone proteins assemble DNA into nucleosomes, the composition and spacing of which contribute to higher order chromatin packing. The chromatin is organized into regions of less-condensed actively transcribed chromatin (euchromatin) and highly-condensed transcriptionally repressed chromatin (heterochromatin). Epigenetic modifications of histone proteins have been shown to correlate with spatial distribution of active and repressed chromatin (2,3). Acetylation of lysines 9 or 27 of histone H3 (H3K9ac and H3K27ac) and tri-methylation of lysine 4 (H3K4me3) correlate with transcriptional activity, while repressed chromatin is characterized by e.g. tri-methylation or di-methylation of the same H3 lysine residues (H3K9me3 and H3K27me3; H3K9me2 and H3K27me2) (4-8).

Foreign DNA introduced into mammalian cells can be recognized as threat by the host cell. The cellular responses to the foreign DNA, such as viral DNA, include chromatinization of the entering DNA, leading to its transcriptional silencing (9–11). How viruses resist cellular chromatinization and silencing is known for only few viruses (9,12,13). These include herpes simplex viruses (HSVs) (14-16), polyomaviruses (17, 18), adenoviruses (19), and cytomegaloviruses (20,21), all of which encode proteins that promote transcriptionally activating histone modifications on chromatinized viral genome during lytic infection.

In parvovirus infection, the nuclear entry of viral single-stranded DNA is followed by the formation of double-stranded replicative intermediates, nuclear accumulation of viral proteins and DNA, and the formation of autonomous parvovirus-associated replication (APAR) bodies (22–25). Viral gene expression and DNA replication are

dependent on the S-phase of the cell cycle. In infected cells transcription of viral non-structural protein 1 (NS1) at 4 h p.i. is followed by viral genome replication (26). Replication continues throughout the infection and leads to production of viral capsids and their nuclear egress at 20-24 h p.i. Intranuclear chromatinization of parvoviral genome, modification of the assembled histones and effect of these events on viral gene expression are not well understood. To provide answers to these questions we analyzed the infection of an autonomous protoparvovirus, canine parvovirus (CPV), by confocal imaging techniques and *in situ* proximity ligation assays (PLA) complemented with chromatin immunoprecipitation coupled with high-throughput sequencing (ChIP-seq). Our results demonstrated enrichment of acetylated histones (H3K27ac) in close proximity to viral DNA-binding NS1 in APAR bodies and, especially H3K27ac accumulated in the viral promoters. Inhibition of histone acetyltransferase (HAT) activity led to interruption of viral life cycle. These results reveal that histone acetylation on chromatinized parvoviral genome is necessary for expression of viral genes and successful progress of infection.

MATERIALS AND METHODS

Cells, viruses and constructs. Norden laboratories feline kidney (NLFK; Quality Control of Pfizer Animal Health, Lincoln, NE) cells were grown in Dulbecco´s modified Eagle medium (DMEM) supplemented with 10 % fetal bovine serum (Gibco, Paisley, UK) at 37 °C in the presence of 5 % CO_2. CPV-2d isolates originated from an infectious plasmid clone (a gift from C.R. Parrish, Cornell University, Ithaca, NY (27)). The viruses had been isolated as described by Suikkanen et al. (28). For infection, the cells were inoculated with CPV (MOI 1–2) and kept at 37 °C until fixation. In order to synchronize infections, the cells were incubated on ice at 4 °C for

20 min prior to virus addition. The cells were then inoculated on ice to allow for the virus adsorption to occur for 30 min. Next, the cells were rinsed at 4 °C with phosphate-buffered saline (PBS) containing 0.3 % bovine serum albumin (BSA) followed by addition of 37 °C medium. The plasmid encoding fluorescent H3-EGFP was a generous gift from J. Langowski (German Cancer Research Center, Heidelberg, Germany). A NLFK cell line stably expressing H3-EGFP was established by transfection (TransIT-LT1 reagent; Thermo Fisher Scientific Inc, Waltham, MA) with an expression vector at 24 h after seeding. After 2 days the DMEM was replaced by DMEM containing 1 mg/ml of geneticin (Sigma Aldrich, St. Louis, MO). The cells were then seeded at different intervals until a stable expression was observed by microscopy.

Confocal microscopy For immunolabeling, cells seeded on round coverslips were infected with CPV and fixed at 8, 10, 12, 16, and 24 h p.i. with 4 % paraformaldehyde (PFA; 15 min at room temperature). Viral proteins were detected with an NS1-specific monoclonal antibody (MAb) (generous gift from Caroline Astell) (29), an intact capsid MAb and a polyclonal capsid-protein VP2 antibody (Ab; generous gifts from Colin R. Parrish) (27) followed by goat anti-mouse or anti-rabbit Alexa-555 or Alexa-633 conjugated secondary Abs (Molecular Probes, Life technologies, Grand Island, NY). Modified histones were labeled with rabbit Abs against H3K9me3 and H3K27ac (Abcam, Cambridge, MA) followed by goat anti-rabbit Alexa-633 conjugated secondary Abs. For deacetylation and hyper-acetylation studies, cells were either pretreated for 30 min prior to virus inoculation or treated at 0-6 h p.i. with 0.1-mM anacardic acid prepared in 100 % dimethyl sulfoxide (DMSO) (Sigma Aldrich, St. Louis, MO). Cell viability was assessed using propidiun iodide (Sigma) and annexin V Alexa-647 (Molecular Probes) staining following manufacturer instructions (data not

shown). All cells were fixed at 24 h p.i. with 4 % PFA and permeabilized with 0.1 % Triton X-100 in PBS supplemented with 1 % BSA and 0.01 % sodium azide. Imaging was done with an Olympus FV-1000 confocal microscope with the UPLSAPO 60x oil-immersion objective (NA=1.35). EGFP was excited with a 488 nm argon laser and fluorescence was collected with a 515/30 nm band-pass filter. Alexa-555 and the PLA reagent 594 nm were excited with a 543 nm He-Ne laser and the fluorescence were collected with a 570 nm to 620 nm and a 560IF band pass filters, respectively. Alexa 633 was excited with a 633 nm He-Ne laser and the fluorescence was collected with a 647 nm long-pass filter. Image size was between 512 x 512 and 1600 x 1600 pixels with a pixel resolution of 66–69 nm. Deconvolution was performed with Huygens Essential software (SVI, Netherlands). The point-spread function was averaged, and the iterative deconvolution was performed with a signal-to-noise ratio of 5, and the quality threshold was 0.01. Image analysis was done with ImageJ (30).

Correlation analysis was done with ImageJ using the JACoP plugin (31). Distance analysis of the fixed-cell samples was done by first making a Euclidian distance map from an image of interest and then plotting the original image against the distance map. The data from every cell were combined and arranged to 0.1 μm wide bins based on the distance of the voxels from the nuclear envelope. The average intensity was then plotted for each bin. Distance maps were done in ImageJ with the Exact Euclidian Distance Transform (3D) plugin, and the data analysis was done with an in-house Java code. Student's t-test (two-tailed, unequal variance) was used to evaluate statistical significance in the change of the recovery time point values.

Analysis of protein interactions. For *in-situ* proximity ligation assay (PLA) (32), cells were grown on 8-well chamber slides (Nunc™ Lab-Tek™ II Chamber Slide™ System, Nalgene Nunc International, Penfield, NY) to 80-90 % confluence and fixed

145 with 4% PFA. PLA was done with a Duolink®II kit (Olink Bioscience, Uppsala,
146 Sweden) and primary Abs against NS1 and H3K27ac. After labeling the cells with the
147 primary Abs, the PLA probes (oligonucleotide-conjugated secondary anti-mouse and
148 anti-rabbit IgGs diluted in 3 % BSA in PBS) were incubated for 1 h at 37 °C in a
149 humidified chamber followed by ligation and amplification according to the
150 manufacturer´s instructions. Samples were embedded in ProLong Gold antifade
151 mounting medium with 4,6-diamidino-2-phenylindole (DAPI). The specificity of the
152 assay was confirmed using positive, negative and technical control readings. Positive
153 and negative controls consisted of infected and non-infected cells, respectively, labeled
154 with anti-VP2 and intact capsids specific Abs, or anti-NS1 and anti-H3K27ac Abs. For
155 a technical control, non-infected cells were labeled only with PLA probes. Quantitative
156 analysis was done with ImageJ by determining the arithmetic mean of the total number
157 of signals per cell. The PLA signal was normalized as PLA=NA-NT, where NA is the
158 total number of nuclear PLA dots in infected and non-infected cells labeled with
159 H3K27ac Ab and NS1 Mab, and NT is the average number of nuclear PLA dots in the
160 technical control. Student's t-test (two-tailed, unequal variance) was used to evaluate
161 statistical significance.

162 **ChIP-seq and qPCR.** Cells were fixed with formaldehyde and nuclei were isolated,
163 lysed, and sonicated with a Covaris S220 ultrasonicator. The resulting nuclear extract
164 was incubated overnight at 4 °C with Dynal Protein G beads pre-incubated with 5µg of
165 H3K27ac (ab4729) or H3 (ab1791) Abs. Beads were washed and bound complexes
166 eluted, and cross-links were reversed by heating at 65 °C. IP and input DNA were then
167 purified by a treatment with RNAse A, proteinase K, and phenol-chloroform
168 extraction. Before moving forward to ChIP-seq, precipitated DNA was analyzed by
169 qPCR using primers specific for gp1, gp2 and gp5 regions. Libraries were constructed

from IP and input DNA by NEBnext® Ultra™ DNA-library preparation kit for Illumina. DNA in the range 150–350 bp was gel-purified after PCR amplification. The library was quantified using Agilent bioanalyzer and subjected to 50 bp single-end read sequencing with Illumina Hiseq 2000 at EMBL Genecore, Heidelberg. Quality metrics for sequenced reads were gathered with FastQC (33). Adapter sequences were removed with cutadapt (34). Reads were then aligned with Bowtie2 (35) to cat (ICGSC Felis_catus 6.2) and CPV (NCBI Reference Sequence NC_001539.1) reference genomes. Sequencing reads of H3K27ac ChIP-seq from infected cells were normalized to CPV reads per genomic content (RPGC). File conversions to the BAM format were done with SAMtools (36), and visualizations with deepTools (37).

RESULTS

Temporal changes in localization of modified histones in infection. To analyze chromatinization of CPV genome at various times post infection (p.i.), we used H3-EGFP-expressing cells to identify the distribution of histone H3 and APAR bodies represented by NS1. The line profiles showed that at 12 h p.i. H3 was mostly located distinctly from NS1, whereas at 24 h p.i. increased colocalization of H3 with NS1 was observed (Fig. 1A and 1B). Similarly, Pearson correlation coefficient (PCC) analysis indicated correlation of H3-EGFP with NS1 at 16 h p.i. (0.61 ± 0.29, $n=22$, student´s T-test p-values at 16/24 h p.i. in comparison to 10 h p.i., $p<0.05$) and 24 h p.i. (0.77 ± 0.21, $n=26$, $p<0.01$), but not at earlier times (Fig. 1G). To assess how H3-associated active and repressed chromatin is distributed in infected cells, we used H3K27ac and H3K9me3 as markers of active and repressed chromatin, respectively. Immunolabeling indicated that at 12 h p.i., distinct NS1 foci began to emerge, while H3K27ac and H3K9me3 showed a thorough nuclear distribution. The line profiles of H3K27ac and H3K9me3 with NS1 at 12 h p.i. indicated only a weak colocalization (Fig. 1C and 1E).

However, at 24 h p.i. H3K27ac concentrated in the enlarging APAR bodies (Fig. 1D). Similarly, PCC of NS1 with H3K27ac was significantly higher at 16 h p.i. (0.83 ± 0.22, n=18, $p<0.01$) and at 24 h p.i. (0.87 ± 0.08, n=23, $p<0.01$) in comparison to earlier times (Fig. 1G). No correlation of H3K9me3 with NS1 was observed by the line profile and PCC analyses (Fig. 1E, 1F and 1G,). Colocalization of NS1 with H3 and H3K27ac correlated time-dependently with enlargement of APAR bodies. Quantitative 3D distribution analysis of H3K27ac and H3K9me3 as a function of distance from the nuclear rim indicated that the H3K27ac signal was located in the nuclear center in both infected (24 h p.i.) and non-infected cells (n=20, 1000 spots/cell counted; Fig. 1H). In infected cells, the 0.7 µm-thick region at the nuclear periphery, in particular, was enriched in H3K9me3, and its intensity decreased toward the nuclear center. In non-infected cells, the H3K9me3 signal was distributed throughout the nucleus (n=20, 1000 spots/cell counted; Fig. 1I).

In summary, these data showed that in the infected cells both H3K27ac and H3-EGFP were enriched in APAR bodies. At the same time, H3K9me3 was found to accumulate in the nuclear periphery, an area known to harbor the layer of marginalized cellular chromatin. This suggests that the histones associated with the viral genome in the APAR area bear modifications that are characteristic of active gene expression.

Time-dependent intranuclear interplay of H3K27ac and NS1. In order to address the nuclear interaction of H3K27ac with viral DNA-bound NS1, *in situ* PLA was performed. PLA is an immunodetection technique that generates a fluorescent signal only when two antigens of interest are within 40 nm of each other (32). To assess the nuclear interactions of H3K27ac and NS1, we analyzed infected cells by PLA at 8–24 h p.i. Non-infected control cells exhibited only a faint nuclear signal (PLA signal per cell 1.1±0.19, n=111). At 8, 10, 12, and 16 h p.i. time-dependent increase in amount of

220 punctuate intranuclear PLA signals was detected (0.14±0.16, n=105; 0.81±0.30, n=92;
221 3.43±0.91, n=110; 19.46±3.39, n=90) with a maximal signal at 24 h p.i. (36.49±3.97,
222 n=99, Fig. 2A) in the low DAPI nuclear interior (Fig. 2B). The infection-induced
223 compaction of the host chromatin and its dislocation into the nuclear periphery and
224 around the nucleolus at 24 h p.i., were visualized with DAPI.

225 In summary, these results demonstrated a time-dependent increase in interaction or
226 close proximity of H3K27ac with NS1 after 10 h p.i. followed by extensive interaction
227 at 24 h p.i. This finding is consistent with the results of immunofluorescence and PCC
228 analyses of infected cells, which showed increased colocalization and correlation of
229 H3K27ac and NS1 at late-stage infection.

230 **CPV promoters are rich in acetylated histones.** The prominent colocalization and
231 interaction of NS1 with H3K27ac in the nucleus at 24 h p.i. prompted us to study
232 whether the CPV genome *per se* is chromatinized with H3K27ac-enriched
233 nucleosomes. To this end, ChIP-seq with H3K27ac Ab for the infected cells was
234 performed. First, the success of ChIP was assessed with genome-wide occupancy of
235 H3K27ac in infected cells showing a typical occupancy of this histone marker at ±2 kb
236 around the transcription start sites (TSSs) of ~35 % of the genes (data not shown). We
237 found that 9.23 % of the total reads ($2.23x10^7$ reads) were aligned with the viral
238 genome in a unique manner (0.00% in non-infected cells) (38). Alignment of these
239 reads with the genome, after normalization to reads per CPV-genomic content,
240 revealed that H3K27ac is mostly enriched in the TSSs of P4 promoter driven
241 transcriptional units for NS1 and NS2 (gp1 loci) and P38 driven transcriptional units
242 for VP1 and VP2 (gp2 loci) (Fig. 3A) (39).

The viral genome was mostly devoid of this histone marker towards the right-hand end (gp5 loci). The location of gp1 loci was very similar to that of P4 promoter of the parvovirus minute virus of mice (MVM, nucleotides 1–260) and the location of gp2 was similar to that of CPV P38 promoter (1355–2260) (40). The targeted ChIP-qPCR of H3 with gp1, gp2 and gp5-specific primers showed that the CPV genome is thoroughly chromatinized with uniformingly distributed H3 (Fig. 3B, data not shown). Finally, targeted ChIP-qPCR analysis confirmed occupancy of H3K27ac in the genome (Fig. 3C).

In conclusion, our data verified that the parvoviral genome is chromatinized with histones at late-stage infection. Importantly, acetylated histones were observed to be enriched in viral promoters to allow for transcriptional activation of the viral NS and VP genes.

Viral histone acetylation is important for infection progress. In order to determine the importance of histone acetylation of parvoviral genomes during early stages of infection, the effect of histone HAT inhibitor anacardic acid on viral protein synthesis was determined. Treatment with anacardic acid at very early infection (-0.5–1 h p.i.) resulted in decrease in the percentage of cells with nuclear NS1 (~5 %) and VP2 (~2 %) at 24 h p.i. in comparison to non-treated infected cells (68.0 and 50.3 % respectively, $n \geq 250$). This suggests that early inhibition of histone acetylation induced an almost complete block of infection. While the early treatment exerted a maximal effect, anacardic acid was able to inhibit the infection progress when introduced later, up to three hours post infection (Fig. 4A). Here, we cannot rule out the possibility that histone acetylation was not the only factor affecting viral protein production. Also cell cycle progression could be involved. Of note, the treatment did not affect cell viability (data not shown). Our results suggest that histone acetylation is required at very early

infection for the production of viral proteins necessary for efficient progress of infection.

Next, we studied if the anacardic acid-induced inhibition of histone acetylation affected late infection. For this we divided the infected cells in four categories based on intracellular localization of capsids at 24 h p.i. The categories were: (I) capsids localized into cytoplasmic endocytic vesicles due to inhibition of infection or secondary infection (24); (II) capsids localized in discrete foci scattered throughout the nucleus; (III) capsids in enlarged APAR bodies enriched with NS1; (IV) capsids both in APAR bodies and in the cytoplasm (egress initiated) (Fig. 4B). Next, the localization of capsids was determined at 24 h p.i. in cells exposed to anacardic acid at 6 h p.i. Studies indicated that category II, III and IV cells were reduced in treated cells (4.9 %, 12.5 %, 21.2 %; n≥250) in comparison with that of the non-treated infected control cells (7.9 %, 26.8 %, 26.8 %; n≥250). Moreover, an increase in category I cells was detected in treated cells (61.1 %; n≥250) in comparison to control cells (36.6 %; 1.9 %, n≥250; Fig. 4B). These results demonstrated that efficient progress of infection correlates with histone acetylation in APAR area at late stages of infection.

Finally, we studied whether the timing of viral histone acetylation was critical for progress of late-stage infection. Here, the distribution of H3K27ac and NS1, positioned on viral genome, was analyzed at 24 h p.i. in cells treated with anacardic acid. The inhibitor treatment at 16 h p.i. resulted in exclusion of H3K27ac from the NS1-positive APAR bodies (Fig. 4C). This indicated that the viral genomes produced after drug treatment at 16 h p.i. were not acetylated. However, H3K27ac colocalized with NS1 after exposure to inhibitor at 20 h p.i. (Fig. 4C). These results suggested that histone acetylation on majority of newly synthesized viral DNA occurs until 16 - 20 h p.i.

These inhibition studies provided evidence that histone acetylation of intranuclear viral genomes at early infection plays an essential role in production of viral proteins. Similarly, acetylation of histones on newly synthesized viral genomes during active virus replication is necessary for efficient accomplishment of infection. In summary, our findings demonstrate that viral histone acetylation is essential for both early and late steps in parvoviral life cycle.

DISCUSSION

Although much is known about nuclear replication and gene expression of parvoviruses, little is known about chromatinization of parvoviral genome and histone modifications and effect of these events on progress of infection. For some DNA viruses, such as herpesviruses and adenoviruses, epigenetic mechanisms, including histone modifications play an important role in the regulation of viral gene expression. During herpesvirus lytic infection, the viral genomes are associated with histones immediately after injection into the nucleus, and viral proteins are required to enhance histone acetylation to allow for efficient viral gene expression (41-45). Moreover, studies of infection by a adenovirus have shown that viral proteins mediate transcriptional activation of viral promoter regions (46-48). In this work, we first observed that the progress of CPV infection was accompanied by enrichment of H3 histones in the enlarged APAR body area. This is consistent with earlier studies showing nuclear chromatinization of adeno-associated parvoviruses, used as gene therapy vectors, and MVM genomes (49-52). Next, we addressed the existence of histone modifications in the parvoviral APAR bodies and revealed accumulation of histones with modifications characteristic of transcriptionally active chromatin (H3K27ac). Moreover, our analysis demonstrated that H3K27ac was located in close proximity to the viral NS1 protein in the APAR bodies. This interaction is likely to be

caused by accumulation of NS1 on viral genome because of its involvement in transcription and replication (53-55). NS1 of autonomous parvoviruses not only controls the viral activities but also regulates the host-gene expression through histone acetylation in cancer cells by recruiting innate proteins with HAT-activity (56,57). To date, evidence for involvement of NS1 in regulation of histone acetylation on parvoviral promoters has not been reported. Here, our PLA studies revealed that CPV NS1 was located in close proximity to H3K27ac in the enlarged APAR bodies. Our earlier studies indicated that CPV NS1 has two distinct binding sites in the viral genome (54,55,58). It is tempting to speculate that NS1 is involved in histone acetylation of viral P4 and P38 promoters by recruiting host proteins with acetyl-transferase activity. However, the specific role and interactions of NS1 in viral histone modifications remains to be determined. Our ChIP-seq analyses demonstrated for the first time that in the extensively chromatinized parvoviral genome H3K27ac accumulated in viral P4 and P38 promoter areas. This suggests that parvoviral gene expression is regulated by histone acetylation in promoter areas. The importance of histone acetylation for the progress of infection was supported by our results showing that inhibition of histone acetylation was accompanied by extensive repression of infection. Earlier studies have shown that formation of parvoviral gene transcription templates and intermediates for genome replication are temporally phased in infection (59-62). In line with these, our results suggest that parvoviruses regulate their gene expression via histone acetylation in a temporal fashion.

In summary, CPV genome is chromatinized inside the nuclei of infected cells and histone modifications associated with transcriptional activation are enriched in viral promoters. Our results highlight a critical role of epigenetic modification in progression of the parvoviral life cycle.

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FIGURE LEGENDS

Figure 1

Intranuclear distribution of H3-EGFP, modified histones and viral NS1 protein. Confocal microscopy images of cells stably expressing H3-EGFP (cyan) (A) at 12 h and (B) at 24 h pi labeled with NS1 (yellow) antibody. Infected cells at 12 h p.i. and 24 h p.i. labeled with antibodies for H3K27ac (cyan) (C, D), H3K9me3 (cyan) (E, F) and NS1 (yellow). In order to clarify the changes in colocalization, pseudo-colour images with intensity increasing from blue to yellow are shown. Fluorescence line-profile analysis of the intensity of H3-EGFP/H3K27ac/H3K9me3 (cyan) and NS1 (yellow) in a single optical section through the center of nucleus is shown beside each image. Analysis was performed with ImageJ and Plot RGB Profile-plugin. Scale bars, 5 μm. (G) Quantitative colocalization analysis of H3K27ac (dove grey), and H3K9me3 (dark grey) and H3-EGFP (black) with NS1. The mean values of PCCs with standard deviation (SD) are shown. Statistical significance of colocalization at 16 and/or 24 h p.i. in comparison to 10 h p.i. is shown (Student´s T-test p-values: *P<0.05; **$P<$ 0.01). Plots of intensity of H3K27ac (H) and H3K9me3 (I) in infected cells at 24 h p.i. (dark grey) and in non-infected (dove grey) as a function of distance from the nuclear envelope.

Figure 2

***In situ* PLA of H3K27ac interaction with NS1 in infected cells.** (A) Number of PLA signals in infected cells at 8, 10, 12, 16 and 24 h p.i. The mean values of PLA signals with ±standard error from ~100 cells per time point are shown. The negative control, PLA signal in non-infected cells, is shown by a dashed line. Statistical significance of PLA signals per cell at 12, 16 and 24 h p.i. in comparison to negative control is shown

(Student´s T-test p-values: *P<0.05;**P< 0.01). (B) Distribution of PLA signals in DAPI-stained nucleus at 24 h p.i. Scale bar, 10 μm.

Figure 3

Enrichment of acetylated histones on gp1 and gp2 transcription start sites and effect on gene expression. (A) Histogram depicts occupancy of H3K27ac at CPV genome. Sequencing reads of H3K27ac ChIP-seq from infected cells at 24 h p.i. are normalized to RPGC. Bent arrows indicate TSSs of transcriptional units for NS1 and NS2 (gp1), VP1 and VP2 (gp2) and right-hand terminal gp5 loci and red bars depict regions where qPCR primers anneal. ChIP-qPCRs for targeted measurements of (B) H3 and (C) H3K27ac level at gp1 and gp2 TSSs and VP2-gp5 inter-region is shown as enrichment of DNA relative to input DNA. Two primer pairs for each of the three regions were used; columns represent the means and ±SD of triplicate wells of two independent ChIP experiments.

Figure 4

Importance of the acetylation on progress of infection. (A) Percentage of cells showing viral NS1 and VP proteins in the nucleus in the cells exposed to HAT inhibitor, anacardic acid (0.1 mM). Cells were either pretreated for 30 min with the drugs prior to infection or treated at 0-6 h pi, and the treatment continued until fixation at 24 h p.i. (B) Effect of anacardic acid treatment on localization of viral capsids. Images of cells representing four type categories based on intracellular localization of capsids at 24 h p.i. and percentages of cell showing various types after treatments at 6

583 or 16 h p.i. The mean values with ±SD are shown. (C) Distribution of H3K27ac in
584 comparison to NS1 at 24 h p.i. in cells treated with anacardic acid at 16 or 20 h p.i.
585 Scale bars, 10 μm.